## SUPPLEMENTAL MATERIAL

## Kasheta et al., https://doi.org/10.1084/jem.20162084

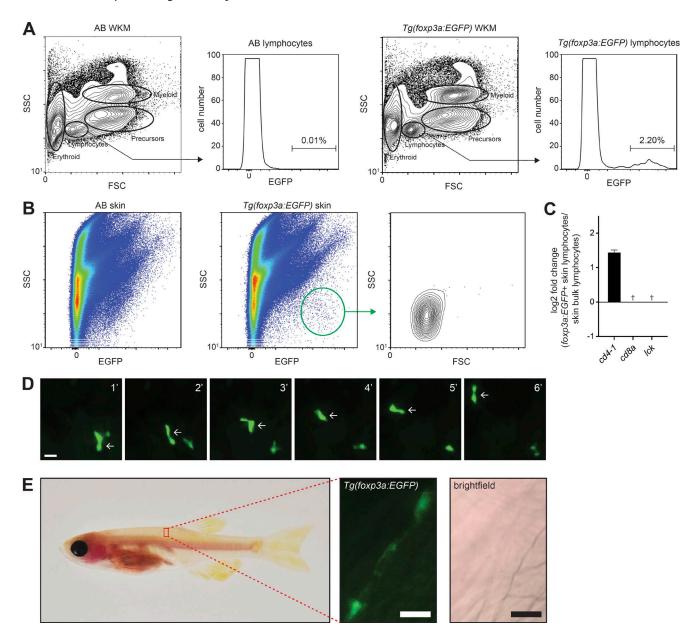


Figure S1. **Analysis of** foxp3a:EGFP-positive cells in multiple tissues. (A) Gating strategy for quantification of EGFP-positive lymphocytes from WKM. Related to Fig. 1 D. (B) Flow cytometry analysis of skin from representative AB and Tg(foxp3a:EGFP) animals. (C) qRT-PCR of selected genes in foxp3a:EGFP-positive thymocytes relative to bulk thymocytes. +, Expression of cd8a and lck was below the limit of detection in foxp3a:EGFP-positive skin lymphocytes. Error bar indicates SEM; n=3. (D) Still frames from live imaging (Video 1) of a foxp3a:EGFP-positive cell migrating through skin (arrows). Each panel represents a 1-min interval. Bar,  $10 \mu m$ . (E) foxp3a:EGFP-positive cells between segment of dorsal musculature. Bars,  $50 \mu m$ .

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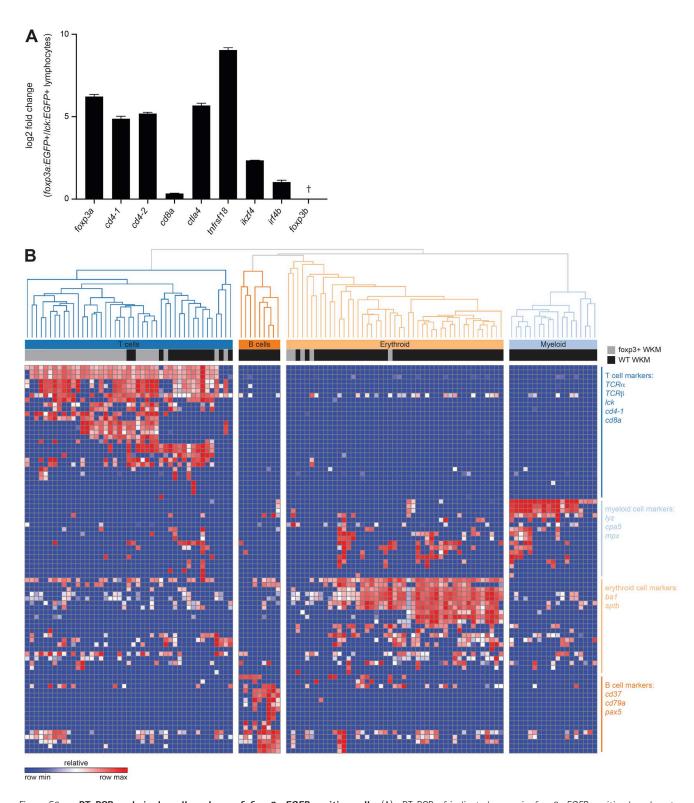


Figure S2. **qRT-PCR and single-cell analyses of** *foxp3a:EGFP*-**positive cells.** (A) qRT-PCR of indicated genes in *foxp3a:EGFP*-positive lymphocytes compared with lymphocytes sorted from Tg(lck:EGFP) animals. †, Expression of *foxp3b* was below the limit of detection in Tg(foxp3a:EGFP) animals. Error bar indicates SEM; n = 3. (B) Unsupervised hierarchical clustering using gene expression analyses of single cells. Hematopoietic lineage assignments of control WKM cells in each cluster are shown. These assignments were made based on expression of lineage-specific marker genes as well as a comparison of these expression profiles to those of cells isolated from animals with lineage-specific transgene expression (Moore et al., 2016a). Hematopoietic lineage-specific genes representing each lineage profiled are indicated (right).

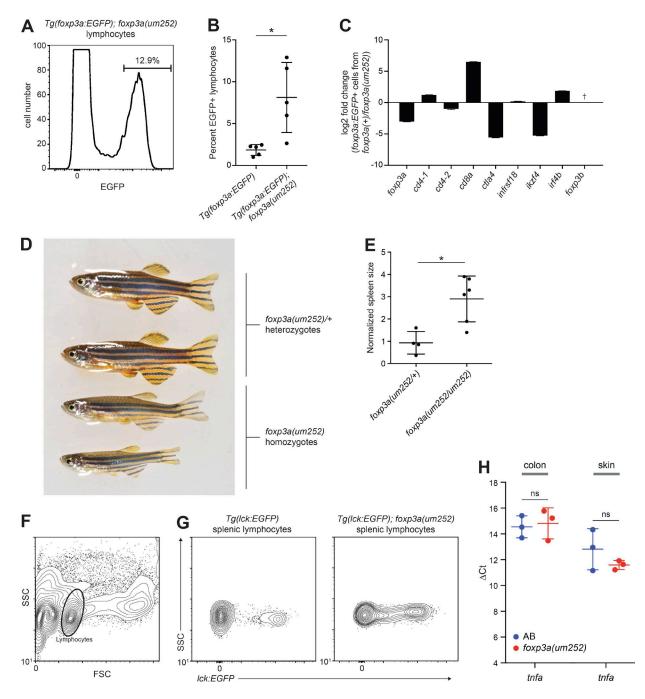


Figure S3. **Phenotypic analyses of** *foxp3a* **mutants.** (A) Flow cytometry analysis of WKM from a representative Tg(foxp3a:EGFP); foxp3a(um252) animal. Gated lymphocytes are plotted as a histogram with the percentage of EGFP-positive lymphocytes indicated. (B) Percentages of foxp3a:EGFP-positive lymphocytes, based on flow cytometry analysis, are increased in a foxp3a(um252) mutant (n = 5) compared with a wild-type (n = 5) background. Two-tailed Student's t test, \*, P < 0.05. Error bar indicates SEM. (C) qRT-PCR of indicated genes in EGFP-positive lymphocytes sorted from Tg(foxp3a:EGFP); foxp3a(um252) animals compared with EGFP-positive lymphocytes from Tg(foxp3a:EGFP) animals is plotted. †, Expression of foxp3b was below the limit of detection in EGFP-positive lymphocytes from both Tg(foxp3a:EGFP); foxp3a(um252) and Tg(foxp3a:EGFP) animals. Error bar indicates SEM; n = 3. (D) Images of representative sibling foxp3a(um252)/+ heterozygotes and foxp3a(um252) homozygotes. (E) Normalized spleen sizes of sibling foxp3a(um252)/+ heterozygotes and foxp3a(um252) homozygotes. (E) Normalizing maximum spleen area (area at level of splenic artery) by fish volume. Two-tailed Student's t test, \*, P = 0.007. (F) Flow cytometry analysis of a representative Tg(lck:EGFP) spleen. The lymphocyte-containing gate is indicated. (G) Gated lymphocytes from representative animals are plotted to indicate EGFP positivity. (H) qRT-PCR analysis of inflammation marker gene tnfa.  $\Delta$ Ct values were calculated relative to a  $\beta$ -actin control. Two-tailed Student's t test, t, t and t split indicates SEM; t and t split indicates

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Video 1. **Long-term live-imaging of** *foxp3a:EGFP***-positive cells.** Cells were imaged as described in Materials and methods. An area of skin posterior to the operculum was imaged. A time-lapse rendered video of an originally 20-min video is shown.

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