

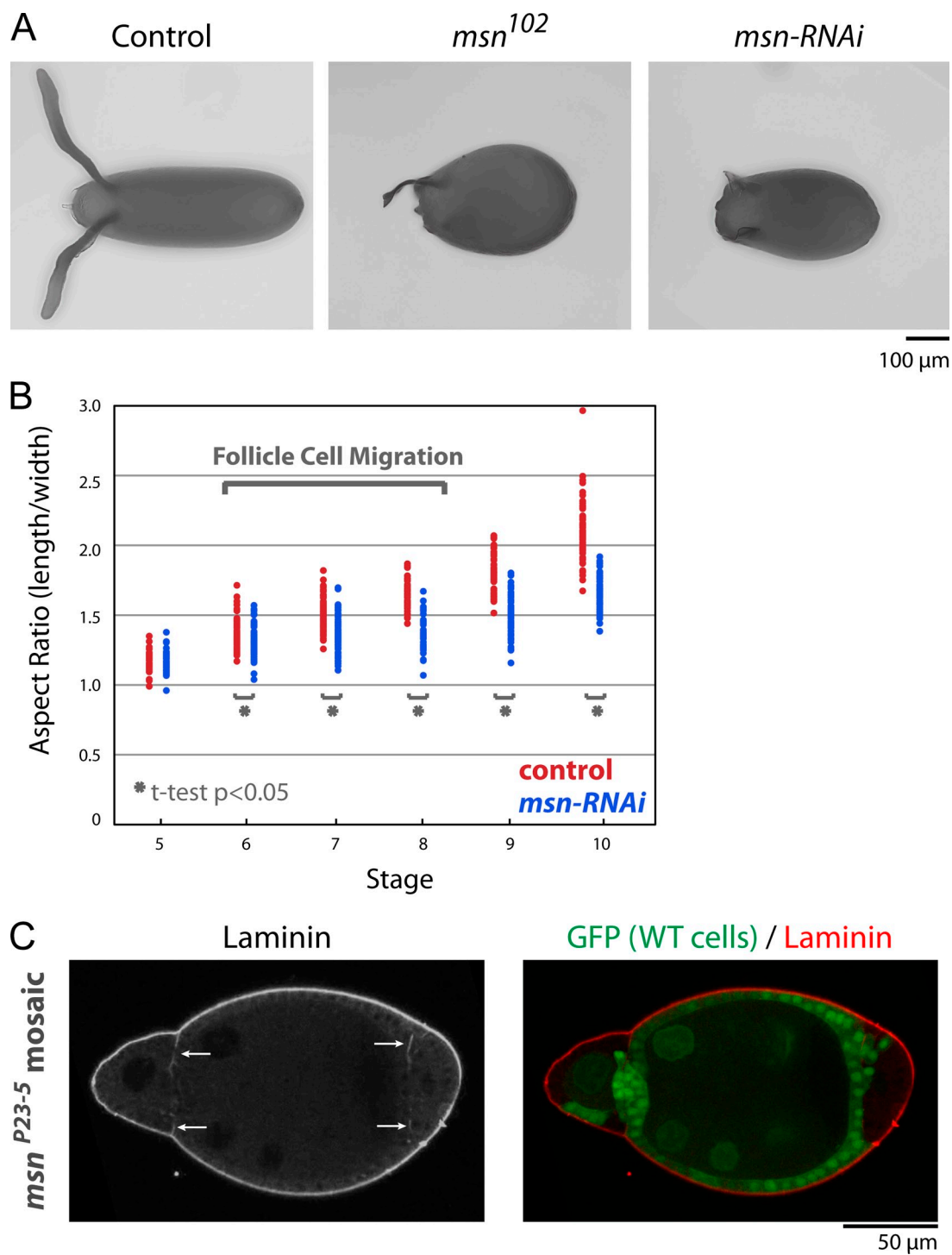
Lewellyn et al., <http://www.jcb.org/cgi/content/full/jcb.201209129/DC1>

Figure S1. **Misshapen is required for egg chamber elongation.** (A) Expression of *msn*-RNAi in the follicle cells produces round eggs, similar to follicle cell mosaics of a published allele, *msn*¹⁰². The wild-type image has been replicated from Horne-Badovinac et al. (2012). (B) Aspect ratios for individual control and *msn*-RNAi egg chambers are plotted for each developmental stage. *msn*-RNAi egg chambers are significantly rounder than controls starting at stage 6. These data are from a single experiment that involved dissection of >12 flies per condition, with $n = 35$ –80 egg chambers for each condition at each developmental stage. (C) Transverse optical section of an *msn* mosaic egg chamber that has mutant clones (indicated by lack of GFP) at the anterior and posterior poles, causing wild-type follicle cells to invade the germ cells. Laminin staining shows that invading wild-type cells maintain contact with a BM (arrows). Bar measurements are indicated in the figure.

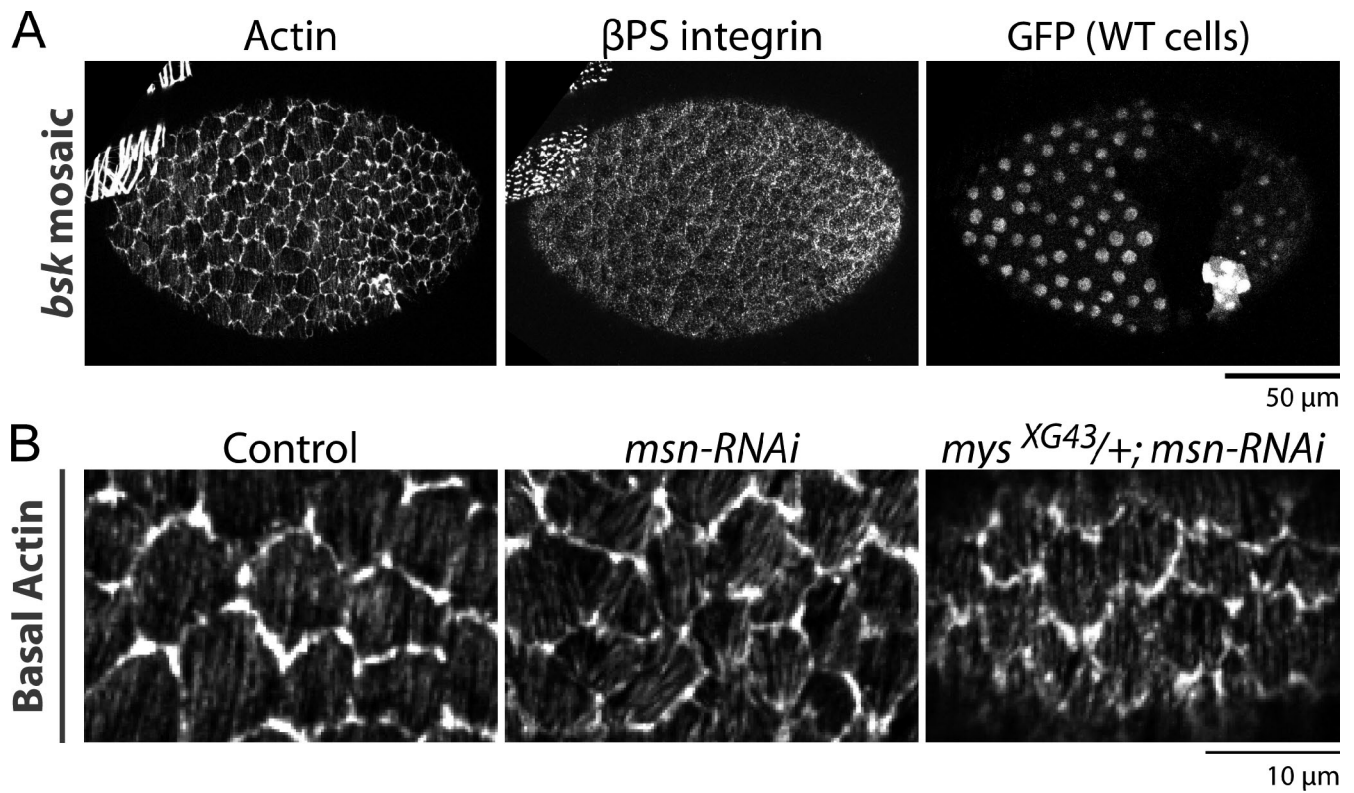
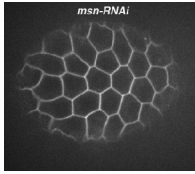
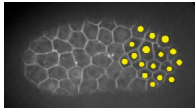


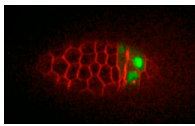
Figure S2. **Msn's effect on integrin levels is JNK independent, and reducing integrin levels rescues the basal actin organization defect in *msn-RNAi*.** (A) A basal view of a *Df(bsk)^{fbp147E}* mosaic epithelium stained for the β -PS integrin reveals that defects in JNK signaling do not affect integrin levels. (B) Reducing integrin levels in *msn-RNAi* epithelia restores the planar organization of the basal actin filaments (only 5% show disruptions in basal actin organization compared with 20% in *msn-RNAi* alone, $n = 20$ for control and *msn-RNAi*. $n = 22$ for *mys*^{XG43/+}; *msn-RNAi*).



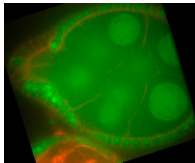
Video 1. **Msn promotes follicle cell migration.** Live imaging of control and *msn-RNAi* egg chambers (Fig. 1 C). Samples were analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Cell membranes are labeled with FM4-64. Frames were taken every 30 s for 22.5 min. Playback rate is 300x real time.



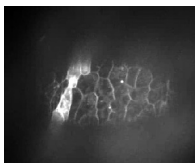
Video 2. **A large *msn* mutant clone blocks global follicle cell migration.** *msn*¹⁷² cells (yellow dots) are stationary, while neighboring wild-type cells attempt to move past them (Fig. 1 D). The *msn* mosaic egg chamber was analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Cell membranes are labeled with FM4-64. Frames were taken every 30 s for 61 min. Playback rate is 300x real time.



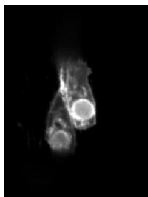
Video 3. ***msn-RNAi* mosaic egg chambers exhibit a cell-autonomous defect in follicle cell migration.** The cells in an *msn-RNAi* flipout clone (GFP) move very little while neighboring wild-type cells efficiently migrate past them (Fig. 1 F). The mosaic egg chamber was analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Cell membranes are labeled with FM4-64 (red). Frames were taken every 30 s for 32 min. Playback rate is 300x real time.



Video 4. **The presence of an *msn*¹⁷² mutant clone at the anterior leads to invasion of neighboring wild-type cells (GFP) into the germ cell cluster.** The time-lapse begins at a transverse section, then goes up to a more basal plane before returning back to the transverse section (Fig. 2, D and E). The mosaic egg chamber was analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Cell membranes are labeled with FM4-64 (red). Frames were taken every 30 s for 90 min. Playback rate is 300x real time.



Video 5. **“Tails” form at the back of individual *msn-RNAi* cells in a small clone (mCD8-GFP, Fig. 5 B).** An egg chamber containing a clone of cells expressing *UAS-msn-RNAi* and *UAS-mCD8-GFP* was analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Frames were taken every 30 s for 28 min. Playback rate is 300x real time.



Video 6. **“Tails” can retract as the *msn-RNAi* follicle cells migrate forward (mCD8-GFP, Fig. 5 C).** An egg chamber containing a clone of cells expressing *UAS-msn-RNAi* and *UAS-mCD8-GFP* was analyzed by time-lapse microscopy using a spinning disk confocal microscope (Axiovert 200M; Carl Zeiss) with a spinning disk unit (model CSU-10; Yokogawa Corporation of America). Frames were taken every 30 s for 60 min. Playback rate is 300x real time.

Table S1. Experimental genotypes

Figure	Panel	Genotype
1	C and D	<i>w¹¹¹⁸</i> / +; <i>TubP-GAL80^{ts}</i> / +; <i>TubP-GAL4</i> / + <i>TubP-GAL80^{ts}</i> / UAS- <i>msn</i> -RNAi; <i>TubP-GAL4</i> / +
1	E	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁷²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i>
1	F	<i>hsFLP</i> / +; UAS- <i>msn</i> -RNAi / +; <i>act5c>>GAL4</i> , UAS-GFP / +
2	A	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁷²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> (top, no clones) <i>e22c-GAL4</i> , UAS-FLP / +; <i>msn^{P23-5}</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> (bottom, clone)
2	B	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁰²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i>
2	C	<i>C306-GAL4</i> / +; UAS- <i>msn</i> -RNAi / +; UAS- <i>mCD8-GFP</i> / +
2	D and E	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁷²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i>
3	A	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁰²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> (top) <i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁷²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> (bottom)
3	B	<i>hsFLP</i> / +; UAS- <i>HA-myr-msn</i> / <i>act5c>>GAL4</i> , UAS-GFP
3	C	<i>w¹¹¹⁸</i> / +; <i>tj-Gal4</i> , <i>vkG-GFP</i> / + (top left) <i>vkG-GFP</i> / <i>e22c-GAL4</i> , UAS-FLP; <i>msn¹⁰²</i> , <i>FRT80</i> / <i>hs-piMyc</i> , <i>FRT80</i> (top right) <i>tj-Gal4</i> , <i>vkG-GFP</i> / UAS- <i>msn</i> -RNAi (bottom left) <i>mys^{XG43}</i> / +; <i>tj-GAL4</i> , <i>vkG-GFP</i> / UAS- <i>msn</i> -RNAi (bottom right)
3	D	<i>w¹¹¹⁸</i> / +; <i>tj-Gal4</i> / + <i>mys^{XG43}</i> / +; <i>tj-GAL4</i> / + <i>tj-Gal4</i> / UAS- <i>msn</i> -RNAi <i>mys^{XG43}</i> / +; <i>tj-GAL4</i> / UAS- <i>msn</i> -RNAi <i>tj-Gal4</i> / UAS- <i>msn</i>
4	A and B	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn-YFP</i> , <i>FRT80</i> / <i>FRT80</i>
4	C	<i>w¹¹¹⁸</i> (left) <i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁰²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> (right)
5	A	<i>w¹¹¹⁸</i> / <i>hsFLP</i> ; <i>act5c>>GAL4</i> , UAS-GFP / + <i>hsFLP</i> / +; UAS- <i>msn</i> -RNAi / +; <i>act5c>>GAL4</i> , UAS-GFP / +
5	B and C	<i>hsFLP</i> / +; UAS- <i>msn</i> -RNAi / +; <i>act5c>>GAL4</i> , UAS-GFP / UAS- <i>mCD8-GFP</i>
5	D	<i>hsFLP</i> / +; UAS- <i>msn</i> -RNAi / +; <i>act5c>>GAL4</i> , UAS-RFP / <i>tal</i> -GFP
S1	A	<i>w¹¹¹⁸</i> / +; <i>TubP-GAL80^{ts}</i> / +; <i>TubP-GAL4</i> / + <i>e22c-GAL4</i> , UAS-FLP / +; <i>msn¹⁰²</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i> <i>TubP-GAL80^{ts}</i> / UAS- <i>msn</i> -RNAi; <i>TubP-GAL4</i> / +
S1	B	<i>w¹¹¹⁸</i> / +; <i>TubP-GAL80^{ts}</i> / +; <i>TubP-GAL4</i> / + <i>TubP-GAL80^{ts}</i> / UAS- <i>msn</i> -RNAi; <i>TubP-GAL4</i> / +
S1	C	<i>e22c-GAL4</i> , UAS-FLP / +; <i>msn^{P23-5}</i> , <i>FRT80</i> / <i>ubi-eGFP</i> , <i>FRT80</i>
S2	A	<i>y,w</i> , <i>hsFLP</i> / +; <i>Df(2L)flp147E</i> , <i>FRT40A</i> / <i>ubi-GFP</i> , <i>FRT40A</i> (<i>Df</i> removes <i>bsk</i>)
S2	B	<i>w¹¹¹⁸</i> / +; <i>tj-Gal4</i> / + <i>tj-Gal4</i> / UAS- <i>msn</i> -RNAi <i>mys^{XG43}</i> / +; <i>tj-GAL4</i> / UAS- <i>msn</i> -RNAi

Table S2. Conditions for transgene expression

Figure	Panel	Temp at which cross was raised	Temp females on yeast	Time females on yeast
		°C	°C	h
1	C and D	18	29	68–72
3	C and D	18	25	24–29
S1	A and B	18	29	68–72
S2	B	18	25	26.5