

Supplemental material

White et al., https://doi.org/10.1083/jcb.201712041

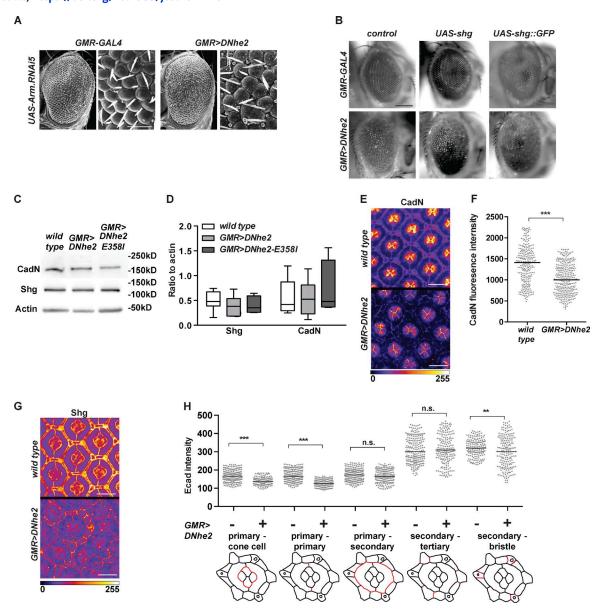


Figure S1. **Overexpression of DNhe2 has no effect on total cadherins and mixed effects on cadherin localization at cell-cell contacts. (A)** Scanning electron micrographs of adult *Drosophila* eyes depicting genetic interactions between *control* (*GMR-GAL4*) and overexpression of *DNhe2* (*GMR>DNhe2*) with RNAi-mediated knockdown of Arm (*UAS-Arm.RNAi5*) using a distinct hairpin sequence. **(B)** Representative images show no suppression of the *GMR>DNhe2* overexpression phenotype with coexpression of two different *UAS-shg* transgenic lines (*UAS-shg* and *UAS-shg::GFP*). **(C)** Representative immunoblots showing total protein levels for Shg (*Drosophila* E-cadherin), CadN, and actin from adult *Drosophila* head lysates made from three *Drosophila* lines: *WT*, overexpression of *DNhe2* (*GMR>DNhe2*), and overexpression of an inactive *DNhe2* mutant (*GMR>DNhe2E358I*). **(D)** Quantitative analysis of replicate Western blots performed as described in C. *WT* and *GMR>DNhe2*, N = 8; *GMR>DNhe2E358I*, N = 4. **(E)** Confocal micrographs of pupal retinae from *WT* and *GMR>DNhe2*-expressing flies labeled for CadN and pseudocolored to show pixel intensities. **(F)** Quantitative measurements of CadN fluorescence intensity (in AU) measured at cone cell-cone cell contacts. *n* = 267–360 junctions, N = 4 individual flies per condition. **(G)** Confocal micrographs of pupal retinae from *WT* and *GMR>DNhe2*-expressing flies labeled for Shg. Bars: 25 μm (A and B); 10 μm (E and G). **(H)** Quantitative measurements of Shg fluorescence intensity (in AU) measured at distinct cell-cell junctions. Labeled schematics show which cell junctions (labeled in red) were measured. *n* = 267–360 junctions, N = 4 individual flies per condition. In D, Tukey boxplots are shown, and significance was determined using an unpaired, two-tailed Student's t test with Holm-Sidak's multiple comparisons correction. In F and H, medians are shown, and significance was determined using the Mann-Whitney test. ***, P < 0.001; ****, P < 0.001.



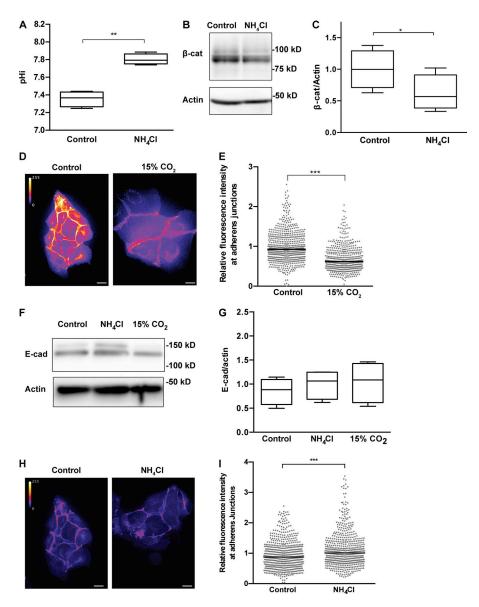


Figure S2. Higher pHi decreases total and junctional β-catenin abundance in clonal NMuMG epithelial cells and MDCK cells; while E-cadherin total levels are unchanged with higher pHi, junctional localization is altered. (A) NMuMG cells maintained for 24 h with 5 mM NH₄Cl have a higher pHi than control untreated cells. n = 4. (B) Representative immunoblots showing total protein levels for β-catenin from NMuMG cells grown under conditions as described in A. (C) Quantitative analysis of replicate Western blots performed as described in B with β-catenin intensity normalized to actin for each replicate. n = 4. (D) Confocal images of MDCK cells grown under normal conditions (control) or acclimated to 15% CO₂ for two passages, fixed and immunolabeled for β-catenin, and pseudocolored to show fluorescence intensities. (E) Quantitative measurements of β-catenin fluorescence intensity at adherens junctions under conditions described in D. n = 3, n = 508-631 junctions. (F) Representative immunoblots showing total protein levels for E-cadherin from MDCK cells grown under normal conditions (control), acute 24-h treatment with 5 mM NH₄Cl, or acclimated to 15% CO₂ for two passages. (G) Quantitative analysis of immunoblots with E-cadherin intensity normalized to actin for each replicate. Control and NH₄Cl, n = 6; 15% CO₂, n = 4. (H) Confocal images of MDCK cells grown under conditions described in F, fixed and immunolabeled for E-cadherin, and pseudocolored to show fluorescence intensities. Bars, 20 μm. (I) Quantitative measurements of images collected as described in H. Data were normalized to the mean fluorescence intensity in control cells for each replicate. Control and NH₄Cl, n = 3, n = 581-617 junctions. In A, C, and G, Tukey boxplots are shown, and significance was determined by unpaired, two-tailed Student's t tests. In E and I, medians are shown, and significance was determined using the Mann–Whitney test. *, P < 0.005; ** , P < 0.01; ***, P < 0.001.



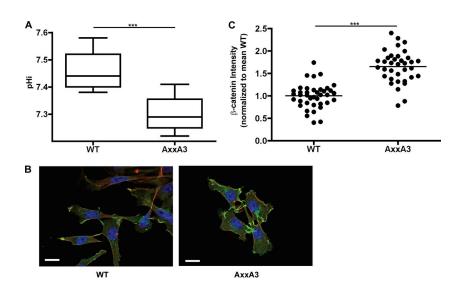


Figure S3. Lower pHi increases β-catenin abundance at membrane protrusions. (A) pHi measurements in PS120 cells stably expressing WT Nhe1 (WT) or mutant Nhe1 (AxxA3). n = 4. (B) Representative immunofluorescence images of cells treated as described in A showing β-catenin– (green), phalloidin-(red), and DAPI-stained nuclei (blue). Bars, 20 μm. (C) Quantitative analysis of β-catenin intensity at membrane protrusions. n = 3. In A, Tukey boxplots are shown; in C, means are shown; in A and C, significance was determined by unpaired, two-tailed Student's t tests. ***, P < 0.001.



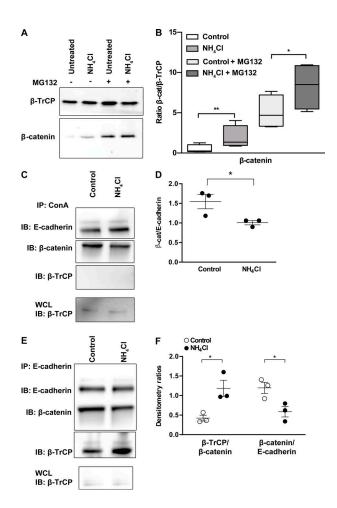


Figure S4. Higher pHi increases association of β-catenin with the E3 ligase β-TrCP and increases the amount of β-TrCP associating with cadherinassociated β-catenin. (A) Representative immunoblots (IBs) of β-catenin and β-TrCP abundance in endogenous β-TrCP immune complexes (IP) from MDCK cells untreated (control) or treated with 5 mM NH₄Cl for 24 h in the absence and presence of proteasome inhibitor MG132 (see Materials and methods for details). (B) Quantitative analysis of immunoblot replicates from A. n = 4. (C) Representative immunoblots of E-cadherin, β-catenin, and β-TrCP abundance in ConA precipitations from MDCK cells untreated (control) or treated with 5 mM NH₄Cl for 24 h. Immunoblot of β-TrCP in whole-cell lysates (WCLs) shows overall abundance. (D) Quantitative analysis of immunoblot replicates from C. n = 3. (E) Representative immunoblots of E-cadherin, β-catenin, and β-TrCP abundance in endogenous E-cadherin immune complexes (IP) from MDCK cells untreated (control) or treated with 5 mM NH₄Cl. Immunoblot of β-TrCP in whole-cell lysates shows overall abundance. (F) Quantitative analysis of immunoblot replicates from E. n = 3. In B, Tukey boxplots are shown. In D and F, scatterplots and means are shown. In B, D, and F, significance was determined using an unpaired, two-tailed Student's t test. *, P < 0.05; **, P < 0.01.

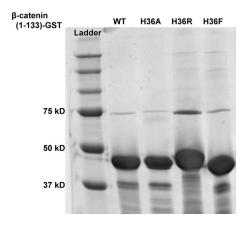


Figure S5. Coomassie of purified β-catenin-(1-133)-GST proteins.