

John Peter et al., <https://doi.org/10.1083/jcb.201610055>

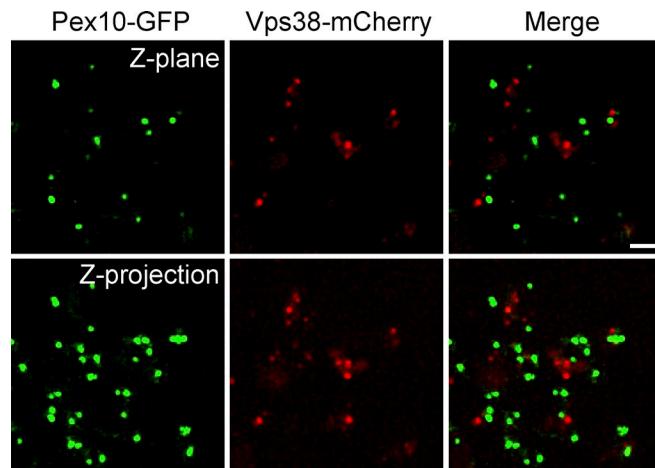


Figure S1. **Pex10 and Vps38 do not colocalize.** Pex10 and Vps38 are C-terminally tagged at their endogenous loci with GFP and mCherry, respectively. Bar, 2 μ m.

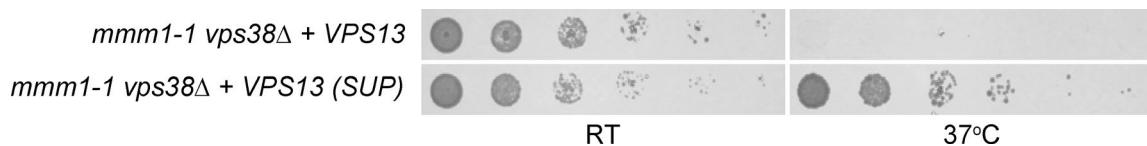


Figure S2. **Vps13 suppressor rescues loss of ERMES function in the absence of Vps38.** The ts strain *mmm1-1* with the indicated genotypes was spotted as serial dilutions on minimal media plates and grown at RT and 37°C. Both WT Vps13 and the suppressor version are expressed from a low-copy plasmid.

Table S1. Yeast strains used in the study

Strain	Genotype	Reference
ByK45	BY4741 MAT α his3Δ leu2Δ0 met15Δ0 ura3Δ0	Euroscarf
ByK46	BY4742 MAT α his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0	Euroscarf
ByK498	ByK45 VPS13 ⁺ GFP	Lang et al., 2015
ByK486	ByK45 VPS13 ⁺ GFP vps38Δ::KANMX	This study
ByK495	ByK45 VPS13 ⁺ GFP VPS38::mCHERRY-HIS	This study
ByK497	ByK45 VPS13 ⁺ GFP(L1627S) VPS38::mCHERRY-HIS	This study
ByK566	ByK45 VPS13 ⁺ GFP PEX10::mCHERRY-NATNT2	This study
ByK567	ByK45 VPS13 ⁺ GFP(L1627S) PEX10::mCHERRY-NATNT2	This study
ByK499	ByK45 VPS13 ⁺ GFP VPS39::KANMX-GPDpr-VPS39	This study
ByK393	ByK45 VPS13 ⁺ GFP MCP1::NATNT2-GPDpr-MCP1	This study
ByK226	YH8 ura3-167 trp1-A1 leu2-A1 his3-A200 mmm1-1	This study
ByK394	ByK226 mmm1-1 MCP1::NATNT2-GPDpr-MCP1	This study
ByK258	ByK226 mmm1-1 vps13Δ::HIS	This study
ByK395	ByK226 mmm1-1 vps13Δ::HIS MCP1::NATNT2-GPDpr-MCP1	This study
ByK405	ByK226 mmm1-1 MCP1::NATNT2-GPDpr-MCP1 vps39Δ::KANMX	This study
ByK539	ByK226 mmm1-1 MCP1::NATNT2-GPDpr-MCP1 vps38Δ::KANMX	This study
ByK627	ByK226 mmm1-1 MCP1::NATNT2-GPDpr-MCP1 pex1Δ::KANMX	This study
ByK638	ByK226 mmm1-1 VPS39::NATNT2-GPDpr-VPS39	This study
ByK636	ByK226 mmm1-1 vps13Δ::HIS VPS39::NATNT2-GPDpr-VPS39	This study
ByK676	ByK226 mmm1-1 vps38Δ::KANMX	This study
ByK447	ByK45 VPS39::GFP-KANMX	This study
ByK392	ByK45 MCP1::NATNT2-GPDpr-MCP1	This study
ByK448	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS39::GFP-KANMX	This study
ByK536	ByK45 MCP1::6xHA-HPHNT1	This study
ByK537	ByK45 VPS13 ⁺ GFP MCP1::6xHA-HPHNT1	This study
ByK538	ByK45 VPS13 ⁺ GFP(L1627S) MCP1::6xHA-HPHNT1	This study
ByK433	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS13::GFP-KANMX	This study
ByK424	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS13(1-2766aa)::GFP-KANMX	This study
ByK462	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS13(1-2599aa)::GFP-KANMX	This study
ByK460	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS13(1-2447aa)::GFP-KANMX	This study
ByK458	ByK45 MCP1::NATNT2-GPDpr-MCP1 VPS13(1-2247aa)::GFP-KANMX	This study
ByK413	ByK45 VPS13::NATNT2-CYC1pr-yeGFP-VPS13	This study
ByK469	ByK45 VPS13::NATNT2-CYC1pr-yeGFP-VPS13 (2247-3144aa)	This study
ByK655	ByK45 MCP1::KANMX-GPDpr-MCP1 VPS13::NATNT2-CYC1pr-yeGFP- VPS13	This study
ByK470	ByK45 MCP1::KANMX-GPDpr-MCP1 VPS13::NATNT2-CYC1pr-yeGFP-VPS13 (2247-3144aa)	This study
ByK719	ByK499 MCP1::NATNT2-GPDpr-MCP1	This study
ByK720	ByK719 VPS39::mCHERRY-HIS3MX	This study
ByK324	ByK45 VPS39::NATNT2-GPDpr-GFP	This study
ByK325	ByK324 vps13Δ::KANMX	This study
ByK660	ByK46 x ByK324 (diploid)	This study
ByK661	ByK660 mdm10Δ::KANMX	This study
ByK630	ByK226 mmm1-1 mcp1Δ::KANMX	This study

Table S2. Plasmids used in the study

Plasmid	Genotype	Reference
pBK417	pRS413-TEFpr-mCHERRY-UBC6(TA)	This study
pBK250	pRS413-TEFpr-MCP1(1-61)-mCHERRY-UBC6(TA)	This study
pBK251	pRS413-TEFpr-MCP1(122-173)-mCHERRY-UBC6(TA)	This study
pBK416	pRS413-TEFpr-mCHERRY-FIS1(TA)	This study
pBK415	pRS413-TEFpr-MCP1(1-61)-mCHERRY-FIS1(TA)	This study
pBK336	pVPS13 ⁺ GFP	This study
pBK239	pVPS13 ⁺ GFP (L1627S)	This study
pBK509	pRS415-GPDpr-MCP1(WT)	This study
pBK510	pRS415-GPDpr-MCP1(H187A)	This study
pBK511	pRS415-GPDpr-MCP1(P195A)	This study
pBK512	pRS415-GPDpr-MCP1(H237A)	This study
pBK513	pRS415-GPDpr-MCP1(G241A)	This study

Table S3. Primers used in the study

Number	Name	Sequence
Primers for genomic tagging		
1	Vps38_F1_pringle	5'-TGATGGTTTACCTATTAGGGATAGTAATCATAATTAAAAATATCGGATCCCCGGTTAATTAA-3'
2	Vps38_R1_pringle	5'-GAAAAGATTAATGGCAGTCAAAAGAGATTGATTTCACTGAGCTAGAATTGAGCTCGTTAAC-3'
3	Vps38_F2_pringle	5'-CTGATATATTGCAATTAAACAGATTCTGGAGAATTACCGTCAACACGGATCCCCGGTTAATTAA-3'
4	Pex10_F2_pringle	5'-GTGCAGACAACACTGTCAACCACAGGAATTCTGGTCTCGGGCAACGGATCCCCGGTTAATTAA-3'
5	Pex10_R1_pringle	5'-GTGACAATGCTAAAAGAGTAGTCAAATTATTGATTAGTCTCTAGAATTGAGCTCGTTAAC-3'
6	Vps39_F1_pringle	5'-AGAAAAACCCCTCAAATATCAATTATAACAAAATTACGGATCCCCGGTTAATTAA-3'
7	Vps39_R1_pringle	5'-TAAGAAATACTAACACAATAACAGCAGCTTAAGGGATAATTGAGCTCGTTAAC-3'
8	Vps39_F2_pringle	5'-AGAATTAAACAGAAGACTTGGAGAAGTTATAATGAGCTAATAATAACGGATCCCCGGTTAATTAA-3'
9	Mcp1_S1_knop	5'-GAATTACTAGGGCATATACCAAGTTAGCCCAGAGTTGTTTACGATGCGTACGCTGAGTCGAC-3'
10	Mcp1_S4_knop	5'-GAGACTAGCAGGTCTACTGGTTCTGGAGGCACTTCATGCAACTTATCATGATGAATTCTCTGTCG-3'
11	Mcp1_S3_knop	5'-CAATTGAAACCATTTCAAAAGATCGGTTGCTTGCAGCTGAATCGTACGCTGAGTCGAC-3'
12	Mcp1_S2_knop	5'-GAATTGTCGCAAATTAACTAGCTTACACTACTCCTACAAGGATTCTAATCGATGAATTGAGCTCG-3'
13	Vps13_pringle_F1	5'-AAAGGAAAAGGCAGAAAAAGAAAATTAAAGAACAGTTACGGATCCCCGGTTAATTAA-3'
14	Vps13_pringle_F2	5'-CTACATAGTGACAAAGCGGTATATACATTCTATGTAAGTCTGAGCTCGTTAAC-3'
15	Pex1_F1_pringle	5'-GAAGGAGCGCAGTAACAAGAACACCTGAGGAACCTGCTTCAAATGGGATCCCCGGTTAATTAA-3'
16	Pex1_R1_pringle	5'-TGCCCTTAAAGGAAACGCCCTTGTCTTCTCTCTTCAAGGTTGAGTCGAC-3'
17	Vps13_S3_cknop_2247aa	5'-GCACAACTCCAAGATTGCTGAAAAGCAGAAAAAGATATAACTATTCCACGTAACGCTGAGTCGAC-3'
18	Vps13_S3_cknop_2447aa	5'-CCATGACTTATATTACCAAGAGATCTCGTAAAGTTAGTGTACCCACGCTACGCTGAGTCGAC-3'
19	Vps13_S3_cknop_2599aa	5'-CACAGATAGTTGTTCGTTCAAAGGAGTTGATCTCATTAATCAATGGTGTACGCTGAGTCGAC-3'
20	Vps13_S3_cknop_2766aa	5'-CATAGTACAATGATGACGAAATGACTCCTCGGTCTCTCAGCCAATCCATCGATGAATTCTCTGTCG-3'
21	Vps13_S3_cknop	5'-CGCCATTGCTTTAGAGAATACAATAAGTACTGTGAAGCTACCTACGTCAGCTGAGTCGAC-3'
22	Vps13_S2_cknop	5'-ATAGCTACATAGTGACAAAGCGGTATATACATTCTATGATCAATCGATGAATTGAGCTCG-3'
23	Vps13_S1_knop	5'-GTGACTACAAAAGGAAAGGCAGAAAAAGAAAATTAAAGAACAGTTAACGCTACGCTGAGTCGAC-3'
24	Vps13_S4_knop	5'-CCACATATGAGCTAGCAATGGTTAGCAAATTAGCAGCTAAAGACTCTAACATCGATGAATTCTCTGTCG-3'
25	Vps13_S4_cknop_2247	5'-ACGAGCTCTGTTACTTTATCATCTCTTATCAAATGAAAACATTTCATCGATGAATTCTCTGTCG-3'
26	Vps39_F2_pringle	5'-AGAATTAAACAGAAGACTTGGAGAAGTTATAATGAGCTAATAATAACGGATCCCCGGTTAATTAA-3'
27	Vps39_R1_pringle	5'-TAAGAAATACTAACACAATAACAGCAGCTTAAGGGATAATTGAGCTCGTTAAC-3'
Primers used for cloning		
17	mcp1_1-61_ov1TEF_f	5'-CAATCTAATCTAAGTTCTAGAACTAGTGGATCCATGATAAGTTGATGAAGTGC-3'
18	mcp1_1-61_ov1mCh_r	5'-CACCATGTTAATTAAACACGACCCGTCACCGATGTCAGCGAAATCTTGTACAGTTCA-3'
19	mcp1_122-173_ov1TEF_f	5'-CATAGCAATCTAATCTAAGTTCTAGAACTAGTGGATCCGAATCGTGAACAACTGGAAC-3'
20	mcp1_122-173_ov1mCh_r	5'-CCATGTTAATTAAACACGACCCGTCACCGATGTCAGCTGACTTGGAGGAATGCGGAAG-3'
21	TEFov1_mChATG_f	5'-TAATCTAAGTTCTAGAACTAGTGGATCCATGTCGAGGTGCTGGTT-3'
22	Xhol_Ubc6TA_mCh_rv	5'-GACCCCTGAGTCATTTCTAAAGGCAACCAAAACAAAAATAGCGATACCAATATAACCATTGAAGAACTCTGTACAGCTCGCCATG-3'
23	mCh_Fis1TM_Xhol_r	5'-GACCCCTGAGTCATTTCTAAAGGCAACCAAAACAAAAATAGCGATACCAATATAACCATTGAAGAACTCTGTACAGCTCGCCATG-3'
24	GPD_MCP1_gaprep_f	5'-TTTTAGTTAAACACCAAGAACTAGTTGCAATGATAAGTTGATGAAGTGCCTCC-3'
25	pRS4 × 5_MCP1_gaprep_r	5'-GCCGCCCTCGAGGTCACGGTATCGATAAGCTGCTAATTACGTCACAGCAACC-3'

Provided online are four alignments in one PDF and three scripts in one Word file. Alignments S1 and S2 show homology search results for Vps13 (1-3,000 aa) and Vps13 (1,501-3,144 aa), searched against the Pfam database and yeast proteome, respectively. Alignments S3 and S4 show the results for the C terminus of Vps13 (3,030-3,144 aa) and full-length Mcp1, respectively, searched against the Protein Data Bank. All homology alignments were generated using the HHpred server. Scripts S1, S2, and S3 are ImageJ macros used to calculate the percentage of colocalization between Vps13[^]GFP/Pex10-mCherry foci, Vps13[^]GFP/Vps38-mCherry foci, and Pex10-GFP/Vps38-mCherry foci, respectively.

Reference

- Lang, A.B., A.T. John Peter, P. Walter, and B. Kornmann. 2015. ER-mitochondrial junctions can be bypassed by dominant mutations in the endosomal protein Vps13. *J. Cell Biol.* 210:883–890. <http://dx.doi.org/10.1083/jcb.201502105>