

Supplemental material

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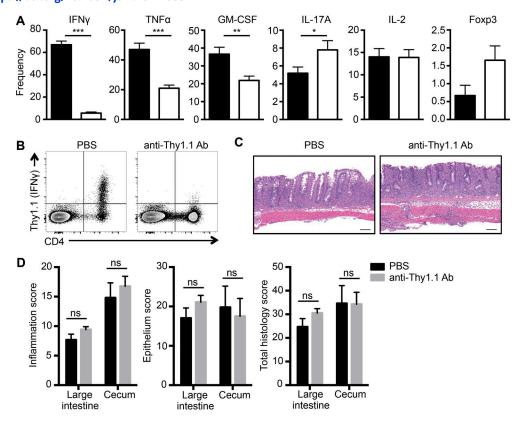


Figure S1. IFNy-producing CD4 T cells produce proinflammatory effector cytokines during IBD but are dispensable to induce intestinal inflammation. (A) Naive CD4 T cells from IFNy KI mice were transferred to Rag1^{-/-} recipient mice. After 6–8 wk, CD4 T cells from large intestine were segregated by expression of Thy1.1, the IFNy reporter molecule, and staining of indicated molecules was assessed by flow cytometry (n = 15-19, four to five experiments). (B–D) After transfer of IFNy KI naive CD4 T cells, recipient mice were treated with anti-Thy1.1 depleting antibody (clone 19e1.2) or PBS for 6–8 wk. (B) Thy1.1 expression on CD4 T cells in spleen after treatment. (C) Representative hematoxylin and eosin–stained sections from the large intestine of treated mice. The images were made with a $10 \times$ objective. Bars, 135μ m. (D) Disease scores from treated mice (n = 8-10, two experiments). Graphs show the mean \pm SD; unpaired t test: *, P < 0.05; **, P < 0.01; ***, P < 0.001.



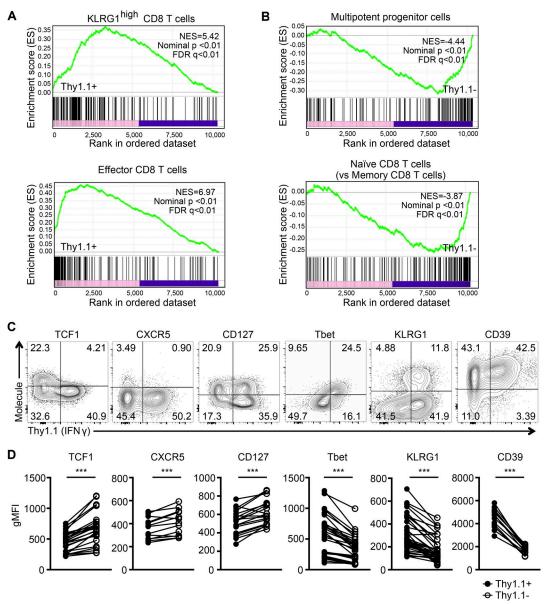


Figure S2. IFNy-producing CD4 T cells express a terminally differentiated signature, whereas IFNy-nonproducing CD4 T cells possess a self-renewing, progenitor-like profile. (A and B) GSEA plots illustrate enrichment of Thy1.1⁺ and Thy1.1⁻ CD4 T cell RNA sequencing data with previously published gene sets. (A) Representative GSEA of effector CD8 T cell gene sets (GSE10239, top; dataset from Kaech et al., 2002, bottom). (B) Representative GSEA of naive CD8 T cell gene sets in comparison to memory CD8 T cell gene signature (GSE37301, top; dataset from Luckey et al., 2006, bottom). (C and D) Expression of indicated molecules in the splenic CD4 T cells was determined by flow cytometry at 4–7 wk after colitis induction. The geometric mean fluorescent intensity for Thy1.1⁺ and Thy1.1⁻ CD4 T cells is shown. (n = 14–35, two to five independent experiments). Paired t test: ***, P < 0.001.

References

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