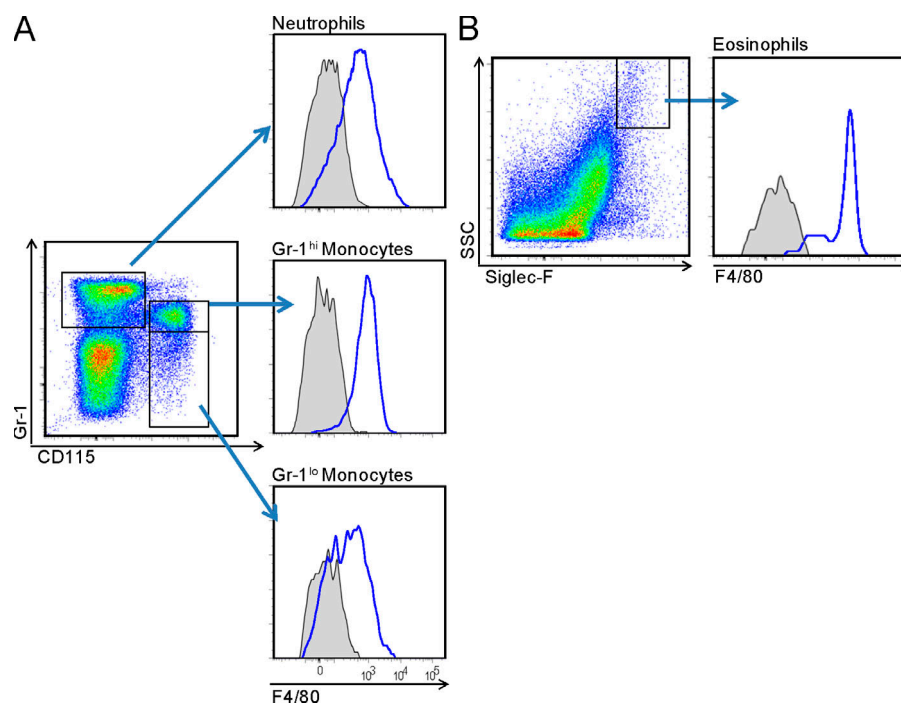
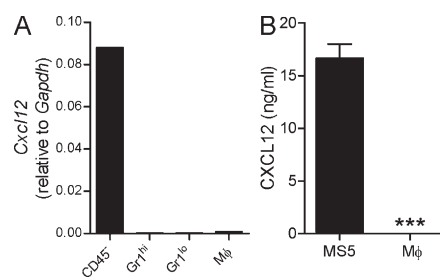


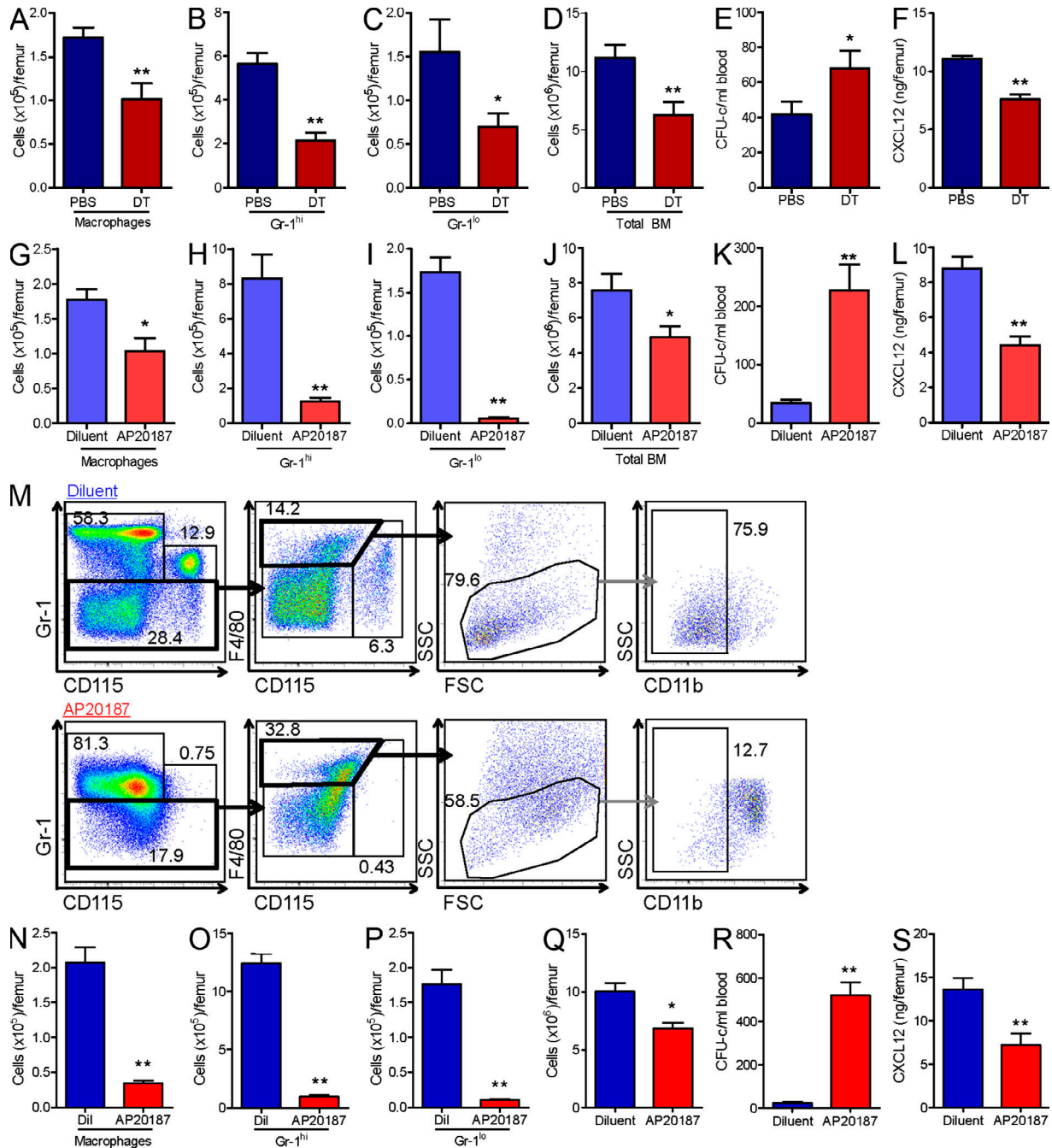
## SUPPLEMENTAL MATERIAL

Chow et al., <http://www.jem.org/cgi/content/full/jem.20101688/DC1>

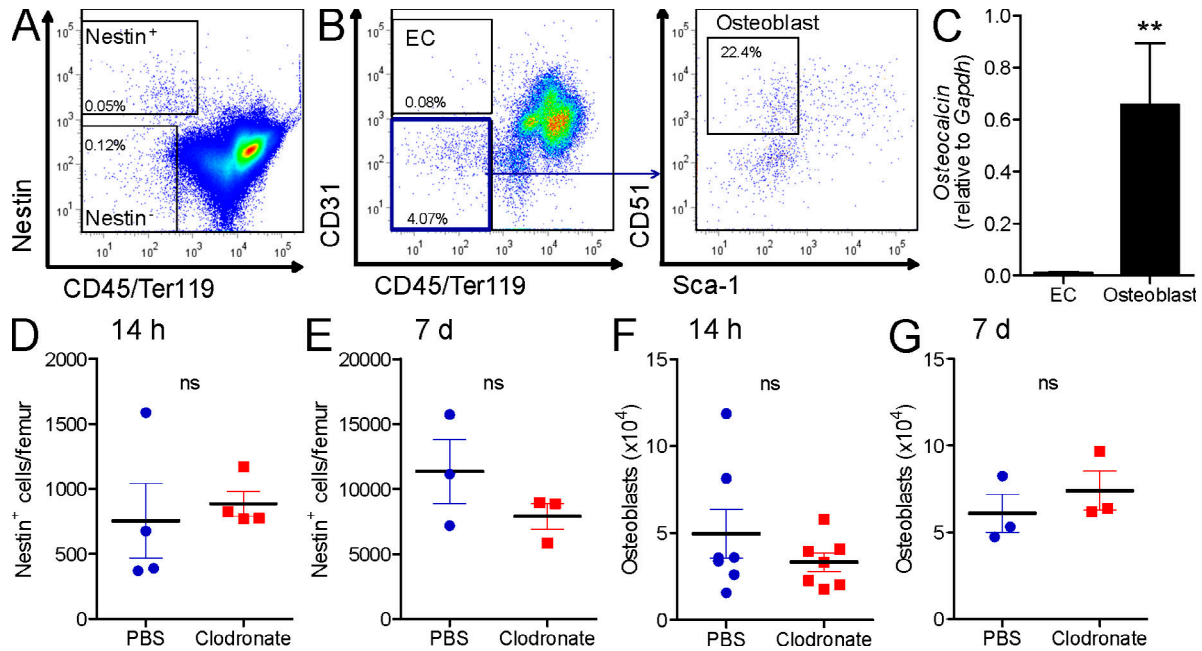
**Figure S1. BM neutrophils, MOs, and eosinophils express F4/80.** (A) Total DAPI<sup>-</sup> single cells were gated by Gr-1 and CD115 staining. Staining with F4/80 (blue line) and isotype control (gray shaded histogram) on Gr-1<sup>+</sup> CD115<sup>-</sup> neutrophils, Gr-1<sup>+</sup> CD115<sup>+</sup> Gr-1<sup>hi</sup> MOs, and Gr-1<sup>-</sup> CD115<sup>+</sup> Gr-1<sup>lo</sup> MOs is displayed. (B) Eosinophils were gated as SSC<sup>hi</sup>Siglec-F<sup>+</sup> cells and expression of F4/80 (blue) and isotype control (gray) is displayed.



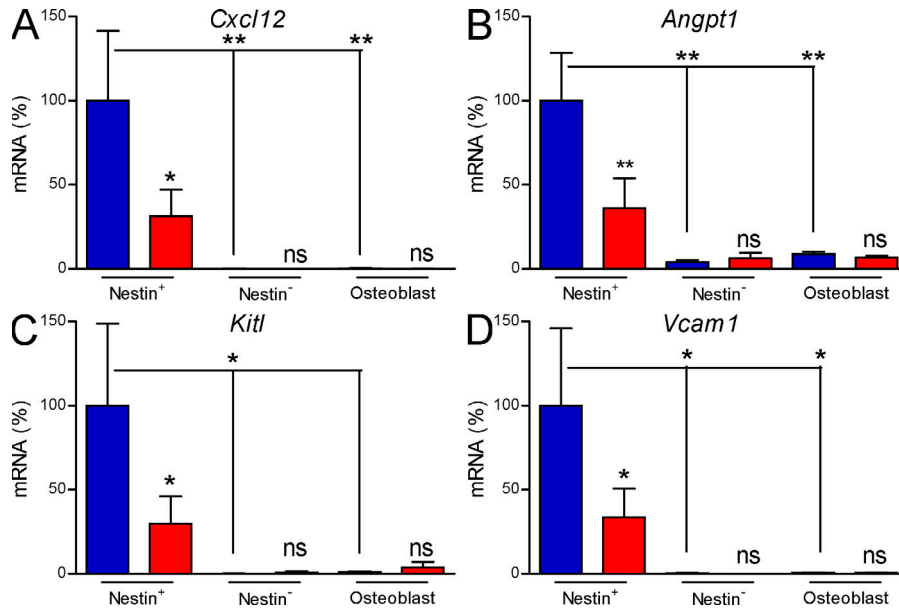
**Figure S2. MΦ do not produce CXCL12.** (A) CD45<sup>-</sup> cells, Gr-1<sup>hi</sup> MOs, Gr-1<sup>lo</sup> MOs, and MΦ were sorted from the BM, and expression of *Cxcl12* relative to *Gapdh* was assessed by Q-PCR. (B) Levels of CXCL12 in the supernatant of MS-5 culture in complete medium and in the supernatant from BMDM were measured by ELISA. Each experiment was performed once. \*\*\*, P < 0.001



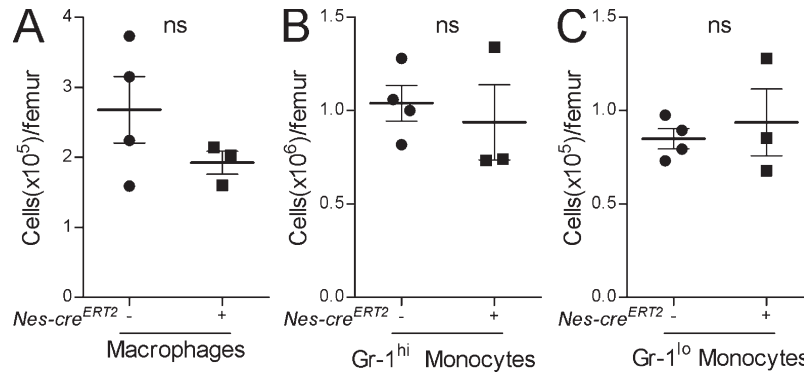
**Figure S3. Three other models in which mononuclear phagocyte depletion is associated with robust HSC/progenitor mobilization and CXCL12 reduction.** (A–F) CD11b-DTR BM chimeras treated with two doses of PBS (dark blue) or DT (dark red). Quantification of BM mononuclear phagocytes ( $n = 9$ ; A–C), total nucleated cells ( $n = 9$ ; D), CFU-C ( $n = 15$ –16; E), and BMEF CXCL12 levels ( $n = 7$ ; F). Data are pooled from three independent experiments. (G–L) Mafia BM chimeras were injected with diluent (purple) or AP20187 (light red) and analyzed 24 h after the last injection. Quantification of mononuclear phagocytes ( $n = 5$ ; G–I) and total nucleated cells per femur ( $n = 5$ ; J), CFU-C per milliliter of blood ( $n = 11$ –13; K), and BMEF CXCL12 ( $n = 5$ ; L). G–J and L show representative data from three independent experiments; K represents pooled data from three independent experiments. (M–S) Non-transplanted Mafia mice were injected with diluent (blue) or AP20187 (red) and analyzed 24 h after the final injection. (M) MΦ were gated by Gr-1<sup>lo</sup> CD115<sup>int</sup> F4/80<sup>+</sup> SSC<sup>int/lo</sup> CD11b<sup>lo</sup>. Gr-1<sup>lo</sup> CD115<sup>int</sup> F4/80<sup>+</sup> SSC<sup>int/lo</sup> CD11b<sup>hi</sup> gating demonstrated BM neutrophilia that leaked into the Gr-1<sup>lo</sup> CD115<sup>int</sup> F4/80<sup>+</sup> SSC<sup>int/lo</sup> gate after treatment with AP20187. (N–P) Quantification of absolute numbers of mononuclear phagocytes ( $n = 5$ –6) and CXCL12 levels in the BMEF by ELISA ( $n = 5$ –6) 1 d after the last injection of diluent (Dil) or AP20187 in nontransplanted Mafia mice. Data are pooled from two independent experiments and represented as mean  $\pm$  SEM. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ .



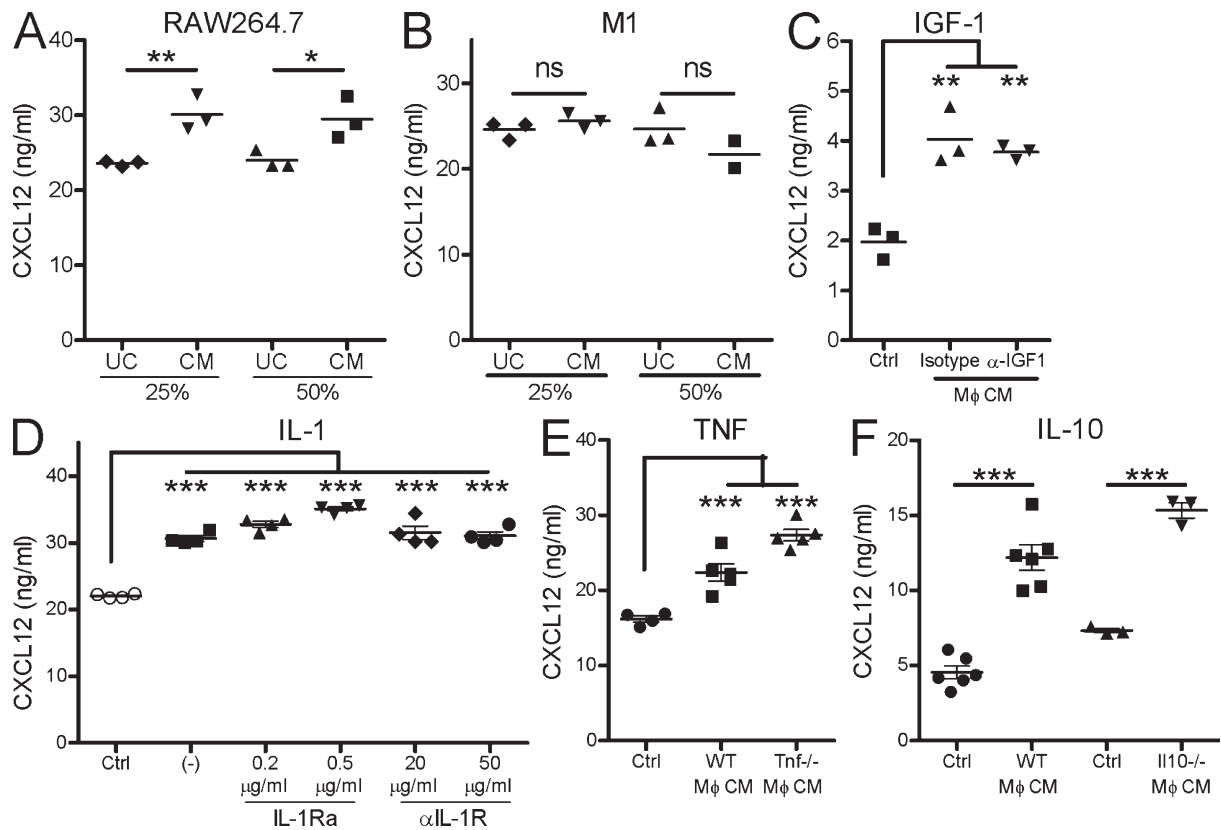
**Figure S4. Sorting BM Nestin<sup>+</sup> and Nestin<sup>-</sup> fractions and bone endothelial cell and osteoblast fractions.** (A and B) Sorting strategy for Nestin<sup>+</sup> and Nestin<sup>-</sup> fractions from the BM (A) and endothelial cells (EC) and osteoblasts from the bone (B). FACS plots were pregated on DAPI<sup>-</sup> single cells. (C) Q-PCR of *Osteocalcin* expression relative to *Gapdh* in in vivo-sorted endothelial cells (EC) and osteoblasts. Representative data from three independent experiments are shown ( $n = 3-4$ ). (D-G) Counts of Nestin<sup>+</sup> cells in one femur (D and E) and osteoblasts digested from femurs, tibias, and humeri (F and G) 14 h (D and F) and 7 d (E and G) after treatment with PBS- (blue circles) or clodronate-encapsulated (red squares) liposomes. (D) Representative data from three independent experiments are shown ( $n = 3-4$ ). (E and G) Experiments were performed once ( $n = 3$ ). (F) Data pooled from two independent experiments ( $n = 7$ ). \*\*,  $P < 0.01$ ; ns, not significant.



**Figure S5. Retention gene expression is reduced in Nestin<sup>+</sup> cells 7 d after mononuclear phagocyte treatment.** (A-D) Relative expression of *Cxcl12* (A), *Angpt1* (B), *Kitl* (C), and *Vcam1* (D) in Nestin<sup>+</sup> and Nestin<sup>-</sup> fractions sorted from the BM and osteoblasts sorted from the bone 7 d after treatment with PBS- (blue bars) or clodronate-encapsulated (red bars) liposomes. Data were analyzed by one-way ANOVA/Newman-Keuls test.  $n = 3$ . \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; ns, not significant



**Figure S6. Mononuclear phagocytes are not reduced 7 d after depletion of Nestin<sup>+</sup> cells.** Tamoxifen-treated *Nes-Cre<sup>ERT2</sup>/iDTR* (+) and control *iDTR* (–) animals were analyzed 7 d after administration of DT for numbers of MΦ (A), Gr-1<sup>hi</sup> MOs (B), and Gr-1<sup>lo</sup> MOs (C). ns, not significant.



**Figure S7. Soluble factor from MΦ, but not myeloblast, cell line enhances stromal CXCL12 production.** 200,000 RAW264.7 MΦ (A) and M1 myeloblast (B) cells were cultured in 300  $\mu$ l of their respective media. After 3 d of culture, the supernatant or fresh control medium was mixed with complete medium at either 25 or 50% concentration to make conditioned medium (CM) or unconditioned medium (UC), respectively. CM or UC was transferred onto 5,000 MS-5 stromal cells plated 24 h prior. CXCL12 levels were determined by ELISA. (C–F) 5,000 MS-5 cells were cultured in complete medium (Ctrl) or media conditioned by MΦ (MΦ CM) that was supplemented with blocking antibody to IGF-1 (C), IL-1 receptor antagonist (D), and blocking antibody to IL-1R (D). In E and F, the ability of MΦ CM derived from TNF- (E) and IL-10-deficient (F) animals to up-regulate CXCL12 production from MS-5 cells were compared with MΦ CM derived from wild-type animals. All experiments were performed 1–2 times. Data were analyzed by one-way ANOVA/Newman-Keuls test. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ ; ns, not significant.