

SUPPLEMENTAL MATERIAL

Cariappa et al., <http://www.jem.org/cgi/content/full/jem.20081399/DC1>**Mice**

Cmah mutant mice (1) were backcrossed for 10 generations into the C57BL/6 background and then studied. *Siae*^{Δ2/Δ2} and *Cmah*^{-/-} mice, both backcrossed for 10 generations into the C57BL/6 background, were intercrossed to generate *Siae*^{Δ2/Δ2}/*Cmah*^{-/-} double mutant animals.

B lymphocyte purification

The method used by Collins et al. (2) was adhered to.

RT-PCR

Standard methodology was used to prepare total cellular RNA and first-strand complementary DNA from splenocytes. The HPRT primers used in the PCR were as previously published (3). The *Siae* primers used to detect the deletion of exon 2 were the following: forward, 5'-AGTGCTGCTCATAATCGCGC-GAG-3'; and reverse, 5'-AAGTTTCCTGCGGTGGGCTTG-3'. The reverse primer spanned exons 4 and 5.

CFSE staining and B cell proliferation

Analysis of CFSE-stained cells was performed on nonstimulated and anti-IgM-activated B cells as described earlier (4) with the following modification: CFSE (Invitrogen) was added to a final concentration of 1 μM and no additional surface staining of the B cells was performed.

Lymphocyte proliferation assay

The method described previously (5) was used. Sorted cell fractions were plated at 10⁵/well in triplicate in 96-well flat-bottomed microtiter plates with medium alone or with medium and one of the following: goat anti-mouse IgM (Fab')₂ (μ chain specific; Jackson ImmunoResearch Laboratories), 1 μg/ml of purified hamster anti-mouse CD40 (BD), and 10 μg/ml LPS from *Escherichia coli* serotype O55:B5, (Sigma-Aldrich). The cells were cultured for 48 h and then pulsed with 2 μCi/well of [³H] thymidine (PerkinElmer) for an additional 18 h, harvested, and read in a beta plate reader.

Annexin V assay for apoptosis

Previously published methodology (6) was used. 5 μl of 10 μg/ml stock of APC-Annexin V (BD) was used and cells were assessed for apoptosis by flowcytometry.

Immunization with T cell-dependent and -independent antigens and ELISA for anti-DNP antibodies

Mice were immunized i.p. with 100 μg DNP-KLH (EMD) or DNP-Ficoll (Biosearch Technologies, Inc.) in PBS on day 0 and boosted on day 28. Serum was collected on designated days and antibodies were quantitated by ELISA as previously described (7).

Stable transfection of U2OS cells, and metabolic labeling-based pulse-chase studies

U2OS cells were stably transfected with a murine C-terminal Flag-tagged *Siae* cDNA in the pCDNA3.1 vector using the calcium phosphate method. *Siae*-expressing clones were identified using an anti-Flag Western blot approach. A stable clone expressing *Siae* was used in all immunofluorescence and metabolic labeling pulse-chase studies.

U2OS-*Siae* cells were metabolically labeled with 0.5 mCi of ³⁵S-methionine for 10 min and chased with complete medium containing 2 mM methionine for varying time periods. Culture supernatants were immunoprecipitated at various chase time points with anti-Flag antibodies and protein A Sepharose, separated on an SDS-polyacrylamide gel, and visualized by autoradiography.

Immunofluorescence of transfected cells

U2OS-*Siae* cells were fixed in 4% paraformaldehyde, permeabilized with 0.1% NP-40, and stained with labeled anti-Flag antibodies, Wheat germ agglutinin, LysoTracker, or LAMP-1 as indicated in Fig. S2. The method used by Collins et al. (2) was adhered to for the immunofluorescence of purified B lymphocytes seen in Fig. S6. Images were captured using a spinning disc confocal microscope (IX81; Olympus) equipped with a charge-coupled device camera and images were processed with Slidebook 4.2 (Intelligent Imaging Innovations, Inc.).

SUPPLEMENTAL REFERENCES

- Hedlund, M., P. Tangvoranuntakul, H. Takematsu, J.M. Long, G.D. Housley, Y. Kozutsumi, A. Suzuki, A. Wynshaw-Boris, A.F. Ryan, R.L. Gallo, et al. 2007. N-glycolylneuraminic acid deficiency in mice: implications for human biology and evolution. *Mol. Cell. Biol.* 27:4340–4346.
- Collins, B.E., B.A. Smith, P. Bengtson, and J.C. Paulson. 2006. Ablation of CD22 in ligand-deficient mice restores B cell receptor signaling. *Nat. Immunol.* 7:199–206.
- Cariappa, A., I.B. Mazo, C. Chase, H.N. Shi, H. Liu, Q. Li, H. Rose, H. Leung, B.J. Cherayil, P. Russell, et al. 2005. Perisinusoidal B cells in the bone marrow participate in T-independent responses to blood-borne microbes. *Immunity*. 23:397–407.
- Cariappa, A., L. Chen, K. Haider, M. Tang, E. Nebelitskiy, S.T. Moran, and S. Pillai. 2003. A catalytically inactive form of protein kinase C-associated kinase/receptor interacting protein 4, a protein kinase C beta-associated kinase that mediates NF-kappa B activation, interferes with early B cell development. *J. Immunol.* 171:1875–1880.
- Cariappa, A., C. Boboila, S.T. Moran, H. Liu, H.N. Shi, and S. Pillai. 2007. The recirculating B cell pool contains two functionally distinct, long-lived, posttransitional, follicular B cell populations. *J. Immunol.* 179:2270–2281.
- Cariappa, A., T.J. Kim, and S. Pillai. 1999. Accelerated emigration of B lymphocytes in the Xid mouse. *J. Immunol.* 162:4417–4423.
- Hennet, T., D. Chui, J.C. Paulson, and J.D. Marth. 1998. Immune regulation by the ST6Gal sialyltransferase. *Proc. Natl. Acad. Sci. USA.* 95:4504–4509.

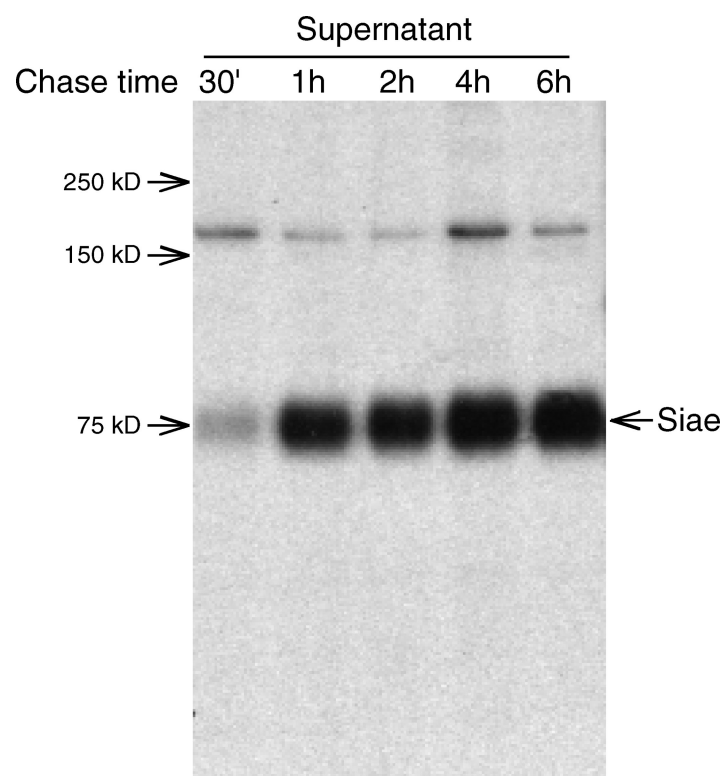


Figure S1. Siae is secreted from stably transfected U2OS cells. U2OS-Siae cells were pulsed with 0.5 mCi of ^{35}S Methionine for 10 min and supernatants were immunoprecipitated immediately after labeling (0') and at the indicated chase time periods. Flag-tagged Siae was identified in cell lysates and in supernatants by immunoprecipitation with anti-Flag antibodies followed by separation on SDS/PAGE and autoradiography.

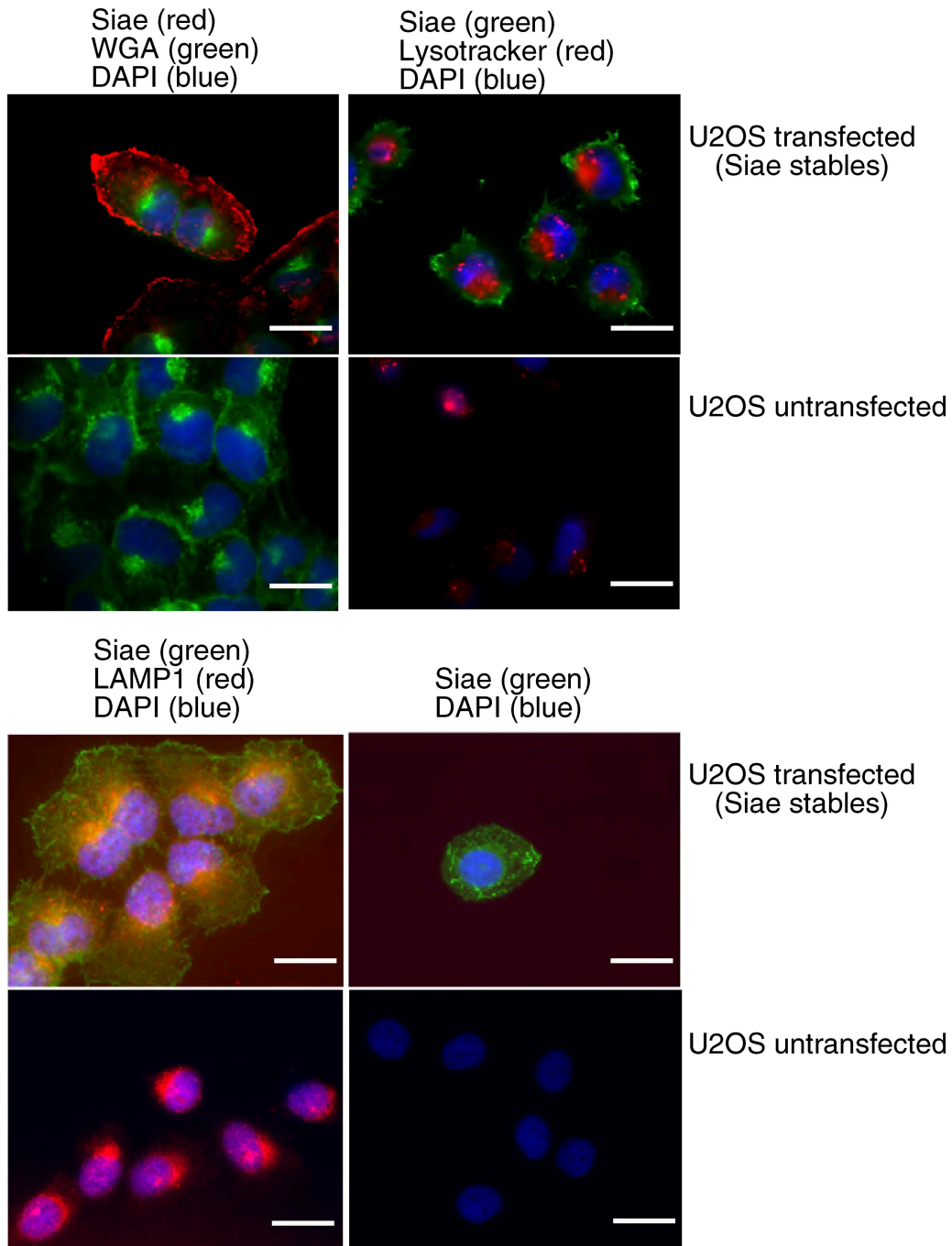


Figure S2. Siae is expressed on the surface of transfected U2OS cells and also partially in lysosomes. U2OS-Siae cells were permeabilized and stained with anti-Flag antibodies (red, top left; and green, top right panels and all bottom), wheat germ agglutinin (WGA; green), LysoTracker (red), or antibodies to LAMP-1 (red). Bars, 20 μ m.

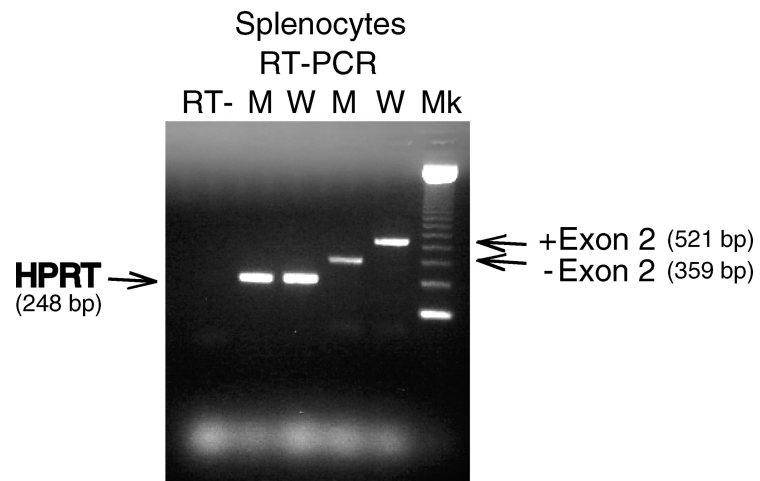


Figure S3. Truncated *Siae* messenger RNA in mutant mice as revealed by RT-PCR. M, mutant; W, WT; Mk, 123-bp marker; RT—, no reverse transcriptase; and HPRT, hypoxanthine phosphoribosyltransferase.

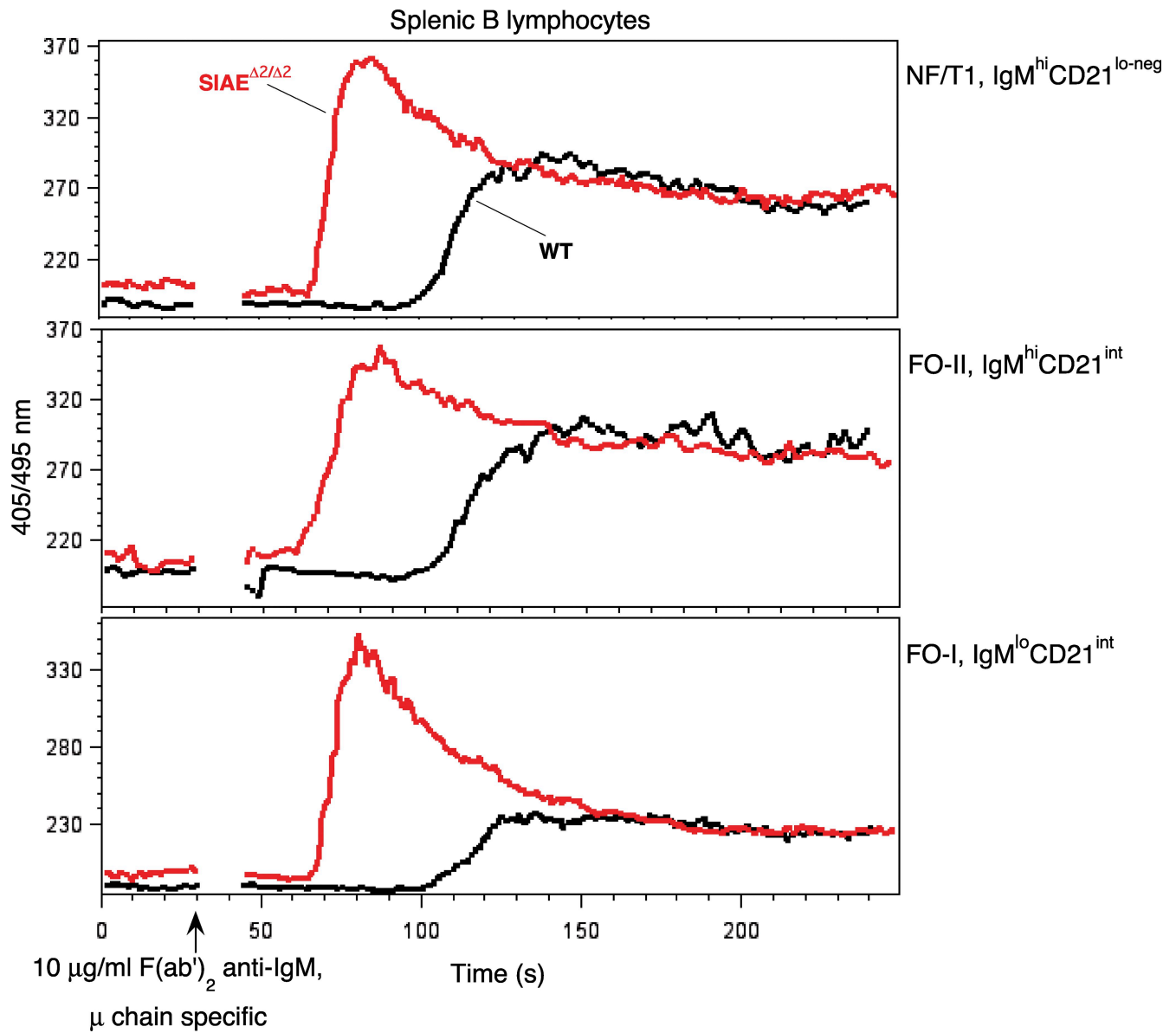


Figure S4. Purified B lymphocytes show accelerated and enhanced BCR signaling in *Siae* mutant mice. B cells were purified from WT and *Siae* mutant mice and intracellular calcium was assessed after BCR cross-linking.

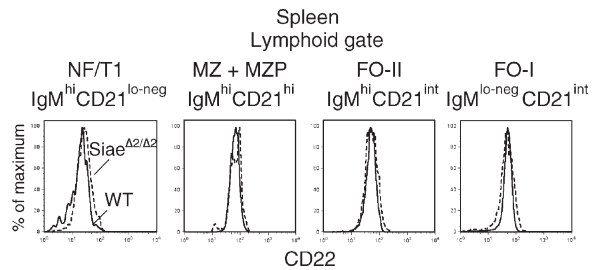


Figure S5. Surface expression of CD22 on splenocytes in *Siae* $\Delta 2/\Delta 2$ mice is unaltered compared to WT mice. Flow cytometric analysis of CD22 expression on WT and mutant B cells was performed. Results are representative of three mice per group.

Splenic B lymphocytes

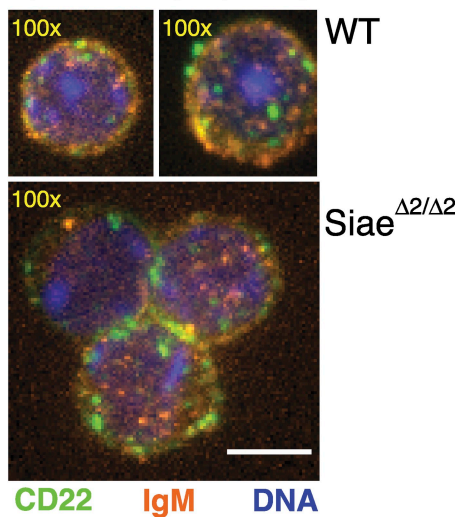


Figure S6. Distribution of CD22 on the surface of unstimulated splenic B lymphocytes is unaltered in *Siae* mutant mice compared to their WT counterparts. B cells were examined using spinning disk confocal microscopy. Bar, 5 μ m. Representative fields are shown.

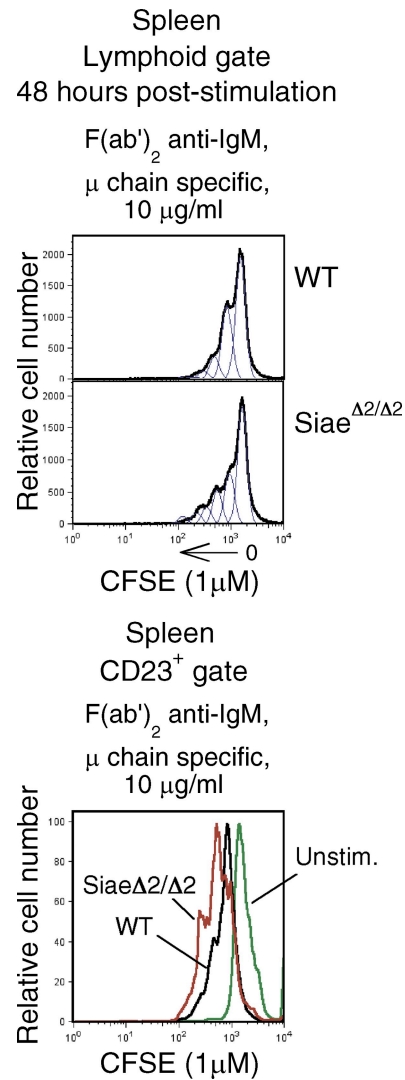
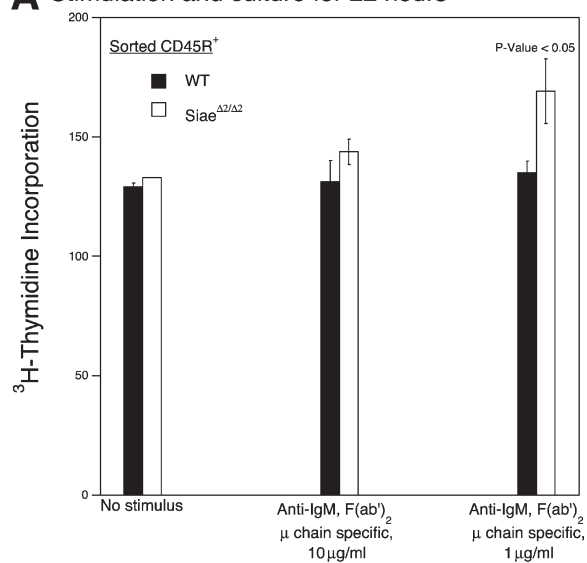
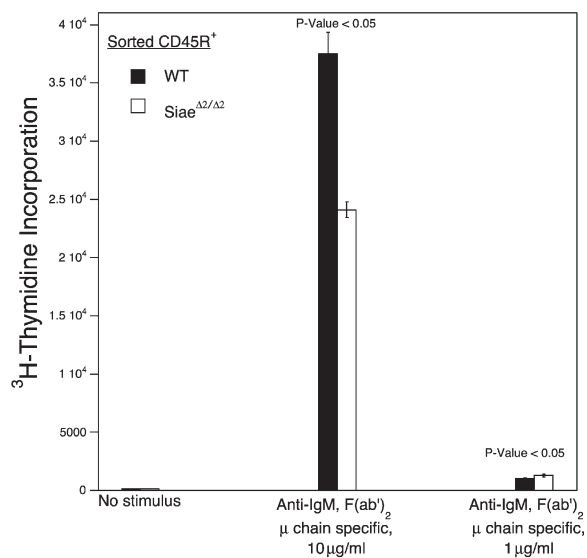


Figure S7. *Siae* mutant splenocytes proliferate more than WT B cells. Generations are marked in blue (top). CD23⁺ cells were gated on (bottom). Results are representative of three mice per group.

A Stimulation and culture for 22 hours



Stimulation and culture for 45 hours



B Stimulation and culture for 45 hours

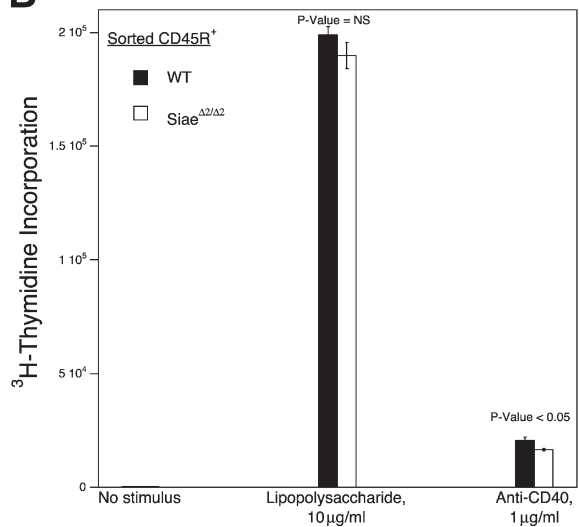


Figure S8. Stimulus strength-dependent inhibition of proliferation in Siae mutant B cells. (A and B) Proliferation after 22 and 45 h (A), and after 45 h (B). $n = 3$ mice per group. Error bars represent SEM.

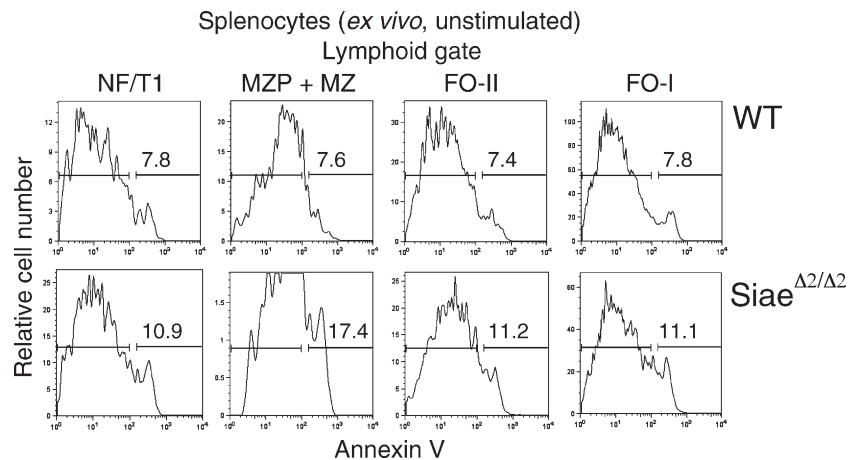


Figure S9. Mutant resting B splenocytes show a mild increase in apoptosis. Annexin V staining was performed on resting B cells from WT and *Siae* mutant mice with three mice per group.

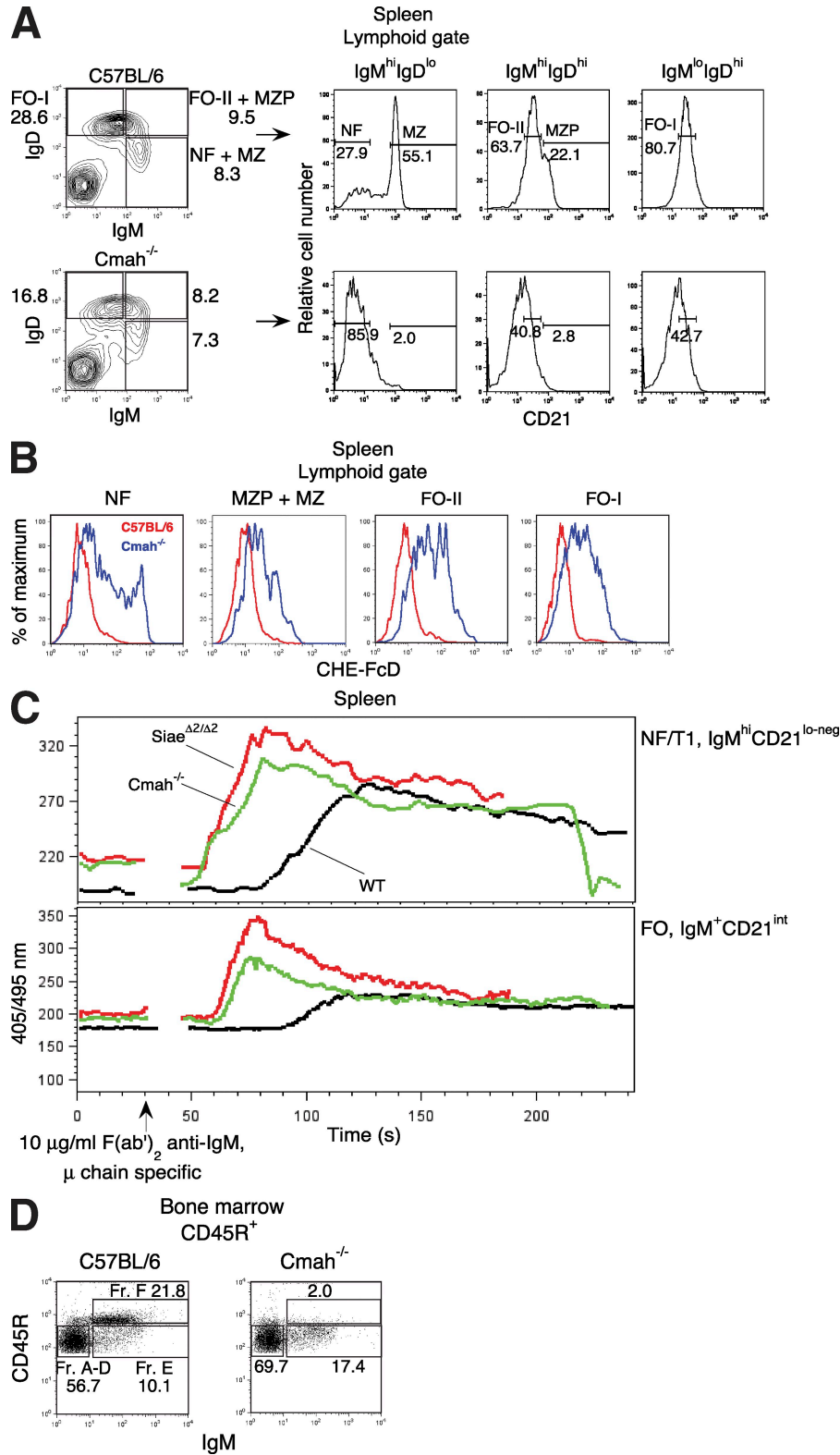


Figure S10. Analysis of *Cmah* mutant mice. (A) Flow cytometric analyses reveal a reduction in MZ B cells in *Cmah*^{-/-} mice. Please refer to Fig. 5B for expansion of abbreviations. Results are representative of more than five mice per group. (B) Increased 9-*O*-acetylation of α2-6-linked sialic acid in *Cmah* mutant B cells. Results are representative of three mice per group. (C) Accelerated and enhanced BCR signaling in *Cmah* mutant mice as assessed by intracellular accumulation of calcium. Results are representative of three mice per group. (D) Perisinusoidal BM B cells are reduced in *Cmah*^{-/-} mice. Results are representative of three mice per group. Please refer to Fig. 5C for expansion of abbreviations.

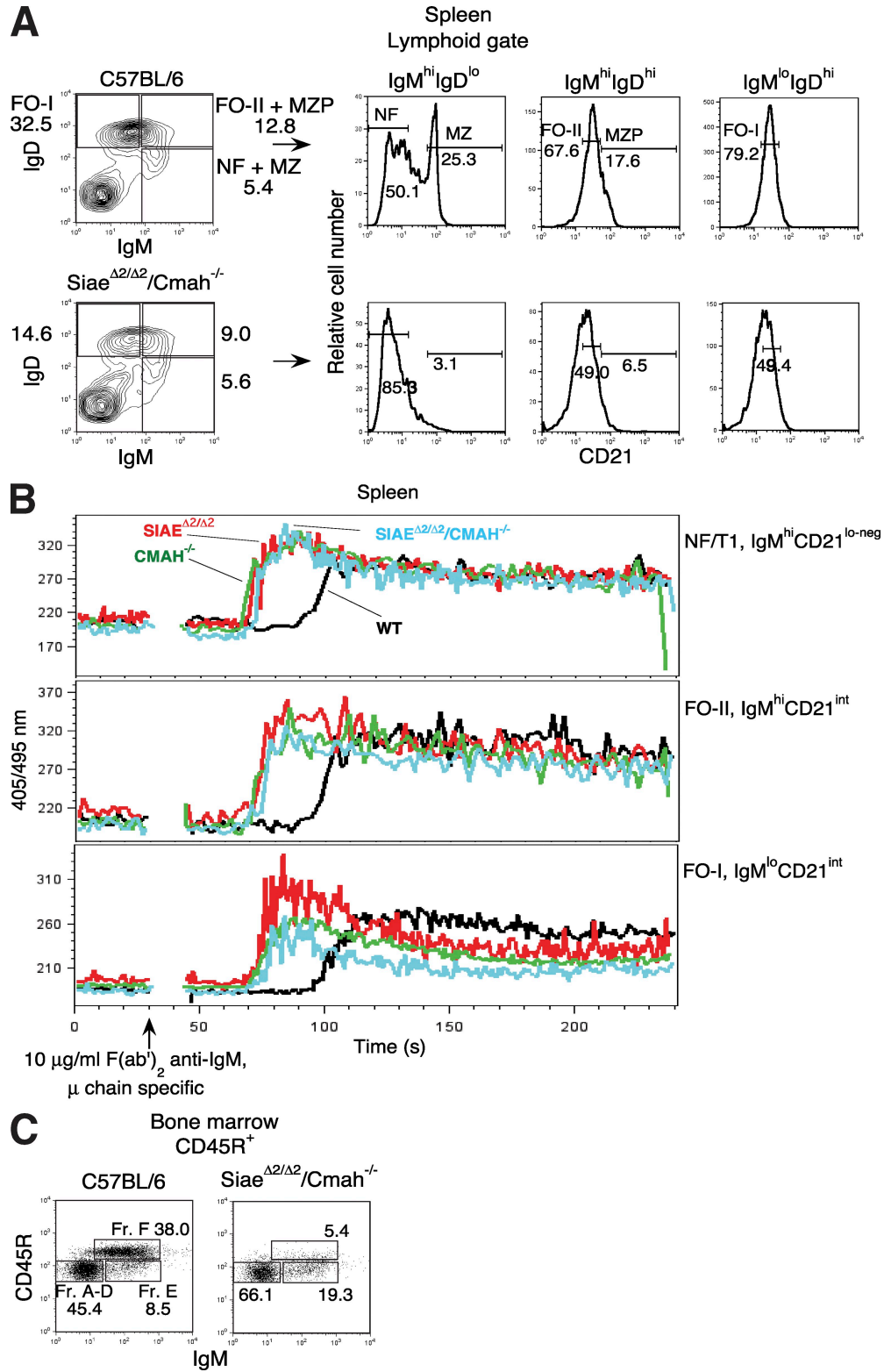


Figure S11. Analysis of *Siae*/*Cmah* double mutant mice. (A) Flow cytometric analyses reveal a reduction in MZ B cells in *Siae*/*Cmah* double mutant mice. Please refer to Fig. 5B for expansion of abbreviations. Results are representative of five mice per group. (B) Accelerated and enhanced BCR signaling in *Siae*/*Cmah* double mutant mice as assessed by intracellular accumulation of calcium. Results are representative of three mice per group. (C) Perisinusoidal BM B cells are reduced in *Siae*/*Cmah* mice double mutant mice. Results are representative of three mice per group. Please refer to Fig. 5C for expansion of abbreviations.

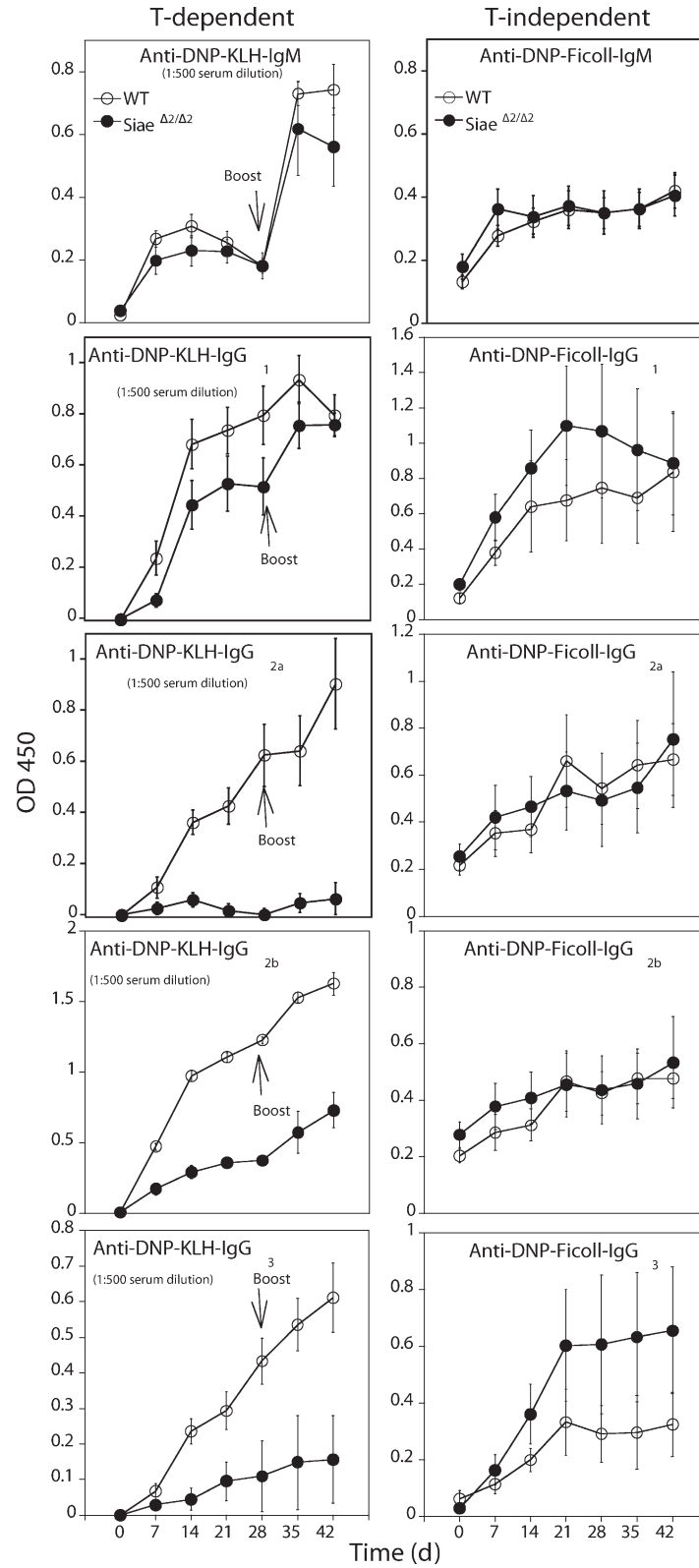


Figure S12. Antibody responses to synthetic T-dependent and T-independent antigens in *SiaE* mutant and WT mice. Five mice were analyzed in each group. Error bars represent SEM.