

## SUPPLEMENTAL MATERIAL

Ge et al., <http://www.jem.org/cgi/content/full/jem.20130731/DC1>**Table S1.** Serum IgM and IgG levels of NZM2328 and R27 at different ages

Strain (age)	IgM (mg/ml)	IgG (mg/ml)	IgG1 (mg/ml)	IgG2a (mg/ml)	IgG2b (mg/ml)
R27 (2–3 mo), <i>n</i> = 7	0.25 ± 0.046				
R27 (5–6 mo), <i>n</i> = 5	0.65 ± 0.20				
R27 (12 mo), <i>n</i> = 13	0.80 ± 0.45				
NZM2328 (6–7 mo), <i>n</i> = 4	0.79 ± 0.40				
R27 (2–3 mo), <i>n</i> = 7		1.08 ± 0.45	0.33 ± 0.57	0.17 ± 0.10	0.21 ± 0.039
R27 (5–6 mo), <i>n</i> = 5		3.62 ± 1.49	2.16 ± 2.04	0.32 ± 0.11	0.51 ± 0.18
R27 (8–9 mo), <i>n</i> = 7		8.12 ± 2.31	2.11 ± 1.89	0.67 ± 0.23	0.48 ± 0.13
R27 (12 mo), <i>n</i> = 15		7.36 ± 2.68	4.99 ± 2.47	0.56 ± 0.36	0.56 ± 0.27
NZM2328 (2 mo), <i>n</i> = 4		1.51 ± 0.46	0.13 ± 0.07	0.098 ± 0.057	0.096 ± 0.025
NZM2328 (5–6 mo), <i>n</i> = 5		5.7 ± 4.03	0.83 ± 0.64	0.87 ± 1.09	0.28 ± 0.21
NZM2328 (7–9 mo), <i>n</i> = 4		9.31 ± 4.22	1.82 ± 0.80*	1.37 ± 0.95*	0.59 ± 0.43

Serum levels of IgM, IgG, IgG1, IgG2a, and IgG2b were determined by ELISA. Values represent the mean ± SD. Student *t* tests were performed between 12-mo-old R27 and 7–9-mo-old NZM2328. \*, *P* < 0.05.

**Table S2.** Percentages and activation status of CD4<sup>+</sup> T cells (CD4<sup>+</sup>CD3<sup>+</sup>), CD8<sup>+</sup> T cells (CD8<sup>+</sup>CD3<sup>+</sup>), and B cells (CD19<sup>+</sup>) in spleens and kidney draining lymph nodes of 2–3-mo-old R27 and NZM2328 female mice were determined by flow cytometry

Cell subpopulation	R27 spleen	NZM2328 spleen	R27 LN	NZM2328 LN
CD4 <sup>+</sup> T of total live cells (%)	41.14 ± 2.30 ( <i>n</i> = 10)	39.71 ± 2.94 ( <i>n</i> = 10)	62.19 ± 7.73 ( <i>n</i> = 5)	60.15 ± 8.19 ( <i>n</i> = 5)
CD69 <sup>+</sup> of CD4 <sup>+</sup> T cells (%)	7.16 ± 1.68 ( <i>n</i> = 8)	7.50 ± 1.39 ( <i>n</i> = 8)	10.30 ± 2.36 ( <i>n</i> = 3)	10.36 ± 0.67 ( <i>n</i> = 3)
CD8 <sup>+</sup> T of total live cells (%)	9.29 ± 1.81 ( <i>n</i> = 10)	9.47 ± 2.29 ( <i>n</i> = 10)	11.69 ± 2.72 ( <i>n</i> = 5)	10.08 ± 5.02 ( <i>n</i> = 5)
CD69 <sup>+</sup> of CD8 <sup>+</sup> T cells (%)	2.47 ± 0.46 ( <i>n</i> = 6)	2.72 ± 0.95 ( <i>n</i> = 6)	3.88 ± 0.77 ( <i>n</i> = 3)	3.60 ± 0.40 ( <i>n</i> = 3)
B cells of total live cells (%)	36.13 ± 2.56 ( <i>n</i> = 10)	38.75 ± 6.10 ( <i>n</i> = 10)	15.52 ± 7.18 ( <i>n</i> = 5)	16.48 ± 6.11 ( <i>n</i> = 5)
CD86 on B cells (MFI)	22.11 ± 12.31 ( <i>n</i> = 8)	21.08 ± 10.06 ( <i>n</i> = 8)	8.85 ± 1.35 ( <i>n</i> = 3)	11.93 ± 2.00 ( <i>n</i> = 3)

Data represent mean ± SD. Two-tailed unpaired student *t* tests were performed to compare values of R27 and NZM2328 counterparts, and no significant differences were detected.

**Table S3.** Genes that are located within the refined Cgnz1 region

Gene (abbreviation)
Fc receptor, IgG, low affinity IV (Fcgr4 [partial]*)
Fc receptor, IgG, low affinity III (Fcgr3*)
RIKEN cDNA 1700009P17 gene (1700009P17Rik)
succinate dehydrogenase complex, subunit C, integral membrane protein (Sdhc)
myelin protein zero (Mpz)
Purkinje cell protein 4-like 1 (Pcp4l1)
nuclear receptor subfamily 1, group I, member 3 (Nr1i3)
translocase of outer mitochondrial membrane 40 homolog-like (yeast) (Tomm40l)
apolipoprotein A-II (Apoa2)
Fc receptor, IgE, high affinity I, gamma polypeptide (Fcer1g)
NADH dehydrogenase (ubiquinone) Fe-S protein 2 (Ndufs2)
a disintegrin-like and metalloproteinase (reprolysin type) with thrombospondin type 1 motif, 4 (Adamts4)
betaGlcNAc beta 1,4-galactosyltransferase, polypeptide (B4galt3)
protoporphyrinogen oxidase (Ppox)
ubiquitin specific peptidase 21 (Usp21)
ubiquitin-fold modifier conjugating enzyme 1 (Ufc1)
death effector domain-containing (Dedd)
nitrilase 1 (Nit1)
prefoldin 2 (Pfdn2)
kelch domain containing 9 (Klhdc9)
poliovirus receptor-related 4 (Pvrl4)
Rho GTPase activating protein 30 (Arhgap30)
upstream transcription factor 1 (Usf1)
F11 receptor (F11r)
RNA and export factor binding protein 2 (Refbp2)
intelectin 1 (galactofuranose binding) (Itln1)
CD244 natural killer cell receptor 2B4 (Cd244)
lymphocyte antigen 9 (Ly9)
SLAM family member 7 (Slamf7)
CD48 antigen (Cd48)
signaling lymphocytic activation molecule family member 1 (Slamf1)
CD84 antigen (Cd84)
SLAM family member 6 (Slamf6)
vang-like 2 (van gogh, <i>Drosophila</i> ) (Vangl2)
nescient helix loop helix 1 (Nhlh1)
nicastatin (Ncstn)
peroxisomal biogenesis factor 19 (Pex19)
WD repeat domain 42A (Wdr42a)
phosphoprotein enriched in astrocytes 15A (Pea15a)
calsequestrin 1 (Casq1)
ATPase, Na <sup>+</sup> /K <sup>+</sup> transporting, alpha 4 polypeptide (Atp1a4)
ATPase, Na <sup>+</sup> /K <sup>+</sup> transporting, alpha 2 polypeptide (Atp1a2)
immunoglobulin superfamily, member 8 (Igsf8)
potassium inwardly rectifying channel, subfamily J, member 9 (Kcnj9)
potassium inwardly rectifying channel, subfamily J, member 10 (Kcnj10 [partial]*)

\*, genes located within the boundaries of the recombination sites.

**Table S4.** Nine candidate genes for Cgnz1 with nucleotide replacements in promoter, exon, and intro regions

Genes	Amino acid mutations	Nuclear acid mutations		
		Promoter region	Exon	Intron
Apoa2	43D>E, 49M>V, 61A>V	-262T>C, -296T>C, -424A>T, -520C>T, -638T>C, -670G>A, -746T>G, -961C>G	635A>G, 644G>A, 668A>G, <u>689T&gt;A</u> , <u>705A&gt;G</u> , <u>742C&gt;T</u> , 922T>C	468T>C, 491G>C, 499C>A, 591A>G, 812C>A, 837A>G, 923G>A, 966T>C, 1062T>C, 1064T>C, 1080C>T
1700009P17Rik	—	-308C>T, -490A>G, -491C>T, -775T>C, -785T>C, -829G>A, -848G>T, -863T>C, -904G>C, -934T>A, -961C>T	1408A>G, 5107T>G	190T>C, 284C>T, 304T>C, 366G>C, 545A>T, 548T>A, 549C>G, 551G>A, 554T>C, 586C>T, 978T>C, 1072A>G, 1107G>A, 1185C>A, 1187G>A, 1214G>T, 1240C>A, 1356C>T, 1549T>G, 1833T>C, 1942T>C, 2272A>C, 2438C>T, 2498A>G, 3014G>A, 3074C>T, 3275G>C, 3440T>A, 3936A>G, 4110T>C, 4205C>T, 4208T>C, 4213T>G
Tomm40l	—	-164G>A, -183T>C, -221T>G, -259T>C, -293A>G, -530G>C, -792C>T	529A>C	180C>A, 224G>A, 405G>A, 725G>C, 735T>C, 786C>T, 833T>G, 834A>G, 1101C>T, 1211T>C, 1543G>C, 2589G>A, 2765G>A, 2811G>A, 3022T>G, 3164C>G
Ndufs2	8R>G	-1138G>A, -1259G>A, -1426T>C	<u>46G&gt;C</u> , 105T>C, 919A>G, 964C>G, 7820A>G, 8782G>A, 9109G>A, 9142G>A, 9166A>G, 9928C>T	210T>C, 492T>C, 647A>G, 739T>C, 874G>A, 895T>C, 1129T>C, 1241C>T, 1323G>A, 1348C>T, 1874G>T, 1938G>C, 1939T>C, 1940G>C, 1982T>C, 2053C>T, 2079A>C, 2110G>A, 2459C>G, 2489C>T, 2496T>G, 2711A>G, 2804A>T, 2805A>G, 2806C>T, 2877C>T, 2915A>G, 2916A>T, 2917A>G, 2940A>G, 2965A>C, 2968A>C, 2971A>C, 3152G>T, 3201G>A, 3226C>T, 3229T>C, 3261T>C, 3284A>G, 3439C>G, 3534A>G, 3540G>A, 3585A>G, 3665T>C, 3763A>G, 4200C>T, 4225T>G, 4538T>C, 4551T>C, 4572A>C, 4590C>T, 4620C>T, 4674A>G, 4935T>C, 4990T>C, 5139C>T, 5143C>A, 5150G>C, 5197C>T, 5255G>A, 5370A>C, 5372C>T, 5843A>G, 5887T>C, 6272G>A, 6284C>G, 7058T>A, 7106G>A, 7190G>A, 7218G>C, 7242C>G, 7273G>C, 7289C>G, 7361G>C, 7569A>C, 7637C>G, 7649A>G, 7929A>G, 7931T>(DEL), 7932A>(DEL), 7987T>C, 8002T>C, 8037C>T, 8159T>G, 8496C>T, 8619T>C, 8675C>T, 8729C>G, 8733C>T, 8978C>T, 9179A>T, 9198T>C, 9218T>A, 9278G>C, 9348T>C, 9535G>A, 9595C>A, 9616T>C, 9665G>A, 9986C>T, 9994C>T, 9996C>G, 9999C>(DEL), 10000C>T, 10033T>G, 10035A>G, 10042G>C, 10045T>C, 10108T>A, 10114G>A, 10115T>C, 10166T>(DEL), 10270T>G, 10278A>T, 10834G>A, 10839A>C, 10861A>C, 11033C>T, 11153A>G, 11216A>T, 11301C>A, 11517G>A, 11738A>G, 11755A>T, 11902C>T
Pex19	55P>S	-47G>A, -75A>G	<u>1969C&gt;T</u> , 6381A>G	174A>G, 311C>A, 1140A>G, 1145C>A, 1209A>G, 1350G>T, 1825G>A, 2197C>A, 2286C>T, 2643A>G, 2689G>C, 2852G>C, 3236A>G, 3484A>G, 3489A>G, 3490C>G, 3516T>C, 3567T>C, 3725T>C, 4204T>C, 4463T>G, 4582C>T, 4710G>C, 4996A>G, 5012A>C, 5081T>C, 5117G>A, 5878G>A, 6074A>G, 6075A>T, 6078A>(DEL), 6198C>G, 6231C>T, 6248T>A, 6589C>T, 6856G>A, 6934A>G, 6980A>G, 7027C>G, 7158T>C

**Table S4.** Nine candidate genes for Cgnz1 with nucleotide replacements in promoter, exon, and intro regions (*Continued*)

Genes	Amino acid mutations	Nuclear acid mutations		
		Promoter region	Exon	Intron
Casq1	–	–543A>G, –749C>T	364G>A, 430G>A, 4861G>C, 6558G>A, 9395G>C	135C>T, 495T>C, 658G>A, 804G>A, 823T>C, 1180A>G, 1224G>A, 1268A>G, 1305T>C, 1312C>A, 1415T>A, 1516T>C, 1551G>A, 1691A> G, 1812G>A, 2452G>T, 2575C>G, 2646A>C, 2671C>A, 2939T>C, 3151C>G, 3249G>A, 3369C>T, 3623G>A, 4075C>A, 4076G>A, 4226G>A, 4235C>T, 4887C>T, 5110A>C, 5305G>A, 5431G>A, 5473G>A, 5640A>G, 6295G>A, 6710T>C, 7148G>A, 7518A>G, 7855A>G, 7975G>A, 8047T>C, 8096A>G, 8124C>T, 8195G>A, 8201A>G, 8213T>A, 8267A>G, 8291T>C, 8299G>C, 8365G>A, 8518C>T, 8743C>T, 8872C>T, 9315C>T, 9321C>T, 9557C>A
Dedd	–	–223T>C, –299T>C, –621C>T, –678G>A, –684T>C	9420C>T, 9492A>C	109C>A, 1000G>T, 1668G>T, 1741T>C, 2213A>G, 2222A>T, 2379A>G, 2433C>G, 2445G>T, 2478C>(DEL), 2488C>T, 2505G>C, 2681C>T, 3094A>G, 3134C>5, 3255G>A, 3279G>T, 3687T>C, 3897G>A, 4465C>G, 5117C>T, 5209T>A, 5337C>A, 5481G>C, 5740A>G, 5790G>T, 6147G>C, 6148G>A, 6355G>A, 6384G>A, 6421G>T, 6439G>A, 6519G>A, 7353T>C, 7696G>A, 7808A>G, 7837T>A, 7838G>A, 8007G>A, 8411G>A, 8700A>G, 8836T>C, 8840G>(DEL), 8853G>T, 9072G>(DEL), 9073C>(DEL), 9868G>A, 10234T>C, 10352G>A, 10356A>G, 10491C>A, 10541G>A, 10597C>T, 10910T>T, 11218A>G, 11219C>A, 11432G>A, 11540A>G
Itln1	207L>V, 289D>N, 295N>T, 302K>E	–45G>A, –51A>G, –96A>G, –191A>T, –752G>C	519A>C, 3681G>T, 4150G>A, 4689T>C, <b>4735A&gt;C</b> , 16961A>G, <b>16977C&gt;T</b> , <b>16996T&gt;G</b> , <b>17016T&gt;C</b>	27G>A, 86A>C, 365A>T, 805T>G, 1119A>C, 1137C>T, 1273A>G, 1306T>C, 1384A>G, 1512T>G, 1518G>A, 1576T>G, 1604A>T, 1646G>A, 1647C>T, 1661T>G, 1706C>T, 1960A>T, 2054C>T, 2262A>G, 2264C>G, 2293C>T, 2327C>G, 2349A>T, 2369G>A, 2505G>A, 2528T>C, 2600A>G, 2714C>T, 2777C>A, 2781G>A, 2813C>G, 2864C>G, 2907A>G, 3151T>G, 3194G>A, 3222C>T, 3272A>G, 3273A>G, 3289T>C, 3336A>C, 3394T>A, 3401T>G, 3415T>A, 3440C>G, 3442T>G, 3444A>C, 3452A>C, 3476C>T, 3477G>A, 3501T>G, 3516T>C, 4273T>A, 4331G>C, 4334A>T, 4409C>T, 4843C>T, 5098G>T, 5246T>C, 5301C>A, 5718G>A, 5929G>C, 6044T>C, 6243A>G, 6405A>G, 6548A>G, 6827T>C, 6840C>A, 6994A>G, 7014C>G, 7030C>T, 7034C>T, 7066G>T, 7067G>C, 7675T>G, 7695G>A, 7906C>A, 8001T>C, 8075C>G, 8182A>T, 8187C>A, 8311T>G, 8408A>T, 8725T>C, 8807C>A, 8895G>T, 8927G>C, 8951C>T, 9053A>G, 9094G>A, 9176A>T, 9355T>C, 9365T>C, 9391A>T, 9451C>T, 9500T>G, 9554T>C, 9596A>C, 9725G>A, 9794C>A, 9839T>C, 9928A>G, 9979C>T, 9988C>G, 10015C>A, 10033T>A, 10039T>C, 10066G>A, 10068T>C, 10093T>C, 10098T>C, 10253C>T, 10303T>C, 10457T>A, 15835A>(DEL), 16121C>T, 16626T>C, 16672A>G, 16729A>T, 16754C>T, 16775A>G, 16779A>G, 16868C>G, 16891T>C, 17126C>T, 17127C>G
Pea15a	–	–237C>G, –262A>G, –297T>C, –316T>C, –575A>T, –670A>G, –753T>C, –775C>T, –846A>G, –859G>T, –891A>G, –946A>T	6331A>G, 7532A>T	321T>C, 595G>T, 712A>C, 733C>G, 839T>C, 1523C>T, 1651T>G, 1797T>C, 1859C>G, 1860T>A, 2252A>G, 2326C>T, 2342G>T, 2402G>T, 2883C>G, 3105G>C, 3193C>T, 3336A>G, 3737G>C, 3800A>G, 3851G>A, 5288T>C, 5353G>A, 5367G>A, 5922C>A, 6181A>C, 7104T>G, 8053C>T, 8381C>T, 8388T>A, 8690A>G, 9191A>G, 9702G>A

Bolding and underlining indicate that the nucleotide change would change the amino acid to be coded. This means that the point mutation is nonsilent mutation.