Shelley et al., http://www.jgp.org/cgi/content/full/jgp.201311072/DC1

## Supplemental results

To examine the consequence of each sequence alteration required to generate the SCN variants $\mathrm{BK}_{0}$ and $\mathrm{BK}_{\text {SRKR }}$ from the parent construct, mbr5, we constructed a series of clones that differed by stepwise addition and deletion of the indicated sequences (Fig. S1 A). We found that the addition of the "MANG" N-terminal sequence found in the SCN clones (C2), removal of IYF at splice site 2 (C3), and switching the "VEDEC" C-terminal sequence to "VYR" (C4) had no significant effects on the G-V relationships or kinetics of activation or deactivation (Fig. S1, B-D). However, comparing mbr5 currents to $\mathrm{BK}_{0}$, the addition of the $\mathrm{Ca}^{2+}$-bowl insert at splice site 3 right-shifted the G-V curve at 0,1 , and 100 $\mu \mathrm{M} \mathrm{Ca}^{2+}$ (Fig. S2 A and Table S1). There was no effect of adding the $\mathrm{Ca}^{2+}$-bowl exon at $10 \mu \mathrm{M} \mathrm{Ca}^{2+}$. The addition of the $\mathrm{Ca}^{2+}$-bowl exon also influenced the kinetics of activation and deactivation. At 10 and $100 \mu \mathrm{M} \mathrm{Ca}^{2+}, \mathrm{BK}_{0}$ had slowed activation compared with mbr5 (Fig. S2 B). $\mathrm{BK}_{0}$ deactivation was faster at $0 \mathrm{Ca}^{2+}$, but slower at 100 $\mu \mathrm{M} \mathrm{Ca}^{2+}$, than mbr5, with no change seen at 1 and 10 $\mu \mathrm{M} \mathrm{Ca}^{2+}$ (Fig. S2 C). Thus, among the sequences tested here, the addition of the $\mathrm{Ca}^{2+}$-bowl exon is the first alteration to produce significant changes in BK current properties. Notably, the changes in G-V and kinetics reported in this study for the addition of the $\mathrm{Ca}^{2+}$-bowl exon are larger than reported previously (Ha et al. 2000. Eur. J. Biochem. 267:910-918). However, the back-
ground variants used in the respective studies were also different.

The addition of the SRKR exon at site $1\left(\mathrm{BK}_{\text {SRKR }}\right)$ produced a further right-shift of the G-V at 0,1 , and 100 $\mu \mathrm{M}$, but not $10 \mu \mathrm{M} \mathrm{Ca}^{2+}$ (Fig. S2 A and Table S1). To determine whether the addition of the $\mathrm{Ca}^{2+}$ bowl at site 3 or the SRKR exon at site 1 had a larger impact on BK current properties, the magnitudes of the $\mathrm{V}_{1 / 2}$ shifts were compared ( $\Delta \mathrm{V}_{1 / 2}$ for $\mathrm{C} 4-\mathrm{BK}_{0}$ vs. $\mathrm{BK}_{0}-\mathrm{BK}_{\text {SRKR }}$ ). At 1 and $100 \mu \mathrm{M} \mathrm{Ca}^{2+}$, the $\Delta \mathrm{V}_{1 / 2}$ values produced by the addition of the $\mathrm{Ca}^{2+}$ bowl and SRKR exons were similar, being 29 and 33 mV at $1 \mu \mathrm{M} \mathrm{Ca}^{2+}$, and both 3 mV at 10 $\mu \mathrm{M} \mathrm{Ca}{ }^{2+}$ (Table S1). However, the addition of SRKR produced a larger $\Delta \mathrm{V}_{1 / 2}$ at $0 \mathrm{Ca}^{2+} \mu \mathrm{M}(49 \mathrm{mV}$ compared with 34 mV for adding $\mathrm{Ca}^{2+}$ bowl). Overall, the addition of SRKR had the largest impact on BK current properties of the sequences tested in this study and produced the most right-shifted G-V curves (see also Results in the main text).

A mbr5 MDAL... S1-S6 $\quad$ RCK1 | LIYF | RCK2 | ...VEDEC |
| :--- | :--- | :--- | :--- |

| C2 | MANG... | S1-S6 | RCK1 | LIYF | RCK2 | VEDEC |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| C3 | MANG... | S1-S6 | RCK1 | L | RCK2 | .VEDEC |

C4 | MANG... | S1-S6 | RCK1 | L | RCK2 | $\ldots$ RKEMVYR |
| :--- | :--- | :--- | :--- | :--- | :--- |

$\mathrm{BK}_{0}$| MANG... | S1-S6 | RCK1 | L | RCK2 [AKP...TEL] |
| :--- | :--- | :--- | :--- | :--- |
|  | ...RKEMVYR |  |  |  |














Figure S1. Properties of BK currents from clones mbr5, C2, C3, and C4. (A) Schematic diagram of mbr5, $\mathrm{BK}_{0}$, $\mathrm{BK}_{\text {SRKR }}$, and the intermediate clones used to examine the functional consequences of stepwise additions or deletions of particular inserts. (B) G-V relationship for constructs mbr5, C2, C3, and C4, at the indicated $\mathrm{Ca}^{2+}$ concentrations. The alternative N terminus, deletion of IYF, and the alternative C terminus had no effect on the G-V curves. $\mathrm{V}_{1 / 2}$ values are given in Table S1. (C and D) Plot of $\tau_{\text {act }}$ or $\tau_{\text {deact }}$ versus voltage for constructs at the indicated $\mathrm{Ca}^{2+}$; as with the G-V curves, no differences in the time constants of activation or deactivation were seen for any of the constructs.


Figure S2. Properties of BK currents from mbr5, $\mathrm{BK}_{0}$, and $\mathrm{BK}_{\text {SRKR. }}$. (A) G-V relationship for mbr5, $\mathrm{BK}_{0}$, and $\mathrm{BK}_{\text {SRKR. }}$. The addition of the $\mathrm{Ca}^{2+}$-bowl exon $\left(\mathrm{BK}_{0}\right)$ and $\operatorname{SRKR}\left(\mathrm{BK}_{\text {SRKR }}\right)$ right-shifts the G-V relationship at all $\mathrm{Ca}^{2+}$ except for $10 \mu \mathrm{M} \mathrm{Ca}{ }^{2+}$, where no shift is observed. The magnitude of the shift was similar between $\mathrm{BK}_{0}$ and mbr5, and between $\mathrm{BK}_{\text {SRKR }}$ and $\mathrm{BK}_{0}$ at 1 and $100 \mu \mathrm{M} \mathrm{Ca}{ }^{2+}$, but was larger for $\mathrm{BK}_{\text {SRKR }}$ at $0 \mathrm{Ca}^{2+}$. $V_{1 / 2}$ values are given in Table S1. ( B and C ) Plot of $\tau_{\text {act }}$ or $\tau_{\text {deact }}$ versus voltage for constructs at the indicated $\mathrm{Ca}^{2+}$. $\mathrm{BK}_{\text {SRKR }}$ is slower to activate than mbr5 at all $\mathrm{Ca}^{2+}$ concentrations, whereas $\mathrm{BK}_{0}$ only activates slower than mbr5 at 10 and $100 \mu \mathrm{M} \mathrm{Ca}$. At $0 \mathrm{Ca}^{2+}$, mbr5 deactivates slower than $\mathrm{BK}_{0}$ and $\mathrm{BK}_{\text {SRKR }}$; however, as $\mathrm{Ca}^{2+}$ is increased, the rate of deactivation of mbr5 increases compared with $\mathrm{BK}_{0}$ and $\mathrm{BK}_{\text {SRKR }}$.


Figure S3. BK currents are far from maximally activated in response to APs. (A) The complete day voltage protocol used to activate channels, consisting of a maximally activating voltage jump to +160 mV , followed by a holding potential identical to the mean daytime neuronal resting membrane potential, and the day AP waveform. (B) Example $\mathrm{BK}_{0}$ current in response to A , clearly showing that the AP does not maximally activate the channel. (C) Overlay of normalized currents from $\mathrm{BK}_{0}$ (black) and $\mathrm{BK}_{\text {SRKR }}$ (red) channels showing the reduced AP current and slower activation of $\mathrm{BK}_{\text {SRKR }}$ currents compared with $\mathrm{BK}_{0}$ currents in response to the day AP command waveform.


Figure S4. Structural model for the alternate splice site 1 region of $\mathrm{BK}_{0}$ and $\mathrm{BK}_{\text {SRKR }}$ variants. The SRKR insert shifts S 642 from a $\beta$-strand region into an extended loop region in the BK channel. Comparison between the cytosolic domains of a human BK channel crystal structure that lacks the SRKR insert (Protein Data Bank accession no. 3NAF; Wu et al. 2010. Nature. 466:393-397) and a zebrafish structure that contains the SRKR insert (Protein Data Bank accession no. 3U6N; Yuan et al. 2012. Nature. 481:94-97) reveals that the SRKR insert enlarges the loop region between two $\beta$ strands. Arrows denote residues within $\beta$ strands. Atomic coordinates of residues marked in red were not determined.

Table S1
$V_{1 / 2}$ values for $B K$ variants

| BK variant | $\mathrm{V}_{1 / 2}(\mathrm{mV})$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | $0 \mu \mathrm{M} \mathrm{Ca}{ }^{2+}$ | $1 \mu \mathrm{M}$ | $10 \mu \mathrm{M}$ | $100 \mu \mathrm{M}$ |
| mbr5 | $173 \pm 2$ | $138 \pm 5$ | $58 \pm 3$ | $-31 \pm 2$ |
| C2 | $174 \pm 3$ | $144 \pm 3$ | $58 \pm 6$ | $-34 \pm 2$ |
| C3 | $167 \pm 5$ | $144 \pm 5$ | $54 \pm 3$ | $-30 \pm 2$ |
| C4 | $166 \pm 6$ | $142 \pm 7$ | $55 \pm 2$ | $-27 \pm 2$ |
| $\mathrm{BK}_{0}$ | $200 \pm 11$ | $171 \pm 4$ | $58 \pm 3$ | $-11 \pm 2$ |
| $\mathrm{BK}_{\text {SRKR }}$ | $249 \pm 3$ | $204 \pm 4$ | $61 \pm 2$ | $8 \pm 2$ |
| $\mathrm{BK}_{0}+$ Alk P |  | $166 \pm 2$ | $59 \pm 4$ | $-8 \pm 3$ |
| $\mathrm{BK}_{\text {SRKR }}+$ Alk P |  | $166 \pm 6$ | $62 \pm 2$ | $-7 \pm 3$ |
| $\mathrm{BK}_{\text {SRKR }}$-S644A |  | $206 \pm 6$ | $62 \pm 3$ | $4 \pm 3$ |
| $\mathrm{BK}_{\text {SRKR }}$-S642A |  | $172 \pm 6$ | $54 \pm 4$ | $-11 \pm 3$ |
| $\mathrm{BK}_{0}$-S642D |  | $208 \pm 3$ | $59 \pm 2$ | $11 \pm 6$ |
| $\mathrm{BK}_{\text {SRKR }}$-S642A/S644A |  | $167 \pm 4$ | $62 \pm 2$ | $-15 \pm 2$ |
| $\mathrm{BK}_{0}+\beta 4$ |  |  | $92 \pm 5$ | $-23 \pm 3$ |
| $\mathrm{BK}_{\text {SRKR }}+\beta 4$ |  |  | $151 \pm 12$ | $15 \pm 8$ |

G-V relationships were constructed from voltage-clamp recordings as described in Materials and methods in the main text. $\mathrm{V}_{1 / 2}$ values at the indicated $\left[\mathrm{Ca}^{2+}\right]_{i}$ were determined from Boltzmann fits to G-V data obtained from inside-out patches using standard square voltage pulses to activate currents in symmetrical $\mathrm{K}^{+}$conditions ( $n \geq 8$ for each condition). Alk P, patches treated with alkaline phosphatase.

