Feiler et al., http://www.jcb.org/cgi/content/full/jcb.201504057/DC1

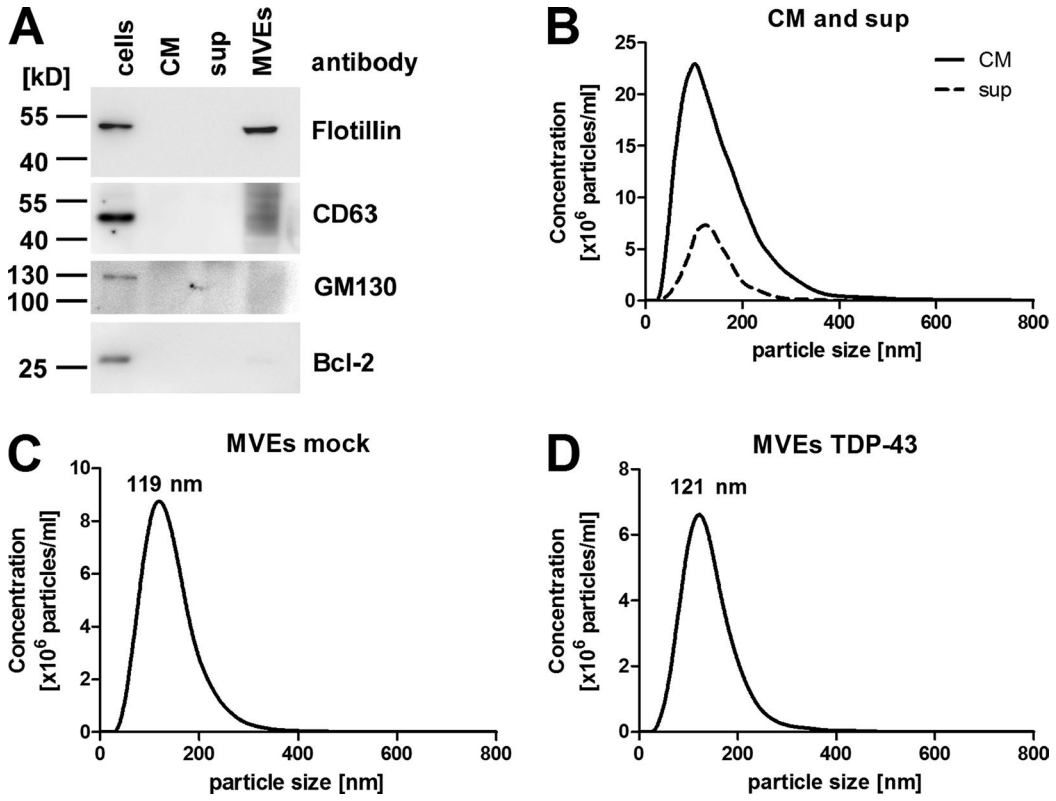

Figure S1. **Characterization of HEK-293–derived MVEs.** (A) Immunoblotting of HEK-293 cells, their corresponding CM directly before ultracentrifugation, MVE-free supernatant (sup) after ultracentrifugation, and MVEs as a proof for MVE purity. 10 µg of total protein of lysate and 30 µg of CM, supernatant, and MVEs were loaded. Flotillin and CD63 were used as an MVE marker, whereas Bcl-2 and GM130 were used as a marker for cellular contaminations. (B–D) Size distribution analysis of microvesicular/exosomal fractions isolated from CM of HEK-293 cells transfected with either mock or myc–TDP-43 by NanoSight particle tracking. (B) CM directly before ultracentrifugation and MVE-free supernatant (sup) after ultracentrifugation. (C and D) MVE fraction of mock- (C) or myc–TDP-43–transfected (D) HEK-293 cells. Mean values from $n = 3$ replicates are shown.

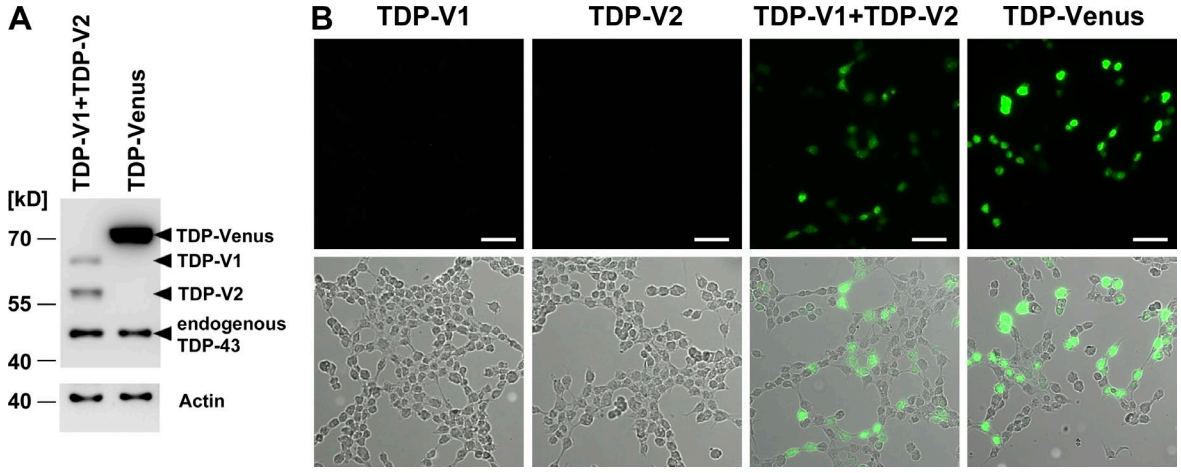

Figure S2. **Characterization of the TDP-43 split Venus and the TDP-Venus constructs.** (A) Western blot analysis of TDP-V1 and -V2 coexpression or TDP-Venus expression in HEK-293 cell lysates 48 h after transfection. (B) Representative micrographs of HEK-293 cells either expressing TDP-V1, TDP-V2, TDP-V1 and TDP-V2, or TDP-Venus. Micrographs were taken 18 h after transfection. Bars, 50 µm.