Supplemental material

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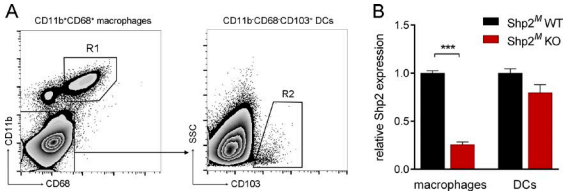

Figure S1. **Efficiency of Cre-driven Shp2 deletion in colonic macrophages and DCs. (A and B)** Colonic macrophages ($CD11b^+CD68^+$; R1) and DCs ($CD11b^-CD68^-CD103^+$; R2) were enriched by fluorescence-activated cell sorting (A) and the expression Shp2 was evaluated by qPCR (B). Data are mean ± SEM and are compiled from two independent experiments. ***, P < 0.001; two-tailed unpaired Student's *t* test.

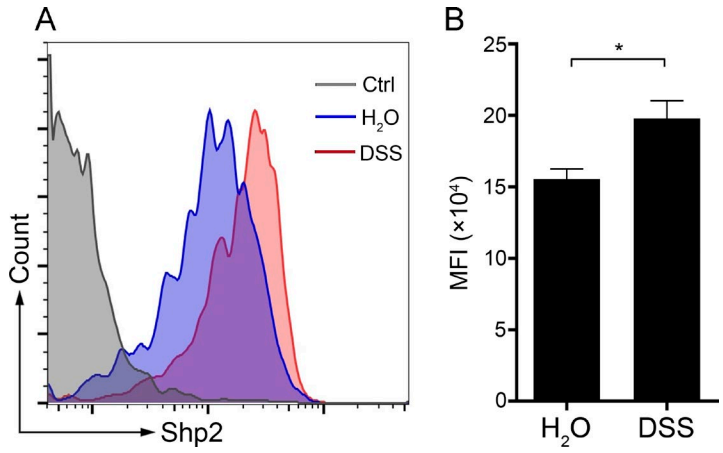

Figure S2. **Expression level of Shp2 in colonic macrophages.** The development of colitis increases Shp2 expression in colonic macrophages. Mice were fed with 2.5% DSS in drinking water for 7 d; the expression of Shp2 in colonic $CD68^+$ macrophages was evaluated by flow cytometry. **(A and B)** Representative plot (A) and quantitative analysis are shown (B). *n* = 4–5. Data are mean ± SEM and are representative of two independent experiments. *, P < 0.05; two-tailed unpaired Student's *t* test.

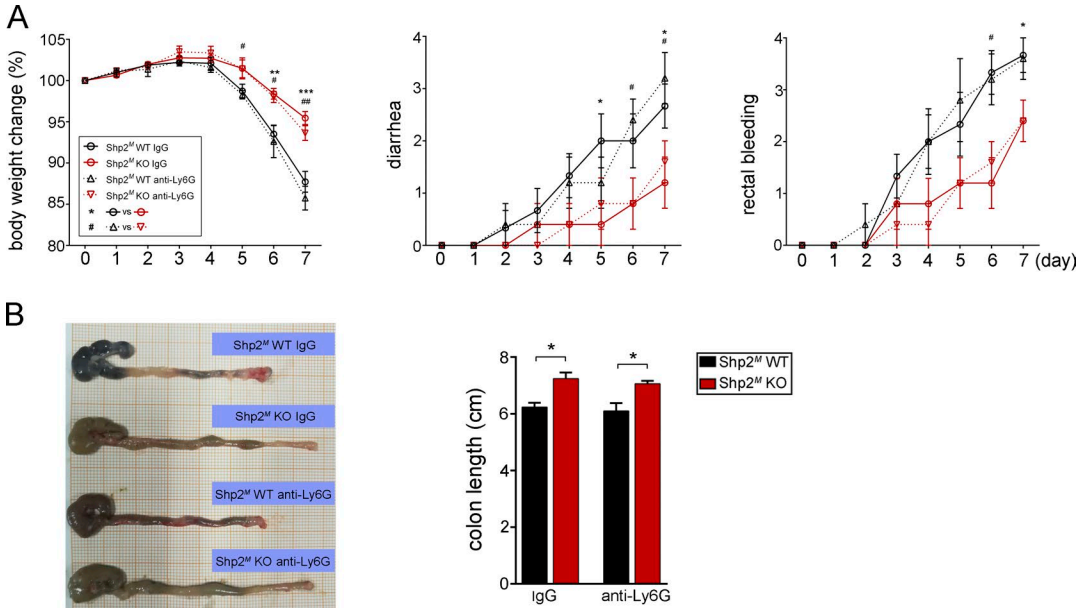

Figure S3. **Development of colitis in neutrophil-depleted mice.** Myeloid Shp2 deficiency alleviates colitis in neutrophil-depleted mice. Mice were injected intraperitoneally with 100 µg anti-Ly6G or IgG isotype at days −2, 0, 2, 4, and 6 after DSS challenge, respectively, to deplete neutrophils. **(A)** Body weight change, diarrhea, and rectal bleeding were monitored daily. **(B)** Colon length was measured at day 7. *n* = 5–6. Data are mean ± SEM and are representative of two independent experiments. * and #, P < 0.05; ** and ##, P < 0.01; ***, P < 0.001; two-tailed unpaired Student's *t* test.

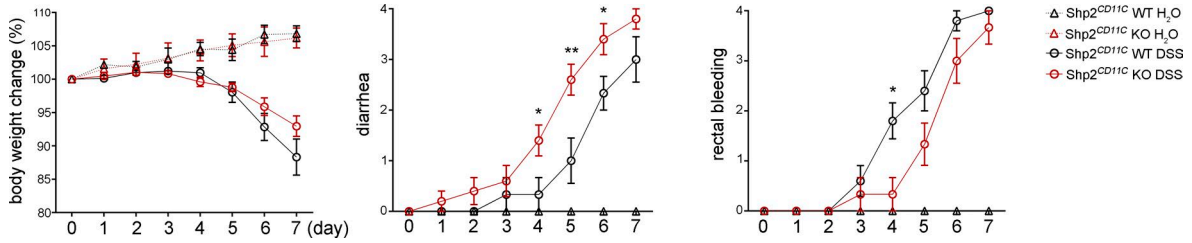

Figure S4. **Severity of colitis between Shp2^{CD11C} WT and Shp2^{CD11C} KO mice Shp2^{CD11C}.** KO mice have decreased colitis susceptibility. Mice were fed with 2.5% DSS for 7 d. Body weight change, diarrhea, and rectal bleeding were monitored daily. *n* = 6–10. Data are mean ± SEM and are representative of two independent experiments. *, P < 0.05; **, P < 0.01, two-tailed unpaired Student's *t* test.

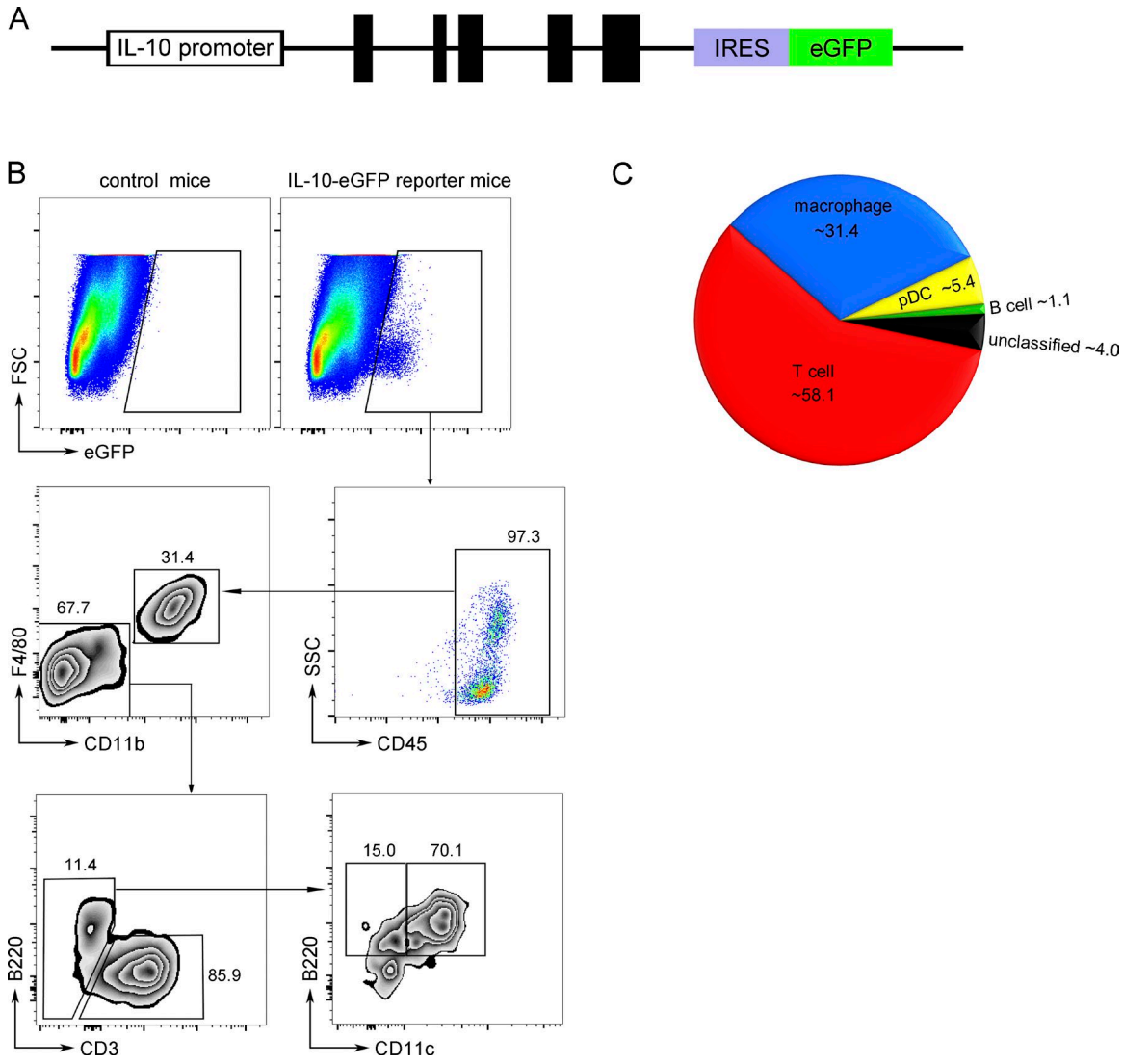

Figure S5. **Cellular sources of colonic IL-10T cells and macrophages are predominant sources of colonic IL-10. (A)** IL-10-GFP mice or control mice were fed with 2.5% DSS; the composition of GFP$^+$ cells were analyzed by flow cytometry. **(B and C)** Representative gating strategy (B) and quantitative analysis (C) are shown. Numbers represent cell percentages. IRES, internal ribosome entry site. Data are representative of two independent experiments.

Table S1. **Primers for qPCR analysis**

Gene	Direction	Primer (5′–3′)
β-Actin	F (forward):	GGCTGTATTCCCCTCCATCG
	R (reverse):	CCAGTTGGTAACAATGCCATGT
TNF-α	F:	CCTGTAGCCCACGTCGTAG
	R:	GGGAGTAGACAAGGTACAACCC
IL-6	F:	CTGCAAGAGACTTCCATCCAG
	R:	AGTGGTATAGACAGGTCTGTTGG
IL-1β	F:	GAAATGCCACCTTTTGACAGTG
	R:	TGGATGCTCTCATCAGGACAG
IL-12	F:	TGGTTTGCCATCGTTTTGCTG
	R:	ACAGGTGAGGTTCACTGTTTCT
IFN-γ	F:	ATGAACGCTACACACTGCATC
	R:	CCATCCTTTTGCCAGTTCCTC
IL-23	F:	ATGCTGGATTGCAGAGCAGTA
	R:	ACGGGGCACATTATTTTTAGTCT
IL-17A	F:	TTTAACTCCCTTGGCGCAAAA
	R:	CTTTCCCTCCGCATTGACAC
IL-17F	F:	TGCTACTGTTGATGTTGGGAC
	R:	AATGCCCTGGTTTTGGTTGAA
16S rRNA	F:	GTGGTGCATGGTTGTCGTCA
	R	ACGTCGTCCCCACCTTCCTC
Human β-actin	F:	CATGTACGTTGCTATCCAGGC
	R:	CTCCTTAATGTCACGCACGAT
Human TNF-α	F:	CCTCTCTCTAATCAGCCCTCTG
	R:	GAGGACCTGGGAGTAGATGAG
Human IL-6	F:	ACTCACCTCTTCAGAACGAATTG
	R:	CCATCTTTGGAAGGTTCAGGTTG