SUPPLEMENTAL MATERIAL

Lochner et al., http://www.jem.org/cgi/content/full/jem.20100052/DC1

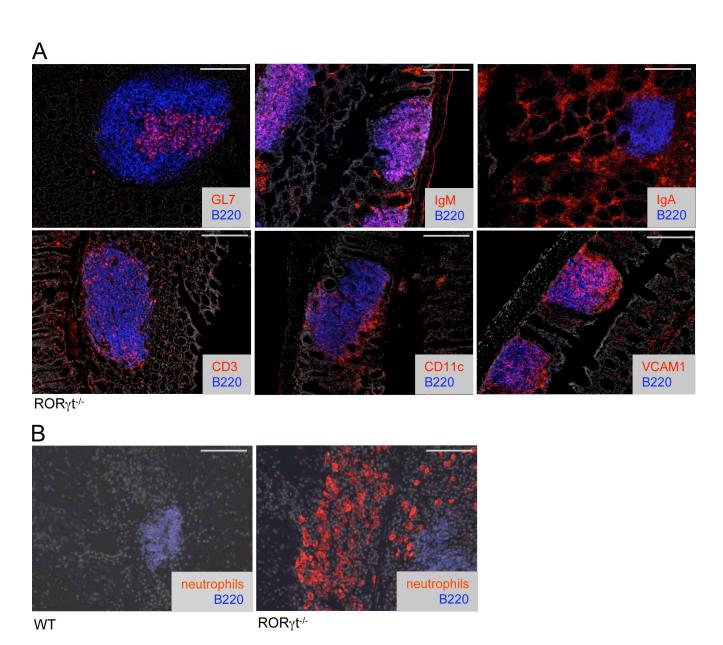


Figure S1. Colonic tLTs in sections of colons from ROR γ t^{-/-} mice after two cycles of DSS treatment. (A) Sections were stained with the indicated antibodies and with DAPI for nuclear staining (shown in gray). Bars, 200 μ m. (B) Sections were stained for neutrophils and B cells. Sections shown are representative for five (A) and three (B) individual mice in at least two individual experiments. Bars, 100 μ m.

JEM S1

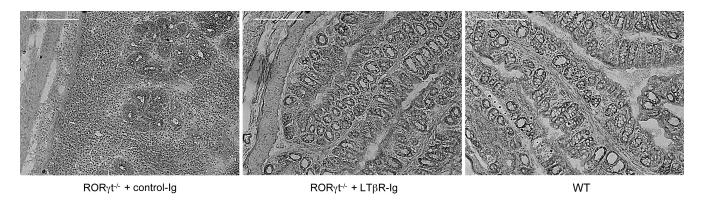


Figure S2. LTβR-Ig and antibiotics prevent DSS-induced colitis in ROR γ t-deficient mice. Wild-type or ROR γ t-/- mice were exposed to two cycles of DSS and treated either by weekly i.p. injections of LTβR-Ig protein or with a control Ig. H&E staining of representative sections form distal colon. Bar, 200 μm. Data are representative of three independent experiments.

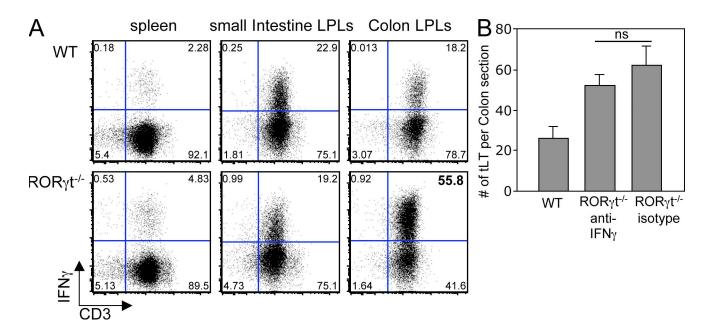


Figure S3. No role for IFN- γ in DSS-induced colitis in ROR γ t-deficient mice. ROR γ t-/- and wild-type control mice were treated with two cycles of DSS. (A) Lymphocytes were isolated from the spleen, small intestine, and colonic lamina propria and stained for IFN- γ production. FACS plots are gated on CD4+ T cells. (B) ROR γ t-/- mice were treated with two injections i.p. of 250 μg of neutralizing anti-IFN- γ antibody before the first and the second DSS cycle. The number of colonic tLTs was assessed as described in Materials and methods. Data are shown from two representative experiments. ns, nonspecific. Statistical significance was assessed by the paired Student's t test. Error bars are SD.

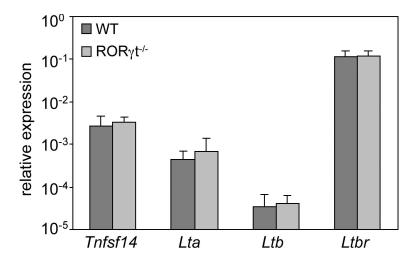


Figure S4. Expression of LTβR and LTβR ligands in DSS-treated ROR γ t-deficient mice. Quantitative real-time PCR on whole colon tissue from DSS-treated wild-type controls and ROR γ t-/- mice. Ct values were normalized to *Gapdh* expression. Data are shown from three independent experiments and four mice per group. Error bars are the SD.

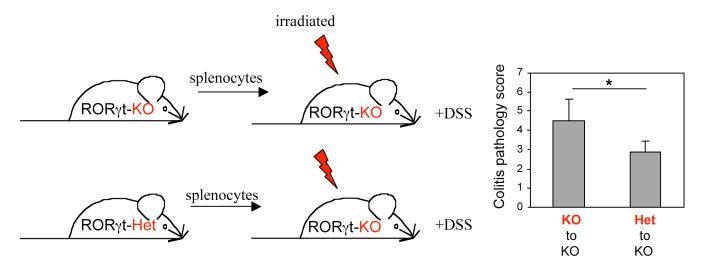


Figure S5. Complementation of ROR γ t-deficient mice with ROR γ t-sufficient spleen cells. Splenocytes were isolated from control or ROR γ t-deficient mice and transferred i.v. into sublethally irradiated ROR γ t^{-/-} mice. 4 wk after transfer, mice were treated with two cycles of DSS. At the end of the second cycle, the histological disease score in the colons of the mice was assessed as described. Data are shown from two independent experiments and three mice per group. Statistical significance was assessed by the paired Student's t test. Error bars are SD.

JEM S3