SUPPLEMENTAL MATERIAL

León et al., http://www.jem.org/cgi/content/full/jem.20131692/DC1

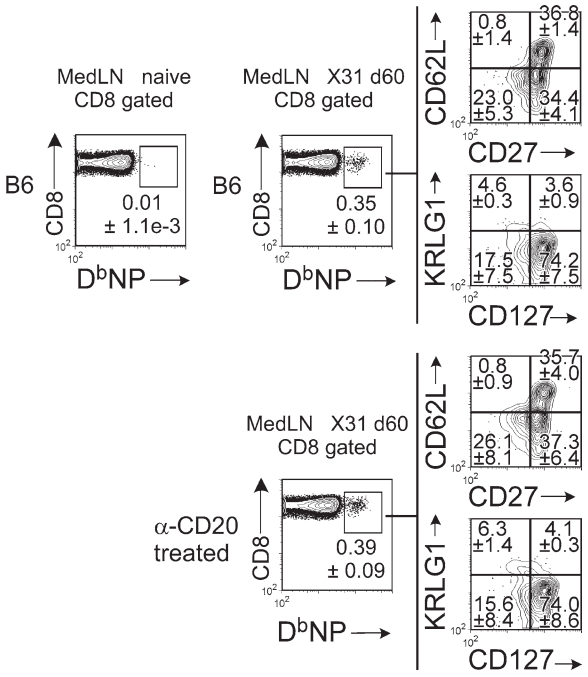

Figure S1. Flow cytometry gating strategy for identification of NP-specific memory CD8 T cell subpopulations. B6 mice were treated with anti-CD20 or control Ab 4 d before X31 infection. 60 d after infection, memory NP+ CD8 T cells from the medLN were identified by flow cytometry, using cells from the medLN of naive mice as a control to set the NP-specific gate. NP-specific CD8 T cells were then analyzed for surface expression of CD62L, CD27, CD127, and KRLG1 using the gates shown. Quantitative data are shown in Fig. 2 F.

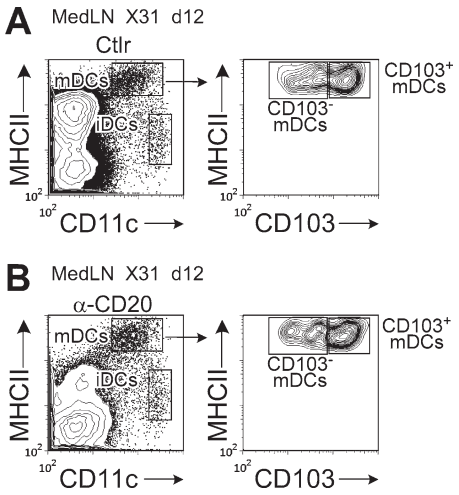

Figure S2. Gating strategy for DC subsets in the medLN of flu-infected B cell depleted animals. B6 mice were treated with control Ab (A) or anti-CD20 (B) 4 d before X31 influenza infection. MHCII^{hi}CD11c^{med} mature DCs (mDCs) and MHCII^{lo}CD11c^{hi} immature DCs (iDCs) in the medLN were identified. The mDCs were further subdivided into CD103^{+} and CD103^{-} populations. Quantitative data are shown in Fig. 3 D.

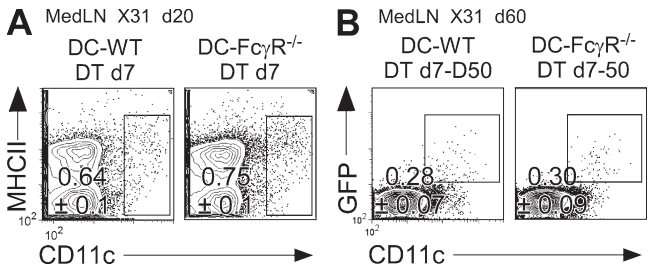

Figure S3. Gating strategy to identify DCs in the medLN of DT-treated DC-WT and DC-FcyR$^{-/-}$ chimeras after flu infection. B6 mice were irradiated and reconstituted with a 80:20 mixture of BM from CD11c-DTR and B6 donors (DC-WT chimeras) or from CD11c-DTR and μMT.FcyR$^{-/-}$ donors (DC-FcyR$^{-/-}$ chimeras). To eliminate DCs derived from the CD11c-DTR progenitors, chimeras were injected with PBS or DT beginning on day 7 after infection with X31 and continuing every 3 d until day 50. MedLN cells were analyzed by flow cytometry on day 20 after infection (A) or day 60 after infection (B). Total DCs were identified as CD11c^{+}MHCII^{+} cells (A). DCs derived from the CD11c-DTR BM precursors express the DTR transgene reporter GFP and were identified as CD11c^{+}GFP^{+} (B). Quantitative data from the flow plots are shown in Fig. 10 (B and I).