

## SUPPLEMENTAL MATERIAL

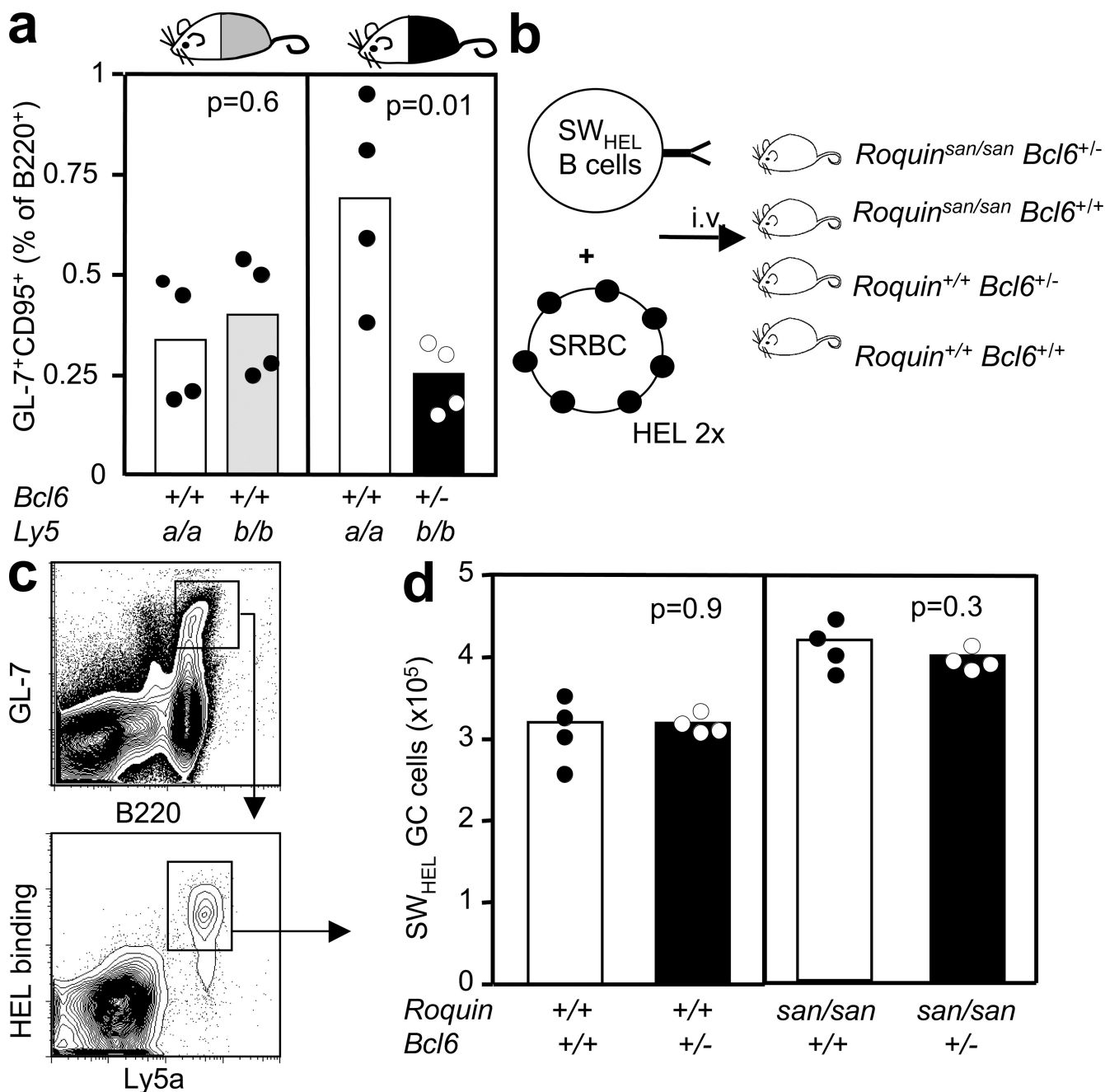
Linterman et al., <http://www.jem.org/cgi/content/full/jem.20081886/DC1>

Figure S1. Heterozygosity for *Bcl6* within the B cell compartment reduces the magnitude of the GC response. (a) Dot plots of B220<sup>+</sup>GL-7<sup>+</sup>CD95<sup>+</sup> GC B cells from mixed bone marrow chimeras 8 d after SRBC immunization. Chimeric mice were generated by sublethal irradiation of recipient C57BL/6-Ly5a mice and reconstitution with a 1:1 mix of either *Bcl6*<sup>+/+</sup>.Ly5<sup>b</sup>/*Bcl6*<sup>+/+</sup>.Ly5<sup>b</sup> or *Bcl6*<sup>+/+</sup>-Ly5<sup>a</sup>/*Bcl6*<sup>+/+</sup>-Ly5<sup>b</sup> bone marrow. Immunizations were performed 12 wk after reconstitution. Data are representative of two independent experiments ( $n = 4$  per group). (b) Experimental outline for evaluating the HEL-specific GC cell response after co transfer of SW<sub>HEL</sub> B cells and HEL-2x-conjugated SRBC. (c) Gating strategy for assessing the HEL-specific GC response. (d) Dot plots showing the total number of CD45.1 donor derived HEL-binding GL-7<sup>+</sup>CD95<sup>+</sup>B220<sup>+</sup> GC B cells 8 d post intravenous transfer of  $10^4$  SW<sub>HEL</sub> B cells and SRBC conjugated to HEL<sup>2x</sup> into 10-wk-old recipient mice of the genotypes indicated. Data are representative of 4 independent experiments ( $n = 4$  per group). In a and d each symbol represents one mouse and P values are indicated on the graphs.

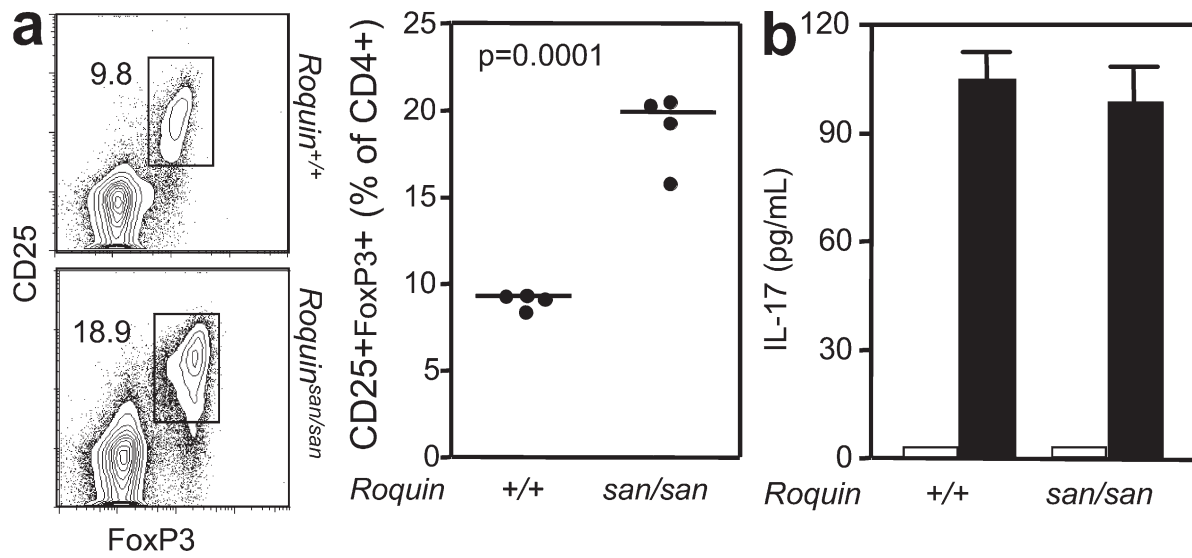


Figure S2. T reg cell number and IL-17 production are not reduced by the *san* allele of *Roquin*. (a) Representative flow cytometric contour plots (left) and graphical representation (right) of CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> T reg cells in 10-wk-old *Roquin*<sup>san/san</sup> mice compared with littermate controls (*Roquin*<sup>+/+</sup>). Each symbol represents one mouse. Data are representative of five independent experiments. The numbers in the plots represent percentages, and horizontal bars indicate medians. (b) ELISA was used to determine culture supernatant IL-17 levels from 24-h splenocyte cultures from *Roquin*<sup>san/san</sup> mice and littermate controls in the presence (shaded bars) or absence (open bars) of PMA and ionomycin. Data are representative of three independent tests. Error bars indicate means  $\pm$  SEM.

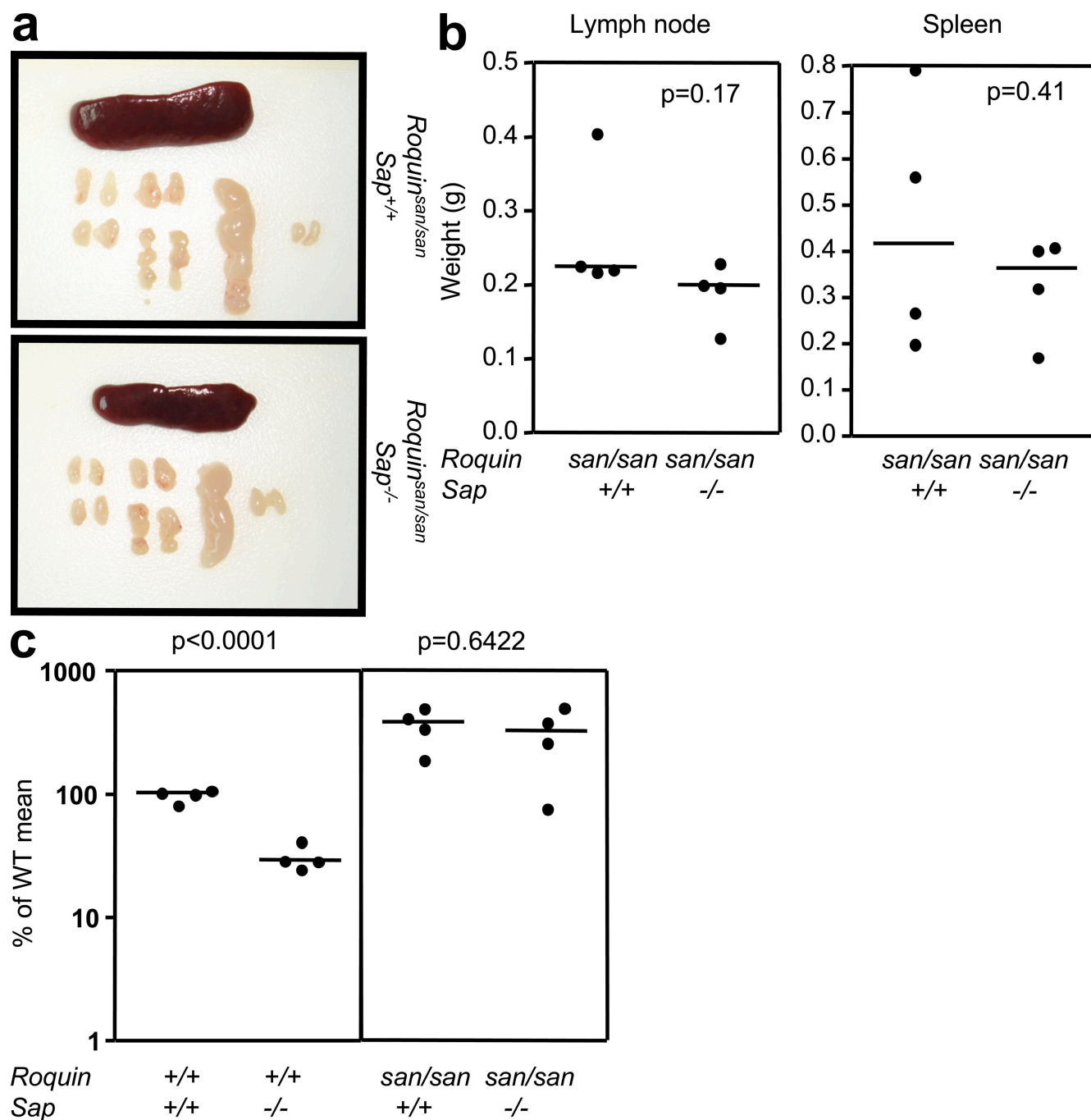


Figure S3. SAP deficiency does not correct the splenomegaly or lymphadenopathy seen in *Roquin<sup>san/san</sup>* mice. (a) Photographs of the spleen and lymph nodes (cervical, inguinal, axillary, subscapular, paraaortic, and mesenteric) from 10-wk-old *Roquin<sup>san/san</sup>* and *Roquin<sup>san/san</sup> Sap<sup>-/-</sup>* mice. (b) Spleen weights and (c) weights of pooled lymph nodes (cervical, inguinal, axillary, subscapular, paraaortic, and mesenteric) from 10-wk-old *Roquin<sup>san/san</sup>* and *Roquin<sup>san/san</sup> Sap<sup>-/-</sup>* mice. Differences are not statistically significant. (d) ELISA analysis of total serum IgG from unimmunized 10-wk-old mice of the indicated genotypes. Data are representative of three experiments. Horizontal bars indicate medians.

**Table S1.** Scoring strategy used to assess the severity of mouse nephritis

Score	Glomeruli		Tubulointerstitium	
	Cells	Matrix	Cells	Matrix
0	NAD	NAD	NAD	NAD
1	Hypercellularity only	Mesangial matrix increase	Very mild	Very mild
2	Proliferative or fibrinoid GN without crescents	Mild scarring	Mild	Mild
3	GN with fibrinoid or crescents in <50% of glomeruli	Moderate scarring	Moderate	Moderate
4	GN with fibrinoid or crescents in >50% of glomeruli	Fibrous obliteration	Severe	Severe

NAD, no abnormality detected.