

Table S1. **Q/N-rich regions in proteins involved in RNA metabolism and function**

A. Yeast proteins containing Q/N-rich regions with links to RNA metabolism and function				
Function	Locus	Protein	Description	Source
RNA metabolism	YAL021C	CCR4	Cytoplasmic mRNA deadenylase component	Michelitsch and Weissman, 2000
	YNR052C	POP2	Cytoplasmic mRNA deadenylase component	Michelitsch and Weissman, 2000
	YCR093W	CDC39	NOT1, component of cytoplasmic mRNA deadenylase	Michelitsch and Weissman, 2000
	YDL160C	DHH1	mRNA decapping activator/translation repressor	this paper
	YDR228C	PCF11	mRNA 3' end processing factor	Michelitsch and Weissman, 2000
	YEL015W	EDC3	mRNA decapping activator/P-body aggregation factor	This paper
	YERO68W	NOT4	Cytoplasmic mRNA deadenylase component	This paper
	YER112W	LSM4	mRNA decapping and splicing	Michelitsch and Weissman, 2000
	YGL044C	RNA15	mRNA 3' end processing factor	Michelitsch and Weissman, 2000
	Nuclear export	YDL088C	ASM4	Nup59, nuclear pore complex
YGL172W		NUP49	Nuclear pore complex	Michelitsch and Weissman, 2000
YGR119C		NUP57	Nuclear pore complex	Michelitsch and Weissman, 2000
YKL068W		NUP100	Nuclear pore complex	Michelitsch and Weissman, 2000
YMR047C		NUP116	Nuclear pore complex	Michelitsch and Weissman, 2000
Translation	Q0140	VAR1	Mitochondrial small ribosomal protein	Michelitsch and Weissman, 2000
	YDR172W	SUP35	Translation termination factor	Michelitsch and Weissman, 2000
	YGL049C	TIF4632	Translation initiation factor	Michelitsch and Weissman, 2000
RNA-binding	YBL051C	PIN4	Contains RRM ¹	Michelitsch and Weissman, 2000
	YBR212W	NGR1	Contains RRM	Michelitsch and Weissman, 2000
	YDL167C	NRP1	Contains RRM and zinc finger domains, interacts with Pub1p	Michelitsch and Weissman, 2000
	YDR515W	SLF1	Contains La RNA binding domain	Michelitsch and Weissman, 2000
	YJR091C	JSN1	PUF1, contains pumilio/puf RNA binding domain	Michelitsch and Weissman, 2000
	YPR042C	PUF2	Contains pumilio/puf RNA binding domain	Michelitsch and Weissman, 2000
	YLL013C	PUF3	Contains pumilio/puf RNA binding domain	Michelitsch and Weissman, 2000
	YGL014W	PUF4	Contains pumilio/puf RNA binding domain	Michelitsch and Weissman, 2000
	YGL178W	MPT5	PUF5, contains pumilio/puf RNA binding domain	Michelitsch and Weissman, 2000
	YGL122C	NAB2	Contains zinc finger domain	Michelitsch and Weissman, 2000
	YNL016W	PUB1	Contains RRM	Michelitsch and Weissman, 2000
Interact with RNA-associated proteins	YPL190C	NAB3	Contains RRM	Michelitsch and Weissman, 2000
	YBL029W	YBL029w	Interacts with CAF4	Michelitsch and Weissman, 2000
	YBL081W	YBL081W	Interacts with RPS28b and NRP1	Michelitsch and Weissman, 2000

YCL028W	RNQ1	Interacts with CCR4	Michelitsch and Weissman, 2000
YDR505C	PSP1	Interacts with NAB2	Michelitsch and Weissman, 2000
YEL007W	YEL007W	Interacts genetically with CCR4 and POP2	Michelitsch and Weissman, 2000
YGL086W	MAD1	Interacts with nuclear pore complex proteins	Michelitsch and Weissman, 2000
YGL181W	GTS1	Interacts genetically with defects in mrna splicing	Michelitsch and Weissman, 2000
YHR082C	KSP1	May modify DOM34, RNT1, and LSM7	Michelitsch and Weissman, 2000
YJL141C	YAK1	Interacts with DCS1, DCS2, POP2, KEM1, and CAF20	Michelitsch and Weissman, 2000
YML017W	PSP2	Interacts with eif4e and 4G, suppresses defects in Group II splicing	Michelitsch and Weissman, 2000
YMR124W	YMR124W	Interacts with CRM1	Michelitsch and Weissman, 2000
YMR216C	SKY1	SR protein kinase	Michelitsch and Weissman, 2000
YNL027W	CRZ1	May be modified by SKY1 and HRR25	Michelitsch and Weissman, 2000
YNL154C	YCK2	May modify many RNA metabolism proteins	Michelitsch and Weissman, 2000
YNL161W	CBK1	Interacts with NOT3	Michelitsch and Weissman, 2000
YNL243W	SLA2	Interacts with PRP45 and SUP35	Michelitsch and Weissman, 2000
YOR290C	SNF2	Interacts with PUB1, PAB1 and several ribosomal proteins	Michelitsch and Weissman, 2000
YOR329C	SCD5	Interacts with CRM1 and NUP116	Michelitsch and Weissman, 2000
YPL016W	SWI1	Interacts with PUB1, LSM1, and several splicing factors	Michelitsch and Weissman, 2000
YPL204W	HRR25	Interacts with DCP1, DCP2, EDC3, DED1, and several ribosomal proteins	Michelitsch and Weissman, 2000
YPL226W	NEW1	Interacts with TEF1, TEF4, DBP8, and ribosomal proteins	Michelitsch and Weissman, 2000

B. Location of conserved Q/N-rich regions in metazoan P-body proteins

Protein	Species	Accession no.	Residue no.	Length	%Q/N	Source	
Dcp2	<i>H. sapiens</i>	NP_689837.2	276–346	70	29	This paper	
	<i>B. taurus</i>	XP_001254141.1	426–498	73	23	This paper	
	<i>M. musculus</i>	XP_980768.1	361–433	72	28	This paper	
	<i>R. norvegicus</i>	XP_001055777.1	350–410	71	28	This paper	
	<i>X. laevis</i>	AAZ38887.1	277–345	74	30	This paper	
	<i>D. rerio</i>	NP_956446.1	272–335	64	20	This paper	
	<i>D. melanogaster</i>	NP_648805.2	616–764	149	31	This paper	
	<i>S. cerevisiae</i>	NP_014281.1	282–326	45	40	This paper	
	Ge-1/Hedls	<i>H. sapiens</i>	AAH64567.1	1135–1274	140	16	This paper
		<i>M. musculus</i>	BAE27236.1	1140–1179	140	16	This paper
<i>R. norvegicus</i>		NP_001028240.1	1141–1180	140	16	This paper	
<i>G. gallus</i>		XP_414022.2	1153–1292	140	18	This paper	
<i>D. rerio</i>		XP_694565.1	1102–1243	142	20	This paper	
<i>X. laevis</i>		AAH44263.1	1125–1266	142	18	This paper	
<i>D. melanogaster</i>		AAO42643.1	1106–1230	125	10	This paper	
GW182		<i>H. sapiens</i>	NP_055309.2	1264–1553	290	25	This paper
	<i>B. taurus</i>	XP_614640.3	1260–1549	290	25	This paper	
	<i>M. musculus</i>	NP_659174.3	1249–1488	240	26	This paper	
	<i>R. norvegicus</i>	XP_001079613.1	1228–1616	289	25	This paper	
	<i>G. gallus</i>	XP_414871.2	1208–1498	291	25	This paper	
	<i>D. rerio</i>	XP_700599.1	760–1125	366	22	This paper	

TNRC6B	<i>D. melanogaster</i>	NP_001014692.1	628–901	274	30	This paper
	<i>H. sapiens</i>	NP_055903.1	1117–1219	103	37	This paper
	<i>B. taurus</i>	XP_001252364.1	1131–1238	108	41	This paper
	<i>M. musculus</i>	NP_659061.2	1200–1307	108	41	This paper
	<i>R. norvegicus</i>	XP_576311.2	1198–1315	117	45	This paper
	<i>G. gallus</i>	XP_416246.2	1165–1265	102	36	This paper
	<i>D. rerio</i>	XP_001345261.1	1267–1374	108	32	This paper

Michelitsch, M.D., and J.S. Weissman. 2000. A census of glutamine/asparagine-rich regions: implications for their conserved function and the prediction of novel prions. *Proc. Natl. Acad. Sci. USA.* 97:11910–11915.

¹RRM, RNA recognition motif.

Table S2. **Yeast Strains used in this study**

Strains	Genotype	Source
yRP1724	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG DHH1GFP (NEO)</i>	Sheth and Parker, 2003
yRP1726	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG DCP1GFP (NEO)</i>	Sheth and Parker, 2003
yRP1728	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG PAT1GFP (NEO)</i>	Sheth and Parker, 2003
yRP1729	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG LSM1GFP (NEO)</i>	Sheth and Parker, 2003
yRP1730	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG XRN1GFP (NEO)</i>	Sheth and Parker, 2003
yRP2162	<i>MATa leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG Dcp2GFP(NEO)</i>	This paper
yRP2217	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 lys2-201 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO DHH1GFP(NEO)</i>	This paper
yRP2225	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO DCP1GFP(NEO)</i>	This paper
yRP2240	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO PAT1GFP(NEO)</i>	This paper
yRP2232	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 lys2-201 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO LSM1GFP(NEO)</i>	This paper
yRP2248	<i>MATa leu2-3,112 trp1 ura3-52 lys2-201 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO XRN1GFP(NEO)</i>	This paper
yRP2164	<i>MATa leu2-3.112 trp1 ura3-52 lys2-201 cup1::LEU2/PGK1pG/MFA2pG edc3::NEO Dcp2GFP(NEO)</i>	This paper
yRP1736	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 DHH1GFP(NEO)</i>	Sheth and Parker, 2003
yRP1936	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 DCP2GFP(NEO)</i>	Bregues et al., 2005
yRP2246	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 XRN1GFP(NEO)</i>	This paper
yRP2307	<i>MATa leu2-3,112 trp1 ura3-52 lys2-201 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 edc3::NEO DHH1GFP(NEO)</i>	This paper
yRP2308	<i>MATa leu2-3,112 trp1 ura3-52 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 edc3::NEO DCP2GFP (NEO)</i>	This paper
yRP2310	<i>MATa leu2-3,112 trp1 ura3-52 lys2-201 cup1::LEU2/PGK1pG/MFA2pG dcp1::URA3 edc3::NEO XRN1GFP(NEO)</i>	This paper
yRP1738	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 DHH1GFP (NEO)</i>	Sheth and Parker, 2003
yRP1923	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 DCP2GFP (NEO)</i>	Teixeira et al., 2005
yRP2239	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 PAT1GFP (NEO)</i>	This paper
yRP2312	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 lys2-201 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 edc3::NEO DHH1GFP (NEO)</i>	This paper
yRP2314	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 lys2-201 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 edc3::NEO DCP2GFP (NEO)</i>	This paper
yRP2316	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 lys2-201 cup1::LEU2/PGK1pG/MFA2pG xrn1::URA3 edc3::NEO PAT1GFP (NEO)</i>	This paper
yRP2325	<i>MATa leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG Dcp2GFP(NEO) petite</i>	This paper
yRP2327	<i>MATa leu2-3.112 trp1 ura3-52 lys2-201 cup1::LEU2/PGK1pG/MFA2pG Dcp2GFP(NEO) edc3:: NEO petite</i>	This paper
yRP2333	<i>MAT leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG Dhh1GFP(NEO) petite</i>	This paper
yRP2335	<i>MATa leu2-3.112 trp1 ura3-52 his4-539 lys2 cup1::LEU2/PGK1pG/MFA2pG Dhh1GFP(NEO) edc3:: NEO petite</i>	This paper
yRP2093	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2]</i>	This paper
yRP2362	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2EDC3]</i>	This paper
yRP2361	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2DCP1]</i>	This paper

yRP2359	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2DCP2(1-300)]</i>	This paper
yRP2360	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2DCP2(102-300)]</i>	This paper
yRP2363	<i>MAT trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOBD-2DHH1(250-461)]</i>	This paper
yRP2364	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAD]</i>	This paper
yRP2368	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAEDDC3]</i>	This paper
yRP2369	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAEDDC3(1-231)]</i>	This paper
yRP2370	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAEDDC3(1-85)]</i>	This paper
yRP2371	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAEDDC3(86-231)]</i>	This paper
yRP2372	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4 gal80 LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ [pOAEDDC3(232-551)]</i>	This paper
yRP2339	<i>MATa leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG lsm4 C:: NEO Dcp2GFP(NEO)</i>	This paper
yRP2340	<i>MAT leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG lsm4 C:: NEO edc3:: NEO Dcp2GFP(NEO)</i>	This paper
yRP840	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG [pRP1193] [pRP1187]</i>	Hatfield et al., 1996
yRP2338	<i>MATa leu2-3.112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG lsm4 C:: NEO edc3:: NEO [pRP1193] [pRP1187]</i>	This paper
yRP1516	<i>MATa his4-539 leu2-3,112 lys2-201 trp1 ura3-52 cup1::LEU2/PGK1pG/MFA2pG dcp2-7::URA3</i>	Dunckley, 2001
yRP1748	<i>MATa his4-539 trp1 leu2-3,112 ura3-52 edc3::NEO dcp2-7::URA3 cup1::LEU2/PGK1pG/MFA2pG</i>	Kshirsagar and Parker, 2004
yRP1502	<i>MATa his4-539 trp1 leu2-3,112 ura3-52 lys2-201 dcp2-7::URA3 ski3::TRP1</i>	Dunckley and Parker, 1999
yRP1750	<i>MAT his4-539 leu2-3,112 ura3-52 lys2-201 edc3::NEO dcp2-7::URA3 ski3::TRP1</i>	Kshirsagar and Parker, 2004
yRP2430	<i>MATa leu2 trp1 ura3 lys2 his3 cup1::LEU2/PGK1pG/MFA2pG Dcp2GFP(NEO) edc3::NEO lsm4::NEO [pRP1551]</i>	This paper

Bregues, M., Teixeira, D., and R. Parker. 2005. Movement of eukaryotic mRNAs between polysomes and cytoplasmic processing bodies. *Science*. 310:486–489.

Dunckley, T., and R. Parker. 1999. The DCP2 protein is required for mRNA decapping in *Saccharomyces cerevisiae* and contains a functional MutT motif. *EMBO J.* 18:5411–5422.

Dunckley, T., Tucker, M., and R. Parker. 2001. Two related proteins, Edc1p and Edc2p, stimulate mRNA decapping in *Saccharomyces cerevisiae*. *Genetics*. 157:27–37.

Hatfield, L., Beelman, C.A., Stevens, A., and R. Parker. 1996. Mutations in trans-acting factors affecting mRNA decapping in *Saccharomyces cerevisiae*. *Mol. Cell Biol.* 16:5830–5838.

Kshirsagar, M., and R. Parker. 2004. Identification of Edc3p as an enhancer of mRNA decapping in *Saccharomyces cerevisiae*. *Genetics*. 166:729–739.

Sheth, U., and R. Parker. 2003. Decapping and decay of messenger RNA occur in cytoplasmic processing bodies. *Science*. 300:753–755.

Teixeira, D., Sheth, U., Valencia-Sanchez, M.A., Bregues, M., and R. Parker. 2005. Processing bodies require RNA for assembly and contain nontranslating mRNAs. *RNA*. 11:371–382.

Table S3. Plasmids and oligonucleotides used in this study

Number	Name/Sequence	Description	Reference
pRP1433	FlagEdc3	To express N-terminal Flag-tagged Edc3 in yeast from TRP1 CEN plasmid	This study
pRP1434	Flag LsmEdc3	To express N-terminal Flag-tagged Edc3 with amino acids 1–85 deleted in yeast from TRP1 CEN plasmid	This study
pRP1435	Flag FDFEdc3	To express N-terminal Flag-tagged Edc3 with amino acids 86–231 deleted in yeast from TRP1 CEN plasmid	This study
pRP1436	Flag Yjef-NEdc3	To express N-terminal Flag-tagged Edc3 with amino acids 232–551 deleted in yeast from TRP1 CEN plasmid	This study
pRP1441	pRS200 HisLsm	TRP1 CEN plasmid prs314 with bglii linkers inserted in SmaI site To express N-terminal His-tagged Edc3 amino acids 1–85 in <i>E. Coli</i>	Sikorski and Hieter, 1989 This study
pRP1442	HisFDF	To express N-terminal His-tagged Edc3 amino acids 86–231 in <i>E. Coli</i>	This study
pRP1210	FlagDcp2(1–300)His	Plasmid to express amino acids 1–300 of Dcp2, which is Flag-tagged at the N terminus and His-tagged at the C terminus in <i>E. Coli</i>	She et al., 2006
pRP1211	FlagDcp2(102–300)His	Plasmid to express amino acids 102–300 of Dcp2, which is Flag-tagged at the N terminus and His-tagged at the C terminus in <i>E. Coli</i>	She et al., 2006
pRP1317	GSTDhh1(46–461)	Plasmid to express amino acids 46–461 of Dhh1, which is Gst tagged at the N terminus in <i>E. Coli</i>	Gift from H. Song
pRP1438	GSTDhh1(46–249)	Plasmid to express amino acids 46–249 of Dhh1, which is Gst tagged at the N terminus in <i>E. Coli</i>	This study
pRP1439	GSTDhh1(250–461)	Plasmid to express amino acids 250–461 of Dhh1, which is Gst tagged at the N terminus in <i>E. Coli</i>	This study
pRP1187	pOAD pOBD-2 U1A-GFP	Activation domain plasmid for two-hybrid interaction analysis Binding domain plasmid for two-hybrid interaction analysis To express GFP tagged U1A RNA binding protein in yeast	Cagney et al., 2000 Cagney et al., 2000 Bregues et al., 2005
pRP1193	MFA2P-U1A	To localize MFA2 mrna in yeast using U1A-GFP	Bregues et al., 2005
pRP1551	Lsm4RNQ1	To express Lsm4 with Q/N domain replaced with RNQ1 prion domain	This study
oRP1321	CCGCCGCATATGTCACAATTTGTTGGTTTCGG	To create prp1441 hisedc3lsm	This study
oRP1322	CCGCCGCTCGAGTTAGCGATTTTGATTATAATCGTTTTG	To create prp1441	This study
oRP1323	CCGCCGCATATGGGTGAACATATTGATTGGCAAGATGATG	To create prp1442 hisedc3fdf	This study
oRP1324	CCGCCGCTCGAGTTAATTTATGGATGGAGAGTTAGTTG CAGAGTG	To create prp1442	This study
oRP1325	GATTACAGGGACATAACAG	To sequence Edc3	This study
oRP1326	CCTATGGCTACACCAGTAC	To sequence Edc3	This study
oRP1327	GAAATAGTCATCGATGCC	To sequence Edc3	This study
oRP1328	CATCTGCATAAGCCTTATGAAATTAATTAATGGAAGA ATTGACTCTAAAGGG	To create prp1438 with a stop codon at position 249 in Dhh1p	This study
oRP1329	CCCTTTAGAGTCAATTCTCCATTTAATTAATTCATAA GGCTTATGCAGATG	To create prp1438	This study
oRP1330	GCGCTGGGATCCATGGAAGAATTGACTCTAAAGGG	To create prp1439 with an N-terminal deletion of Dhh1 up to position 249	This study
oRP1331	CCGGGAGCTGCATGTGTCAGAGG	To create prp1439	This study
oRP1332	ATGATTACGCCAAGCTCGAAATTAACCCTCACTAAAGG GAACAAAAGCTGGAGCTCGCAC ACTTCGAAATGGCCTTCTTTG	To amplify nucleotides –403–1,987 of Edc3 locus to create prp1433-36	This study

oRP1333	GTAAAACGACGGCCAGTGAATTGTAATACGACTCACTA TAGGGCGAATTGGGTACCGATC ACAATGACAGCACTATTCTGCC	To amplify nucleotides -403-1,987 of Edc3 locus to create prp1433-36	This study
oRP1334	CAGTAATTCGTAAAAGAAACCATAATGGACTACAAGG ACGACGATGACAAGATGTC ACAATTTGTTGGTTTCGGA	To insert a Flag tag at the N terminus of Edc3 to create prp1433	This study
oRP1335	TCCGAAACCAACAAATTGTGACATCTTGTCATCGTCGT CCTTGATGTCATTATGGTTTC TTTTACGAATTACTG	To insert a Flag tag at the N terminus of Edc3 to create prp1433	This study
oRP1336	ATGGACTACAAGGACGACGATGACAAGGGTGAACATA TTGATTGGCAAGATGATG	To delete amino acids 1-85 in Edc3 to create prp1434	This study
oRP1337	CATCATCTTGCCAATCAATATGTTCCACCTTGTCATCGT CGTCCCTGTAGTCCAT	To delete amino acids 1-85 in Edc3 to create prp1434	This study
oRP1338	GCAACAAAACGATTATAATCAAAAATCGCGATAAAACG AAAGGTACAGTTATAAATG	To delete amino acids 86-231 in Edc3 to create prp1435	This study
oRP1339	CATTATAACTGTACCTTTCGTTTTATCGCGATTTTGAT TATAATCGTTTTGTGTC	To delete amino acids 86-231 in Edc3 to create prp1435	This study
oRP1340	CACTCTGCAACTAACTCTCCATCCATAAATTAACCAA GAATTACTTTAGCC	To delete amino acids 232-551 in Edc3 to create prp1436	This study
oRP1341	GGCTAAAGTAATCTTGGTTTATTTATGGATGGAGAGT TAGTTGCAGATG	To delete amino acids 232-551 in Edc3 to create prp1436	This study
oRP1342	CTATCTATTCGATGATGAAGATACCCACCAAACCCAA AAAAAGAGATCGAATTCCAGCT GACCACCATG	To clone two-hybrid plasmids by homologous recombination	This study
oRP1343	CTTGCGGGTTTTTCAGTATCTACGATTCATAGATCTCT GCAGGTGACGGATCCCCGGG AATTGCCATG	To clone two-hybrid plasmids by homologous recombination	This study
oRP1344	AATTCCAGCTGACCACCATGTCACTGCCGCTACGACAC G	To amplify Dcp2(1-300) for two-hybrid	This study
oRP1345	AATTCCAGCTGACCACCATGAGTATACCGGTGAGGGGC GC	To amplify Dcp2(102-300) for two-hybrid	This study
oRP1346	GATCCCCGGGAATTGCCATGTTACTCTTGGCTCGAGGG TACCTG	To amplify Dcp2(102-300) for two-hybrid	This study
oRP1347	AATTCCAGCTGACCACCATGGAAGAATTGACTCTAAAG GG	To amplify Dhh1(250-461) for two-hybrid	This study
oRP1348	GATCCCCGGGAATTGCCATGTTAATGATGTTGCTGCGG AGG	To amplify Dhh1(250-461) for two-hybrid	This study
oRP1349	AATTCCAGCTGACCACCATGTCACAATTTGTTGGTTTCG G	To amplify Edc3 for two-hybrid	This study
oRP1350	GATCCCCGGGAATTGCCATGTTACAAATCTAATAGCAG GGACCC	To amplify Edc3 for two-hybrid	This study
oRP1351	AATTCCAGCTGACCACCATGACCGGAGCAGCAACTGC	To amplify Dcp1 for two-hybrid	This study
oRP1352	GATCCCCGGGAATTGCCATGTCAAGCAAAAGAATCTTT TGGCTC	To amplify Dcp1 for two-hybrid	This study
oRP1353	GATCCCCGGGAATTGCCATGTTAATTTATGGATGGAGA GTTAGTTGCAGAGTG	To amplify the lsmfdf region of Edc3 for two- hybrid	This study
oRP1354	GATCCCCGGGAATTGCCATGTTAGCGATTTTGATTATA ATCGTTTTG	To amplify the Lsm region of Edc3 for two- hybrid	This study
oRP1355	AATTCCAGCTGACCACCATGGGTGAACATATTGATTGG CAAGATGATG	To amplify the FDF region of Edc3 for two- hybrid	This study
oRP1356	AATTCCAGCTGACCACCATGGATAAAACGAAAGGTAC AGTTATAAATG	To amplify the Yjef-N region of Edc3 for two-hybrid	This study
oRP1357	GTTTATCAAATTGCAAGATAATATAATTGACAAGGTCA AGTGAGGGGCGCCACTTCTAAA	To delete the C terminus of Lsm4p in yeast	This study
oRP1358	TATTTATATATGTACATAATTATTATAATACAAAATTTG TGAATTCGAGCTCGTTTAAAC	To delete the C terminus of Lsm4p in yeast	This study
oRP1359	GCGCGCGTAATACGACTCACTATAGGGCGAATTGGGTA CCGGGCCCCCCCTCGAGCAGAGTAATTGCTTCGTTCTG TATCTACTTG	To amplify Lsm4 to insert into plasmids	This study
oRP1360	GATTACGCCAAGCGCGCAATTAACCCTCACTAAAGGGA ACAAAAGCTGGAGCTCGCAATACGTTTACAAATCTCTC TGGTTGG	To amplify Lsm4 to insert into plasmids	This study
oRP1361	CTTTTATCAAGTTTATCAAATTGCAAGATAATATAATTG ACAAGGTCAAGGGATCCCTAAACAAATTTTGTATTATAA	To delete the C terminus of Lsm4	This study

oRP1362	TAATTATGTACATATATAAATATATTGG CCAATATATTTATATATGTACATAATTATTATAATACAA AATTGTTTAGGATCCCTTGACCTTGTCAATTATATTAT CTTGCAATTTGATAAACTTGATAAAAAG	To delete the C terminus of Lsm4	This study
oRP1365	CTTTTATCAAGTTTATCAAATTGCAAGATAATATAATTG ACAAGGTCAAGCAAGGTCAGGGACAAGTCAAGG	To insert the prion domain of Rnq1p in Lsm4 C	This study
oRP1366	CCAATATATTTATATATGTACATAATTATTATAATACAA AATTTGTTTAGTAGCGGTTCTGGTTGCCGTTATTG	To insert the prion domain of Rnq1p in Lsm4 C	This study
oRP140	ATATTGATTAGATCAGGAATTCC	Probe for MFA2pG mrna	Caponigro and Parker, 1995
oRP100	GTCTAGCCGCGAGGAAGG	Probe for 7S RNA used as a Loading control	Caponigro et al., 1993

Bregues, M., D. Teixeira, and R. Parker. 2005. Movement of eukaryotic mRNAs between polysomes and cytoplasmic processing bodies. *Science*. 310:486–489.

Cagney, G., P. Uetz, and S. Fields. 2000. High-throughput screening for protein-protein interactions using two-hybrid assay. *Methods Enzymol.* 328:3–14.

Caponigro, G., and R. Parker. 1995. Multiple functions for the poly(A)-binding protein in mRNA decapping and deadenylation in yeast. *Genes Dev.* 9:2421–2432.

Caponigro, G., D. Muhlrads, and R. Parker. 1993. A small segment of the MAT alpha 1 transcript promotes mRNA decay in *Saccharomyces cerevisiae*: a stimulatory role for rare codons. *Mol. Cell Biol.* 13:5141–5148.

She, M., Decker, C.J., Chen, N., Tumati, S., Parker, R., and H. Song. 2006. Crystal structure and functional analysis of Dcp2p from *Schizosaccharomyces pombe*. *Nat. Struc. Mol. Biol.* 13:63–70.

Sikorski, R.S., and P. Hieter. 1989. A system of shuttle vectors and yeast host strains designed for efficient manipulation of DNA in *Saccharomyces cerevisiae*. *Genetics.* 122:19–27.