SUPPLEMENTAL MATERIAL

Bajrami et al., http://www.jem.org/cgi/content/full/jem.20160393/DC1

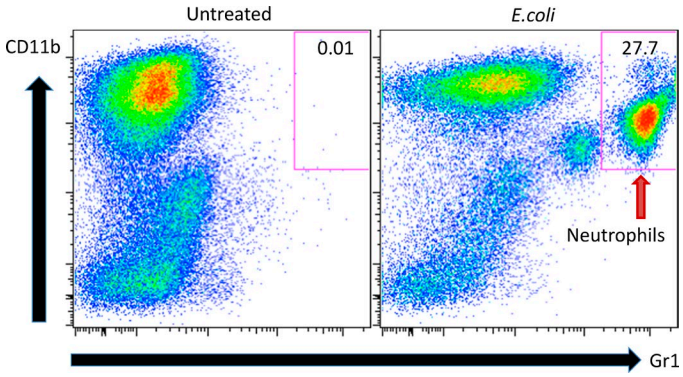
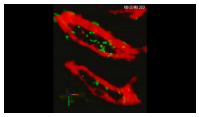
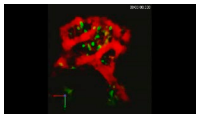


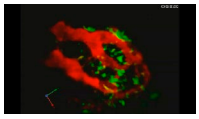
Figure S1. **Flow cytometry analysis of peritoneal cells after i.p. injections of *E. coli*.** C57BL/6 mice were injected i.p. with 3×10^5 *E. coli*. Peritoneal lavage was collected at 4 h after injection. Peritoneal cells were passed through a strainer and stained with Gr1-APC (108412; BioLegend) and CD11b-FITC (101208; BioLegend) antibodies. Neutrophils were defined as $Gr1^+CD11b^+$ cells.



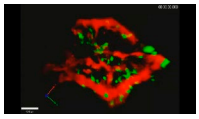
Video 1. **Neutrophil (green) mobilization from the BM to circulation (red) in an untreated mouse.** The mobilization of GFP^+ cells was monitored in vivo by MP-IVM. Images were acquired every 30 s for 70 min. The experiment was conducted as described in Fig. 4. Images were acquired using a 20× water immersion objective.



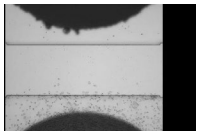
Video 2. **MIP-2–induced neutrophil (green) mobilization from the BM to circulation (red).** The mobilization of GFP^+ cells was monitored in vivo by MP-IVM. Images were acquired every 30 s for 60 min. The experiment was conducted as described in Fig. 4. Images were acquired using a 20× water immersion objective.



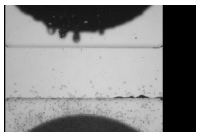
Video 3. **G-CSF–induced neutrophil (green) mobilization from the BM to circulation (red).** The mobilization of GFP^+ cells was monitored in vivo by MP-IVM. Images were acquired every 30 s for 70 min. The experiment was conducted as described in Fig. 4. Images were acquired using a 20× water immersion objective.



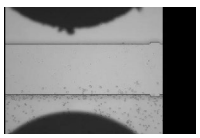
Video 4. **The effect of G-CSF on MIP-2–induced neutrophil (green) mobilization from the BM to circulation (red).** The mobilization of GFP^+ cells was monitored in vivo by MP-IVM. Images were acquired every 30 s for 70 min. The experiment was conducted as described in Fig. 4. Images were acquired using a 20× water immersion objective.



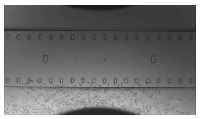
Video 5. **Chemotaxis of mouse neutrophils (untreated) in response to a gradient generated by addition of 1 µl MIP-2 (500 ng/mL) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 5. Images were acquired using a 10× dry lens.



Video 6. **Chemotaxis of G-CSF (1 µg/ml)–treated mouse neutrophils in response to a gradient generated by addition of 1 µl MIP-2 (500 ng/mL) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 5. Images were acquired using a 10× dry lens.

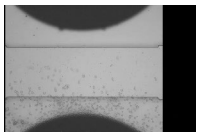


Video 7. **Chemotaxis of mouse neutrophils in response to a gradient generated by addition of 1 µl G-CSF (500 ng/mL) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 5. Images were acquired using a 10× dry lens.



Video 8. **Chemotaxis of mouse neutrophils (untreated) in response to a gradient generated by addition of 1 µl fMLP (1µM) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 5. Images were acquired using a 10× dry lens.

Video 9. **Chemotaxis of G-CSF (1 µg/ml)–treated mouse neutrophils in response to a gradient generated by addition of 1 µl fMLP (1µM) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 5. Images were acquired using a 10× dry lens.



Video 10. **Chemotaxis of mouse neutrophils treated with 1 µg/ml G-CSF and 0.5 µg/mL JAK inhibitor INCB018424 in response to a gradient generated by addition of 1 µl MIP-2 (500 ng/mL) to the chemoattractant reservoir of the EZ-TAXIScan chemotaxis device.** Images were acquired every 30 s for 20 min. The experiment was conducted as described in Fig. 6. Images were acquired using a 10× dry lens.